

# On HIV in the elderly and vitamin B metabolism in HIV infection

Erika Tyrberg

Department of Infectious diseases  
Institute of Biomedicine  
Sahlgrenska Academy, University of Gothenburg



UNIVERSITY OF GOTHENBURG

Gothenburg 2021

Cover illustration: Artwork by Jennie Tyrberg

On HIV in the elderly and vitamin B metabolism in HIV infection

© Erika Tyrberg 2021

[erika.tyrberg@gu.se](mailto:erika.tyrberg@gu.se)

ISBN 978-91-8009-602-7 (PRINT)

ISBN 978-91-8009-603-4 (PDF)

<http://hdl.handle.net/2077/70033>

Printed in Borås, Sweden 2022

Printed by Stema Specialtryck AB



“Vi måste skydda de nya från att komma in i det här gänget,  
och så måste vi ta hand om de gamla.”

Torbjörn

*Ur *Leva Livet – Att åldras med hiv**



# ABSTRACT

The evolution of the human deficiency virus (HIV) field is unparalleled in the history of infectious diseases. From the first cases in the beginning of the 1980s, when an HIV diagnosis was a death sentence, through the discovery of the first effective medicines, up till today when people living with HIV (PLHIV) with access to antiretroviral therapy (ART) can lead a near normal life. The aim of this thesis was to investigate further into two areas where knowledge is still lacking, and important questions remain. We investigated HIV in the elderly and the role of vitamin B metabolism in HIV-associated central nervous system (CNS) disease.

In paper I and II we studied HIV infection in the elderly ( $\geq 65$  years of age) compared to a control group ( $\leq 49$  years of age). In a study of cross-sectional design 100 elderly PLHIV and 99 controls, on ART regimens containing atazanavir, darunavir, or efavirenz were included. In paper I we showed that elderly had a higher number of concomitant medications, comorbidities, and potential drug-drug interactions, than the younger controls. In the darunavir arm, the elderly had higher steady-state concentrations. This was also found in the atazanavir arm, although not statistically different, but suggesting a possible class effect of protease inhibitors. Paper II investigated the role of ART regimen on markers of inflammation and immune activation in elderly PLHIV. The regimens had different inflammatory profiles with lower interleukin-6 levels in the atazanavir arm, and lower ICAM-1 in the efavirenz arm. The darunavir arm had higher CXCL10 levels compared to the efavirenz arm.

Paper III and IV studied the role of homocysteine and vitamin B metabolism in CNS injury in HIV infection. Paper III describes an association between plasma homocysteine, a marker of vitamin B<sub>12</sub> and folate deficiency, and cerebrospinal fluid neurofilament light protein (NfL), a sensitive marker of neuroaxonal damage in HIV infection. In paper IV this association was further studied in a randomised controlled clinical trial. Sixty-one virally suppressed PLHIV were randomised either to the active treatment arm (treatment with vitamin B<sub>12</sub>, B<sub>6</sub>, and folate) or control arm. After 12 months the levels of homocysteine had decreased, and the plasma B<sub>12</sub> and folate levels had increased in individuals in the treatment arm. However, no difference in plasma levels of NfL was found compared to the control arm at 12 months. Furthermore, in the treatment arm, no difference in NfL was found after 24 months, compared to baseline plasma NfL levels.

In conclusion, we found that elderly PLHIV are at risk of adverse drug events through a high prevalence of concomitant medications, potential drug-drug interactions, and higher drug concentrations of protease inhibitors. In addition, we found different inflammatory profiles of efavirenz, atazanavir, and darunavir, a finding that needs to be confirmed in future studies. Furthermore, a novel finding of an association between homocysteine and NfL was made. However, supplementation with B vitamins did not decrease NfL, suggesting a non-vitamin B dependent cause of the association.

**Keywords:** HIV-1, elderly, drug levels, potential drug-drug interactions, inflammation, immune activation, antiretroviral therapy, homocysteine, neurofilament light protein.

ISBN 978-91-8009-602-7 (PRINT)

ISBN 978-91-8009-603-4 (PDF)

# SAMMANFATTNING PÅ SVENSKA

I juni 1981 rapporterades det om de första fallen av vad som skulle bli känt som förvärvat immunbristsyndrom, AIDS. De första fallen diagnosticerades i USA men snart förstod man att det rörde sig om en global pandemi. 1983 lyckades man för första gången med att isolera viruset som orsakar AIDS, humant immunbristvirus (HIV). I slutet av 1980-talet utvecklades de första medicinerna och sedan mitten av 90-talet har det funnits effektiva mediciner. Idag kan en person med HIV, som står på behandling, leva ett normalt liv. Detta har lett till nya frågeställningar kring långtidseffekter av behandling och åldersrelaterade sjukdomar. Denna avhandling fokuserar på två områden där det saknas viktig kunskap, HIV hos de äldre och B-vitamin metabolismens relation till nervskada hos de som lever med HIV.

I takt med att fler och fler fått tillgång till effektiva mediciner mot HIV så har de som lever med HIV blivit äldre. I Sverige är runt hälften över 50 år gamla och allt fler blir äldre än 65 år. Med åldern ökar risken för att utveckla åldersrelaterade sjukdomar och sannolikheten för att ta flera olika läkemedel. Kroppen förändras vilket gör att man kan vara känsligare för läkemedel. Det saknas viktig kunskap om vad detta innebär för de som lever med HIV. För att undersöka detta genomförde vi den tvärsnittsstudie som är grunden för arbete I och II. I arbete I jämförde vi en grupp personer som lever med HIV som var 65 år eller äldre med en grupp som var 49 år eller yngre. Som förväntat hade de äldre fler sjukdomar och tog fler läkemedel än de yngre. Vi fann att koncentrationerna i blodet av en grupp HIV-läkemedel, proteashämmare, var högre hos de äldre. Hos personer som tog ett av dessa läkemedel, darunavir, var biverkningar vanligare. Vidare upptäckte vi att de äldre oftare hade en kombination av läkemedel som kan påverka varandra på ett ofördelaktigt sätt och att det var vanligt med polyfarmaci, dvs. att man använder fler än fem läkemedel varje dag.

Jämfört med sina jämnåriga har personer som lever med HIV en ökad risk att utveckla åldersrelaterade sjukdomar. Forskning talar för att en anledning till detta kan vara att de som lever med HIV har en högre nivå av inflammation i kroppen trots välfungerande behandling än de som inte har HIV. För att undersöka om valet av läkemedel påverkar denna inflammation undersökte vi i delarbete II 10 olika inflammationsmarkörer hos de äldre baserat på läkemedelsregim. Vi fann skillnader för tre olika markörer. De som behandlades med atazanavir hade lägre IL-6, de som behandlades med efavirenz hade lägre ICAM-1 och de som behandlades med darunavir hade högre CXCL10.

Utan antiretroviral behandling skulle många som lever med HIV utveckla en särskild form av demens på grund av infektionen. Även innan symptom utvecklas kan man se tecken på nervskada i hjärnan som ökar i takt med att sjukdomen förvärras. Antiretroviral behandling hindrar utvecklingen av demens och nervskadan minskar. Vissa personer har trots det kvarvarande tecken på en lågradig pågående nervskada, som kan mätas med biomarkören neurofilament light protein (NfL). Orsaken till denna kvarvarande skada är inte helt klarlagd men man kan också se tecken på immunaktivering hos personer med välbehandlad HIV. För att utreda denna kvarvarande skada mätte vi i delarbete III nivåer av NfL i ryggmärgsvätska och nivåer av homocystein i blod. Homocysteinnivåer har tidigare kopplats till demens och kognitiv påverkan hos HIV-negativa personer. Vi fann ett samband där högre NfL-nivåer var kopplat till högre nivåer av homocystein. Detta utredde vi vidare i delarbete IV. Homocysteinnivåerna i blodet är beroende av B-vitaminerna B<sub>12</sub>, B<sub>6</sub> och folsyra och homocysteinnivåer sjunker vid behandling med B-vitaminer. I delarbete IV genomförde vi en randomiserad kontrollerad studie av personer som lever med HIV där vi gav B-vitaminer till den ena gruppen medan den andra gruppen inte fick någon behandling med B-vitaminer. Vi mätte NfL-nivåer och homocysteinnivåer innan och under behandlingen och såg att homocysteinnivåerna sjönk hos de som fick behandling men inte NfL-nivåerna. Detta talar emot att låga B-vitamnivåer är en bidragande orsak till kvarvarande nervskada hos personer som lever med HIV med välfungerande behandling.

Sammanfattningsvis har denna avhandling ökat kunskapen kring HIV och läkemedelsbehandling hos äldre personer med HIV. Vi har visat att denna grupp är i risk för ogynnsamma effekter av läkemedel, genom ökade läkemedelskoncentrationer av vissa HIV-läkemedel, risk för interaktioner mellan läkemedel och polyfarmaci. Vi har också visat att val av antiretroviralbehandling kan ha betydelse för inflammationsnivåer. Utöver det har vi funnit ett samband mellan tecken på nervskada och homocysteinnivåer hos personer med HIV med behandling, men kunde inte se någon effekt av behandling med B-vitaminer vilket talar för att sambandet beror på en annan orsak.



# LIST OF PAPERS

This thesis is based on the following studies, referred to in the text by their Roman numerals.

- I. Tyrberg E, Edén A, Eriksen J, Nilsson S, Treutiger CJ, Thalme A, Mellgren Å, Gisslén M, Andersson LM.  
**Higher plasma drug levels in elderly people living with HIV treated with darunavir**  
PLoS ONE 2021; 16(2): e0246171
- II. Tyrberg E, Skovbjerg S, Samuelsson E, Nilsson S, Edén A, Treutiger CJ, Thalme A, Mellgren Å, Gisslén M, Andersson LM  
**Markers of inflammation and immune activation in elderly HIV-1 infected individuals on stable ART treatment with efavirenz, darunavir, or atazanavir**  
In manuscript
- III. Ahlgren E, Hagberg L, Fuchs D, Andersson LM, Nilsson S, Zetterberg H, Gisslén M  
**Association between Plasma Homocysteine Levels and Neuronal Injury in HIV infection**  
PLoS ONE 2016; 11(7): e0158973
- IV. Tyrberg E, Hagberg L, Andersson LM, Nilsson S, Yilmaz A, Mellgren Å, Blennow K, Zetterberg H, Gisslén M  
**The effect of vitamin B supplementation on neuronal injury in PLHIV – a randomised controlled trial**  
Submitted manuscript

Reprints in this thesis are made with permission from the publishers

# CONTENT

ABBREVIATIONS .....	12
1 INTRODUCTION.....	15
1.1 The pandemic .....	15
1.2 The human immunodeficiency virus .....	16
1.2.1 Life cycle .....	18
1.2.2 Tropism .....	19
1.3 From HIV to AIDS.....	20
1.4 Antiretroviral therapy – the paradigm shift.....	22
1.4.1 Treatment as prevention .....	25
1.5 Latency and reservoirs.....	26
1.6 HIV today .....	26
1.7 HIV and aging .....	28
1.7.1 Comorbidities .....	28
1.7.2 ART and non-ART drugs.....	28
1.7.3 Inflammation and immune activation.....	29
1.8 HIV in the central nervous system.....	30
1.8.1 Neuropathogenesis – a trojan horse?.....	31
1.8.2 Biomarkers of CNS infection.....	33
1.8.3 Neurofilament light protein .....	33
1.8.4 Neopterin .....	34
1.9 B vitamins & homocysteine .....	34
1.9.1 Vitamin B <sub>12</sub> .....	37
1.9.2 Folate .....	38
1.9.3 Homocysteine and neurocognitive disease .....	39
2 AIMS .....	43
3 STUDY POPULATION AND DESIGN .....	45
3.1 Paper I & II .....	45
3.2 Paper III .....	47

3.3 Paper IV .....	48
4 METHODS .....	51
4.1 Laboratory assays .....	51
4.1.1 Drug concentrations .....	51
4.1.2 Markers of inflammation .....	51
4.1.3 Neurofilament light protein .....	51
4.1.4 Homocysteine and B vitamins .....	52
4.2 Drug-drug interactions.....	52
4.3 Neurocognitive testing .....	52
4.4 Statistical methods.....	53
4.5 Ethics.....	53
5 HIV IN THE ELDERLY .....	55
6 VITAMIN B METABOLISM IN HIV INFECTION .....	61
7 CONCLUSION .....	65
8 FUTURE PERSPECTIVES.....	67
9 ACKNOWLEDGEMENTS.....	71
10 REFERENCES .....	75

# ABBREVIATIONS

5-methyl-THF	5-methyl-tetrahydrofolate
AIDS	Acquired immunodeficiency syndrome
ANI	Asymptomatic neurocognitive impairment
ART	Antiretroviral therapy
ATV	Atazanavir
CCR5	CC chemokine receptor 5
CD	Cluster of differentiation
CDC	Centers for Disease Control and Prevention
CNS	Central nervous system
CSF	Cerebrospinal fluid
CXCL10	C-X-C motif chemokine ligand 10
CXCR4	CXC chemokine receptor 4
DRV	Darunavir
EI	Entry inhibitor
EFV	Efavirenz
HAD	HIV-associated dementia
HAND	HIV-associated neurocognitive disorder
HIV	Human immunodeficiency virus
ICAM-1	Intercellular adhesion molecule-1
IL-6	Interleukin-6

INSTI	Integrase strand transfer inhibitor
IF	Intrinsic factor
MMA	Methylmalonic acid
MND	Mild neurocognitive disorder
NfL	Neurofilament light protein
NNRTI	Non-nucleoside reverse transcriptase inhibitor
NRTI	Nucleoside reverse transcriptase inhibitor
P	Plasma
PDDI	Potential drug-drug interaction
PI	Protease inhibitor
PLHIV	People living with HIV
s	Soluble
SAH	S-adenosylhomocysteine
SAM	S-adenosylmethionine
SIV	Simian immunodeficiency virus
SMART	Strategies for Management of AntiRetroviral Therapy
START	Strategic Timing of AntiRetroviral Treatment
STOPP	Screening tool of older people's prescriptions

## Introduction

# 1 INTRODUCTION

When looking back on the last 40 years the picture drawn is remarkable. The evolution of the human immunodeficiency virus (HIV) field is unparalleled in the history of medicine. From the first cases in the beginning of the 1980s, when an HIV diagnosis was a death sentence, through the discovery of the first effective medicines, up till today when people living with HIV (PLHIV) with access to antiretroviral therapy (ART) can lead a near normal life.

## 1.1 THE PANDEMIC

The first notion of the emerging epidemic was when the Morbidity and Mortality Weekly Report on Friday 5<sup>th</sup> of June 1981 published a report of five young men in Los Angeles presenting with pneumocystis pneumonia, a kind of pneumonia associated with immunodeficiency.<sup>1</sup>

1981 June 5;30:250-2

### ***Pneumocystis* Pneumonia – Los Angeles**

**In the period October 1980-May 1981, 5 young men, all active homosexuals, were treated for biopsy-confirmed *Pneumocystis carinii* pneumonia at 3 different hospitals in Los Angeles, California. Two of the patients died. All 5 patients had laboratory-confirmed previous or current cytomegalovirus (CMV) infection and candidal mucosal infection. Case reports of these patients follow.**

*Figure 1. Extract of the first published report on AIDS.<sup>1</sup>*

A month later, in July, it was reported that 26 men living in New York and California were diagnosed with diseases related to immunosuppression, such as pneumocystis pneumonia and Kaposi sarcoma.<sup>2</sup> This was soon followed by several additional reports describing a similar clinical syndrome of low CD4<sup>+</sup> cell counts and diseases related to immunodeficiency. In September 1982 the Centers for Disease Control and Prevention (CDC) in the United States named the disease acquired immunodeficiency syndrome, AIDS.<sup>3</sup>

Initially, it was not known that AIDS was caused by a virus. The recent finding that retroviruses could cause infection in humans<sup>4</sup> led the way to the discovery of HIV-1 in 1983, by French scientists Françoise Barré-Sinoussi and Luc Montagnier, who were later awarded the Nobel prize in 2008.<sup>5</sup> During the next

year Robert Gallo and his group showed that HIV was the causative agent of AIDS which was confirmed by others.<sup>6-8</sup> The groups all named the virus differently, and it was not until 1986 that the newly discovered virus was officially named the human immunodeficiency virus-1 (HIV-1).<sup>9</sup>

The first American reports described cases of men who have sex with men, but soon cases of AIDS were also reported in individuals with haemophilia (receiving blood products), intravenous drug users, and heterosexual individuals.<sup>3</sup> Parallel to the unrevealing of the epidemic in the US, a report came from Belgium of immigrated men from Africa who presented with AIDS.<sup>10</sup> It was soon evident that AIDS affected several countries in Africa.<sup>11-14</sup> The reports from Africa portrayed a disease equally affecting women.<sup>11, 12</sup> In 1983 the first publications on vertical transmission, from mother to child, were published.<sup>15</sup>

The search for the origin of the pandemic led researchers to the African continent. A group of viruses, simian immunodeficiency viruses (SIVs), genetically related to HIV-1 was found among nonhuman primates living in sub-Saharan Africa.<sup>16, 17</sup> The virus with the closest resemblance to HIV-1 was found among chimpanzee.<sup>17</sup> It is believed that cross-species transmission from chimpanzee to humans have occurred at least four times, giving rise to the four known groups of HIV-1 (M, N, O, and P). M and N strains are known to originate from SIV infecting chimpanzee of the subspecies *Pan troglodytes troglodytes*.<sup>18</sup> The specific origin of O and P strains is not established.<sup>19</sup> How HIV-1 was first transmitted to humans is not known, but it is proposed to be through consumption and handling of bushmeat.<sup>19, 20</sup> Based on studies of the evolution of HIV researchers have localised the cradle of the pandemic to Kinshasa (Leopoldville at the time),<sup>19</sup> and the oldest diagnosed case derives from retrospective analysis of a plasma sample from 1959.<sup>21</sup> It is believed that the initial transmission occurred at the beginning of the 20<sup>th</sup> century.<sup>19, 22</sup>

Some years after the discovery of HIV-1, in 1986, another virus capable of causing AIDS in humans was found. The virus was named HIV-2,<sup>23</sup> and it originated from a SIV strain infecting sooty mangabey.<sup>20, 24</sup> Compared to HIV-1, HIV-2 is less pathogenic and less transmissible. It constitutes a smaller epidemic primarily localised to West Africa.<sup>19, 23</sup> This thesis will only cover HIV-1 (hereafter called HIV).

## 1.2 THE HUMAN IMMUNODEFICIENCY VIRUS

HIV belongs to the family of retroviridae, and the genus lentivirus.<sup>25</sup> Retroviruses are unique in that they contain the enzyme reverse transcriptase that translates RNA to DNA,<sup>26</sup> in contrast to the human transcription enzymes that translate

DNA to RNA. The lentiviruses are characterised by the slow disease progression that they give rise to.<sup>25, 26</sup>

The HIV virion is approximately 100 nm in diameter.<sup>25, 27</sup> It is made up of a lipid membrane envelope carrying the glycoproteins gp120 and gp41. Inside the envelope is a cone-shaped capsid containing the viral genome, consisting of two identical single stranded RNA molecules, the important viral enzymes (reverse transcriptase, integrase, protease), and accessory proteins.<sup>25, 27</sup> The genome includes the three major genes *gag*, *pol*, and *env* that in turn are responsible for encoding the structural proteins, viral enzymes, and envelope glycoproteins. In addition, the genome includes genes encoding the different regulatory and accessory proteins (Tat, Nef, Rev, Vif, Vpu and Vpr) important for e.g. viral replication and intracellular transport.<sup>25, 27</sup>

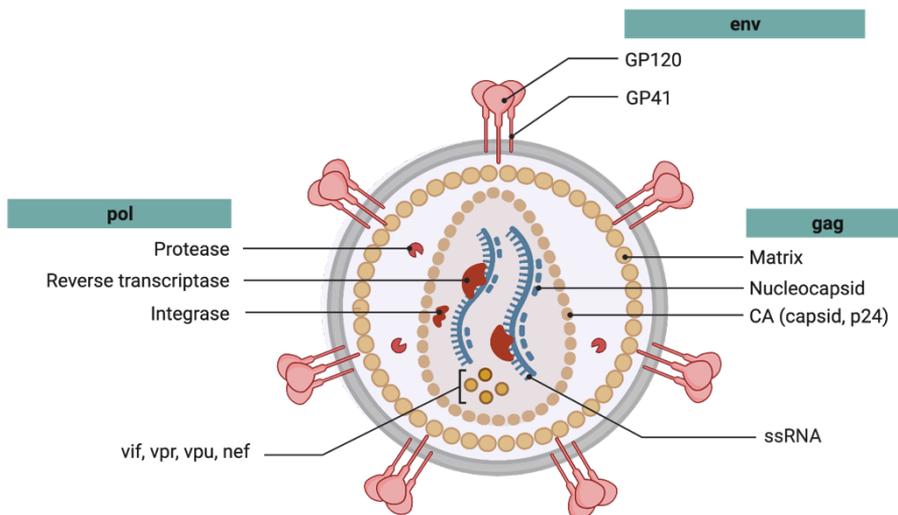


Figure 2. The human immunodeficiency virus.<sup>28</sup>

HIV is divided into four groups, based on genetic differences and origin, M (major), O (outlier), N (non-m/non-O), and P.<sup>29</sup> Group M constitutes the virus responsible for the pandemic, whereas N, O, and P are found primarily in western Africa. The M group is divided in turn into nine subgroups A–D, F–H, J, K.<sup>19</sup> In addition, recombinant forms of different subtypes of HIV exist.<sup>30, 31</sup> The different subtypes are unevenly spread globally where subgroup B predominate in Europe and North America, C in India, whereas a diverse panorama of types is present in Africa.<sup>29</sup>

### 1.2.1 LIFE CYCLE

HIV target cells that present the CD4 receptor (CD4<sup>+</sup> T cells) on their cell surface. These cells include T lymphocytes, monocytes, macrophages, microglia, and dendritic cells.<sup>25, 32</sup> The gp120 glycoprotein on the viral surface binds to the CD4 receptor resulting in a conformational change of the gp120 that enables binding to a co-receptor on the cell surface, CC chemokine receptor 5 (CCR5) or CXC chemokine receptor 4 (CXCR4).<sup>25, 27, 30, 33</sup> The virus envelope hereafter fuses with the cell membrane and the viral capsid is released into the cytoplasm.<sup>25, 27, 30</sup> In the next step viral RNA is translated into DNA by the viral enzyme reverse transcriptase and uncoating occurs.<sup>25, 27</sup> For a long time it has been believed that the uncoating occurs in the cytoplasm, either soon after the viral entry into the cell, stepwise, or at the nucleus.<sup>34, 35</sup> Interestingly, new data propose that the capsid disassembles in the nucleus and that the transcription process is completed within the capsid inside the nucleus.<sup>34, 36, 37</sup> In the nucleus the second viral enzyme, integrase, integrates the proviral DNA into the host genome.<sup>25, 30</sup> The viral DNA is hereafter transcribed by the cell RNA polymerase II to viral RNA and transported to the cytoplasm.<sup>27</sup> The ribosome translates viral RNA to three precursor polyproteins, Gag, Gag-pol, and Env, and the accessory and regulatory proteins.<sup>27, 38</sup> These assemble with viral RNA at the cell membrane and subsequently bud off as a new virion.<sup>27, 38</sup> Concomitantly, the third viral enzyme, the protease, splits the precursor proteins to the structural and enzymatic proteins resulting in a conformational change and the final maturation of the virus.<sup>27, 30, 38</sup>

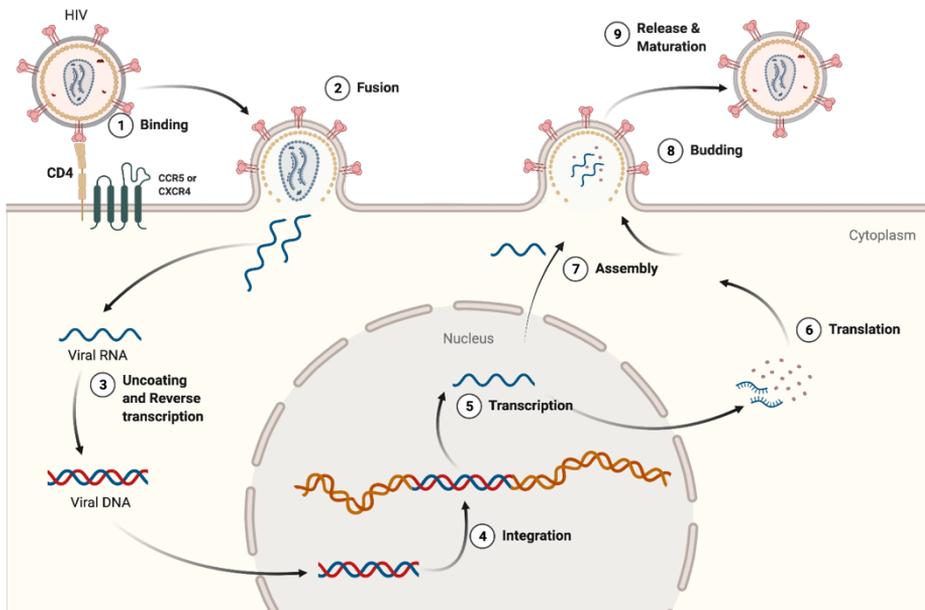


Figure 3. The life cycle of HIV.<sup>39</sup> The virion binds to the CD4 receptor and a co-receptor (CCR5 or CXCR4) (1) and then enters the cell by fusion (2). In the next step uncoating and reverse transcription of viral RNA to DNA by the reverse transcriptase takes place (3). The viral DNA is then integrated into cell DNA by the viral integrase (4). After transcription of viral RNA (5), RNA is translated to viral proteins (6). The viral proteins and two strands of viral RNA assemble at the cell membrane (7) where it buds off as a new virion (8). In the last step the viral protease splits the precursor proteins and a new infective virion is produced (9).

The HIV reverse transcriptase lacks proof reading, making it prone to errors. In combination with the high rate of replication (est.  $10^{10}$  per day in untreated HIV infection), this gives rise to frequent mutations.<sup>30</sup> Furthermore, frequent recombination occurs.<sup>40</sup> The high variability is the basis for how the virus evades the immune response, develops resistance to ART, and one of the reasons why developing a vaccine is so challenging.<sup>25</sup>

## 1.2.2 TROPISM

The HIV strains are divided into two major groups, R5 and X4 viruses. The basis is their use of co-receptor, where R5 virus use the CCR5 receptor and X4 virus the CXCR4 receptor.<sup>33, 41</sup> The R5 virus is dominant during establishment of infection and the early stages of infection.<sup>33, 42</sup> Interestingly, individuals with a homozygous mutation in the gene coding the CCR5 receptor (CCR5 $\Delta$ 32) are protected from HIV infection with R5 virus.<sup>33, 43</sup> The emergence of X4 virus is

seen in many individuals during the course of infection, and is associated with disease progression and loss of immune function.<sup>30, 33, 44</sup> The CCR5 receptor is in addition to T cells found on macrophages, monocytes, dendritic cells, giving rise to the older term macrophage tropic (M-tropic) for R5 virus.<sup>42</sup> Similarly, X4 virus was previously termed T lymphocyte tropic (T-tropic), since the CXCR4 receptor is primarily found in T cell lines.<sup>31, 33</sup> In addition, dual tropic, able to bind both CXCR4 and CCR5 receptors, exist.<sup>25, 33</sup>

### 1.3 FROM HIV TO AIDS

HIV transmission can occur through three different pathways. The most common route is sexual contact. Other routes of transmission are vertical transmission from mother to child and exposure to blood or blood products (e.g. intravenous drug use or iatrogenically).<sup>45</sup>

How HIV crosses the mucosal barrier is not known in detail but infection can occur both by free virus and cell-associated virus.<sup>32</sup> HIV initially infects dendritic cells, macrophages, and T cells in the submucosa but rapidly spreads to local lymph nodes and is subsequently disseminated to the bloodstream and other organs such as the gastrointestinal tract, spleen and bone marrow where infection of a large number of cells occurs.<sup>25, 30, 42</sup> In this initial stage of infection, the lack of specific immune responses allow the virus to rise rapidly to peak levels, resulting in a simultaneous decline in CD4<sup>+</sup> cell count. This coincides with the acute retroviral syndrome seen 2–4 weeks after transmission in a majority of individuals.<sup>45</sup> Clinically, acute infection often presents with flu-like or mononucleosis-like symptoms, but can present with a range of other symptoms such as rash, meningitis, or diarrhoea.<sup>25, 45, 46</sup> Because of the unspecific picture, acute symptomatic infection is often not identified as HIV, and hence often passes undiagnosed. The symptoms in acute infection are self-limiting, and resolve in the course of one to two weeks.<sup>25</sup>

As the immune response awakens, the immune system takes partial control over the infection, viral levels drop, and the infection enters its asymptomatic chronic phase.<sup>25, 31</sup> During this phase the viral levels reach a steady-state, called the viral set point, probably related to the emergence of HIV-specific CD8<sup>+</sup> cytotoxic T cells.<sup>30, 47</sup> The set point level predicts the progress to advanced disease.<sup>48</sup> In some individuals the viral levels drop to very low levels. These individuals are called elite controllers.<sup>49</sup> Parallel to the decrease in viral load, the CD4<sup>+</sup> cell count recovers but usually not to pre-transmission levels.<sup>25</sup>

Although clinically asymptomatic, active viral replication continues throughout the chronic phase, eventually resulting in progressive CD4<sup>+</sup> T cell loss, both in peripheral blood and mucosal tissues, and destruction of the lymphoid tissue.<sup>25, 30</sup> The number of infected CD4<sup>+</sup> T cells are too few to solely explain the quantity of cell loss. It is considered that the chronic activation of the immune system plays a crucial role in the depletion of T cells and pathogenesis of HIV. This is supported by the impact not only on CD4<sup>+</sup> T cells but also on CD8<sup>+</sup> T cells, B cells and NK cells.<sup>50</sup> Over time, the immune system weakens and the immune function deteriorates to a point where the individual is at risk of contracting opportunistic infections (infections that normally do not affect the immunocompetent host) and malignancies, such as Kaposi sarcoma and lymphomas. HIV has developed to AIDS.<sup>25, 30</sup> There are a range of AIDS-defining diagnoses stipulated by the CDC (table 1), which usually occur when CD4<sup>+</sup> cell levels drop below 200 cells/mm<sup>3</sup>. The time lapse from acute infection to onset of AIDS has a large interindividual variation, but is approximately 10 years.<sup>25</sup> (Fig. 4)

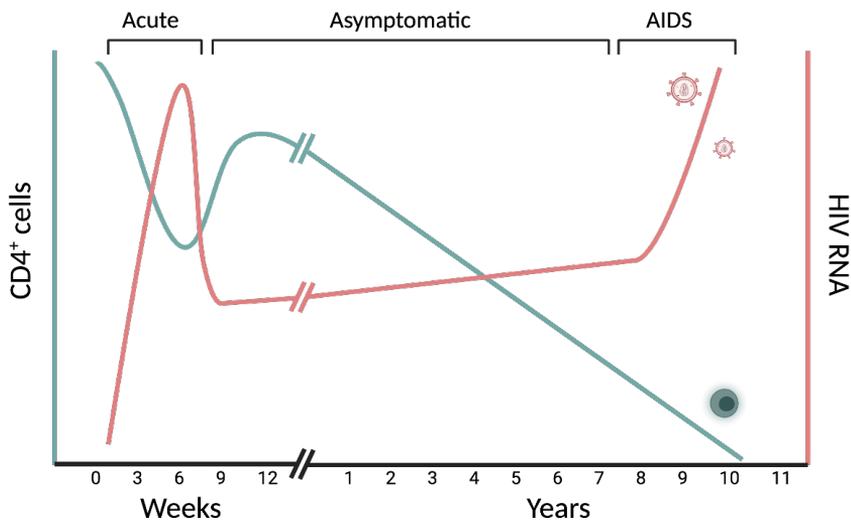


Figure 4. The natural course of HIV infection.<sup>51</sup>

Table 1. AIDS-defining diagnoses.

### **AIDS-defining diagnoses for adults**

Candidiasis of bronchi, trachea, lungs, or esophagus  
Cervical cancer, invasive  
Coccidioidomycosis, disseminated or extrapulmonary  
Cryptococcosis, extrapulmonary  
Cryptosporidiosis, chronic intestinal (>1 month)  
CMV disease (other than liver, spleen, or nodes)  
CMV retinitis  
Encephalopathy, HIV-related  
Herpes simplex: chronic ulcers (>1 month); bronchitis, pneumonitis, esophagitis  
Histoplasmosis, disseminated or extrapulmonary  
Isosporiasis, chronic intestinal (>1 month)  
Kaposi sarcoma  
Lymphoma (Burkitt, immunoblastic, or primary, of brain)  
*Mycobacterium avium* complex or *M. kansasii*, disseminated or extrapulmonary  
*Mycobacterium tuberculosis*, of any site  
*Mycobacterium*, other species or unidentified species, disseminated or extrapulmonary  
*Pneumocystis jirovecii* pneumonia  
Pneumonia, recurrent  
Progressive multifocal leukoencephalopathy  
*Salmonella* septicemia, recurrent  
Toxoplasmosis of brain  
Wasting syndrome due to HIV

## **1.4 ANTIRETROVIRAL THERAPY – THE PARADIGM SHIFT**

Before the introduction of ART, HIV infection was a fatal disease in almost every case. This led to intense research efforts to find effective drugs. The first drug was approved in 1987, the nucleoside reverse transcriptase inhibitor (NRTI) zidovudine. The first randomised trial with a duration of 24 weeks showed benefit of treatment in a group of individuals with late stage disease.<sup>52</sup> But subsequent trials of treatment in earlier stages of disease reported disheartening results,

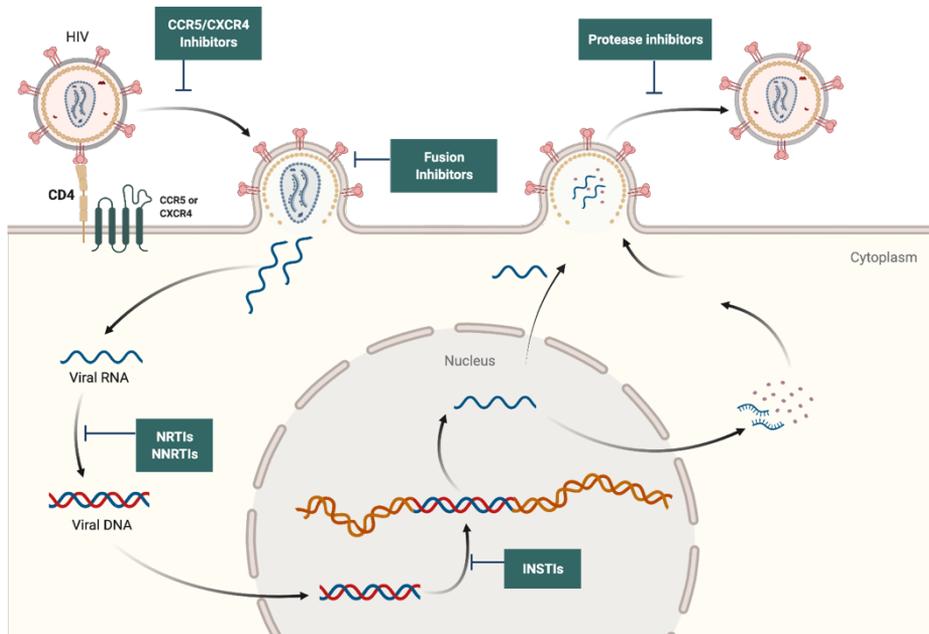
without benefit on disease progression.<sup>53</sup> During the early 90s zidovudine was followed by additional NRTIs, also these without lasting effect.

The big breakthrough came with the introduction of the first protease inhibitors (PIs) in 1995 and the use of combined ART. During the following years, studies showed the durable viral suppression and clinical benefit of combining a PI with two NRTIs compared to earlier NRTI regimens.<sup>54-56</sup> The dramatic effect of combination therapy was well illustrated by the decline in mortality from 29.4 deaths per 100 person-years in 1995 to 8.8 in 1997, showed by Palella et al.<sup>55</sup> Based on the rate of viral decay during treatment, it was estimated that HIV could be cured after 2–3 years of ART but this initial hope was turned into disappointment with the discovery of latent viral reservoirs that were not susceptible to available treatment regimens, as discussed later.<sup>57, 58</sup>

Although effective, ART was also associated with side effects and adverse events. The early drugs induced metabolic changes such as lipodystrophy and there was a fear of increased cardiovascular risk.<sup>59, 60</sup> This gave rise to the idea of treatment-sparing strategies,<sup>61</sup> but initial studies showed discordant results.<sup>62</sup> The SMART (Strategies for Management of AntiRetroviral Therapy) study compared continuous ART with deferred treatment. In the deferred treatment arm, treatment was guided by CD4<sup>+</sup> cell counts, whereby an individual who dropped below 250 cells/mm<sup>3</sup> in CD4<sup>+</sup> cell count initiated ART and subsequently stopped when the CD4<sup>+</sup> cell count raised above 350. In January 2006 the SMART trial was prematurely ended when it became clear that the continuous treatment arm not only experienced less AIDS-related morbidity and mortality, but also less non-AIDS-related morbidity and mortality.<sup>62</sup>

Almost a decade later the results of the START (Strategic Timing of AntiRetroviral Treatment) study were published adding to the knowledge of the beneficial effects of ART. At the time, initiation of ART was recommended to start at a CD4<sup>+</sup> cell count of 350 cells/mm<sup>3</sup> in asymptomatic individuals. The START study randomised participants to either immediate initiation of ART, regardless of CD4<sup>+</sup> cell levels, or to commence ART when the CD4<sup>+</sup> cell count was  $\leq$  350, with the aim of studying the risks and benefits of early ART. In May 2015 the study was stopped early because of the benefits of early ART seen regarding serious AIDS-related events and serious non-AIDS-related events.<sup>63</sup> This knowledge led the way to the revision of treatment guidelines globally to recommend treatment to all, independent of CD4<sup>+</sup> cell count.

Today the arsenal of ART in Sweden consists of five different drug groups with different mechanisms of action. In addition to the above mentioned NRTIs and PIs they include: non-nucleoside reverse transcriptase inhibitors (NNRTIs), integrase inhibitors (INSTIs) and entry inhibitors (EIs). Their mechanisms of action are described in figure 5.



*Figure 5. Entry inhibitors (CCR5/CXCR4 and fusion inhibitors) block the receptor-binding, fusion and entry of the virus into the cell. NRTIs/NNRTIs inhibit reverse transcription, and thus the translation of viral RNA to DNA. INSTIs interfere with the viral enzyme integrase, and thus inhibit the incorporation of viral genome in the DNA of the cell. PIs block the enzymatic cleavage of precursor proteins, inhibiting maturation of functional virions.<sup>64</sup>*

The first Swedish national recommendations for the treatment of HIV were written in the late 1990s and have since been updated on a regular basis. The standard regimen still today consists of a backbone of two NRTIs and one third agent, either an INSTI, a PI, or NNRTI. When starting ART in a treatment naïve individual, the first line recommendation today in Sweden is a combination of the two NRTIs tenofovir and emtricitabine and the INSTI dolutegravir or the NNRTI doravirine, although treatment can be individualised as needed. In the course of the last years a two-drug regimen consisting of lamivudine and dolutegravir has been shown to be effective and is now available as a switch therapy after an initial three drug regimen in patients with a high level of compliance and no pre-existing drug resistance. The most recent additions to the

arsenal are long-acting intramuscular injection formulas. The recommended antiretroviral drugs in Sweden today are presented in table 2.

Group	Generic name	Abbreviation	Trade name
<b>NRTI</b>	abacavir	ABC	Ziagen / Abacavir
	emtricitabine	FTC	Emtriva
	lamivudine	3TC	Epivir / Lamivudine
	tenofovir disoproxil	TDF	Viread / Tenofovir disoproxil
	tenofovir alafenamide	TAF	Vemlidy
<b>NNRTI</b>	efavirenz	EFV	Stocrin / Efavirenz
	nevirapine	NVP	Viramune / Nevirapine
	etravirine	ETR	Intelence
	rilpivirine	RPV	Edurant / Rekambys (injection)
	doravirine	DOR	Pifeltro
<b>PI</b>	atazanavir	ATV	Reyataz / Atazanavir
	darunavir	DRV	Prezista / Darunavir
	ritonavir	RTV	Norvir / Ritonavir
<b>INSTI</b>	raltegravir	RAL	Isentress
	dolutegravir	DTG	Tivicay
	cabotegravir	CAB	Vocabria
	bictegravir	BIC	(Not available as single drug)
	elvitegravir	EVG	(Not available as single drug)
<b>EI</b>	maraviroc	MVC	Celsentri
	enfuvirtide	T-20	Fuzeon
	fostemsavir	FTR	Rukobia
	ibalizumab	IBA	Trogarzo

Table 2. Recommended antiretroviral drugs in Sweden 2021.

### 1.4.1 TREATMENT AS PREVENTION

With neither a cure nor a potent vaccine in the near horizon, effective ART remains our best way of preventing HIV transmission and halting the epidemic. Transmission risk is related to HIV RNA viral load.<sup>65</sup> The first notion of ART as a prevention strategy came with a publication in 1994 by Connor et al., who showed that zidovudine treatment to the mother peripartum and to the child postpartum reduced the risk of vertical transmission of HIV.<sup>66</sup>

In the last decade several studies have investigated the risk of transmission of HIV during ART by studying heterosexual and homosexual serodiscordant couples.<sup>67-69</sup> These studies have presented mounting evidence that HIV is not sexually transmittable when HIV RNA viral load is undetectable in standard assays.<sup>68-72</sup>

This has subsequently led to the revision of the Swedish Communicable Diseases Act, and the rules of conduct, such as the requirement of condom use.

Furthermore, prophylactic ART, preexposure prophylaxis (PrEP), has been proven to effectively prevent HIV transmission in high-risk groups.<sup>73-75</sup>

## 1.5 LATENCY AND RESERVOIRS

In 1995 the first study showing evidence of latent infection was published.<sup>76</sup> Latency is defined as resting cells with integrated proviral DNA that can give rise to viral replication upon stimulation and subsequent activation of the resting cell.<sup>77</sup> This latent infection allows viral persistence despite ART,<sup>78-80</sup> and also evades elimination by the immune system. Latently infected cells are primarily resting memory T cells, but latency can also occur in other cell types such as monocytes, macrophages, and follicular dendritic cells.<sup>81</sup> These latently infected cells constitute the cellular reservoir.

Latent infection is established very early after transmission, probably within days. ART initiation during the acute phase does not hinder the establishment of the reservoir but diminish the size.<sup>81</sup> Only one out of a million resting T cells are latently infected<sup>82</sup> but due to their long half-life ART would have to continue for over 70 years to eradicate the reservoir.<sup>58, 83</sup> Consequently, the reservoir makes cure of HIV through ART unfeasible.

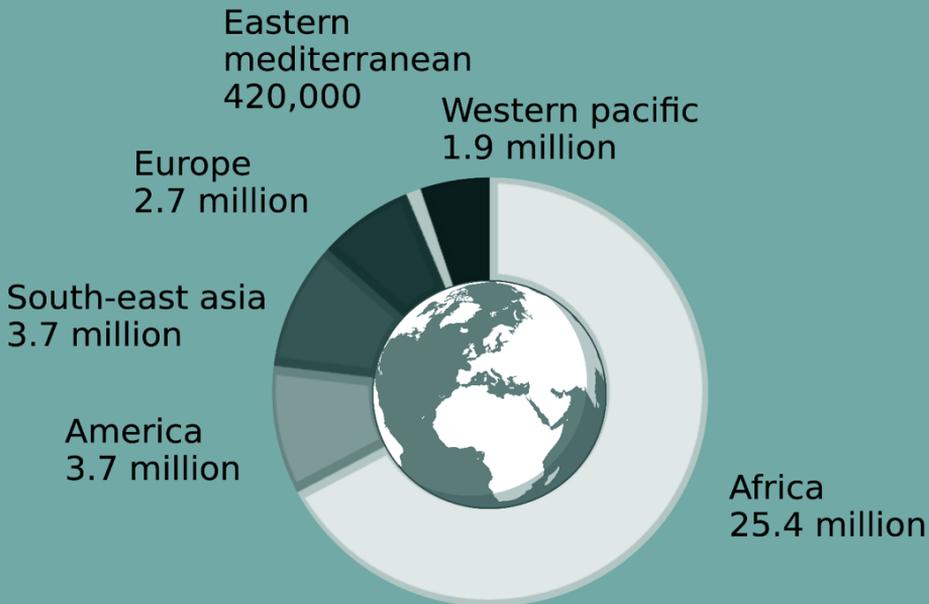
In addition, the reservoir concept is also used to describe anatomical sites where persistence of replication-competent virus may occur.<sup>77</sup> Lymph nodes, gut-associated lymphoid tissue, central nervous system (CNS) and the genital tract are proposed to be such anatomical reservoirs.<sup>81</sup>

## 1.6 HIV TODAY

Up till today the HIV pandemic has claimed more than 36 million lives. Today 37.7 million people live with HIV globally, whereof approximately 8,000 in Sweden (InfCare HIV, December 2021). Even though there is much work left to do, the testing, treatment and prevention strategies are successful. Compared to 2010, 30% fewer people were newly diagnosed and 47% fewer died of HIV-related causes in 2020.<sup>84</sup> In large, this is attributable to the increasing access to ART. The current global situation can be viewed on the next page.<sup>51, 84</sup>

# HIV Globally 2020

37.7 million people living with HIV



73% had access to ART



1.5 million people were infected with HIV



680,000 people died of HIV-related causes

## 1.7 HIV AND AGING

As a result of the success of ART, many PLHIV now reach an older age. In the US today over 50% of PLHIV are 50 years of age or older,<sup>85</sup> and it has been estimated that by 2030 73% of PLHIV in the Netherlands will be over 50 years old.<sup>86</sup> Notably, the prevalence in the age group over 65 years increased with 48% from 2015–2019.<sup>85</sup> In Sweden today 49.5% of those living with HIV are aged 50 years or older and 12.4% are aged 65 years or older (InfCare HIV December 2021). In countries with good access to modern ART and care, the life expectancy of PLHIV is approaching the life expectancy of the general population.<sup>87-89</sup> However, globally, the effect on life expectancy and mortality differs between countries.<sup>90, 91</sup>

There is a general lack of data on elderly PLHIV due to underrepresentation and exclusion from trials. With the increasing age of PLHIV, understanding the particular aspects of aging with HIV has emerged as a research field of great importance.

### 1.7.1 COMORBIDITIES

Aging is associated with an increasing prevalence of comorbidities independent of HIV status.<sup>92-94</sup> However, PLHIV have a higher prevalence of comorbidities compared to HIV negative individuals in the same age group.<sup>94-98</sup> The association between HIV and other comorbidities have been most extensively studied in cardiovascular diseases,<sup>95, 99-101</sup> but associations have also been found with other diseases such as hypertension,<sup>94</sup> chronic renal disease,<sup>94, 95</sup> dyslipidemia,<sup>94</sup> osteoporosis,<sup>102</sup> and cancer.<sup>103</sup> The mechanism underlying this elevated risk is not completely understood but several contributing factors have been proposed, including both HIV-associated and non-HIV-associated.<sup>104</sup> A) A higher frequency of traditional risk factors, such as smoking, in PLHIV.<sup>105, 106</sup> B) A higher frequency of other lifestyle related risk factors, such as alcohol consumption/abuse and substance use.<sup>97, 98</sup> C) A higher frequency of infection with other viruses, such as hepatitis B, hepatitis C and cytomegalovirus.<sup>94, 97</sup> D) ART related toxicity.<sup>107, 108</sup> E) HIV-related causes, such as direct effect of the virus, CD4<sup>+</sup> nadir, and time with immunodeficiency.<sup>95, 96</sup> F) Ongoing low-grade inflammation and immune activation.<sup>109-111</sup>

### 1.7.2 ART AND NON-ART DRUGS

The increased prevalence of comorbidities associated with aging goes hand in hand with an increased number of concomitant drugs. Concomitant medications in turn increase the risk for potential drug-drug interactions (PDDIs).<sup>112</sup> This is of particular importance in PLHIV as they have a higher frequency of

polypharmacy (not including ART) compared to HIV negative individuals in the same age group.<sup>113</sup> During the last years several international guidelines have included recommendations on HIV in the older person. They all highlight the need to take comorbidities and concomitant medications into consideration when caring for the older individual living with HIV.<sup>114-116</sup> Possible ART drug toxicities need to be carefully considered since older individuals are especially vulnerable to drug toxicities.<sup>117</sup>

Aging is related to numerous physiological changes affecting organ function. These changes include alterations that may affect the pharmacokinetics of drugs. With age kidney function decreases with a decline in glomerular filtration rate affecting the clearance of drugs. Furthermore, the mass of, and the blood flow to the liver is reduced, affecting the metabolism and elimination of drugs. Currently, data do not support relevant age-related differences in enzymatic function.<sup>118, 119</sup> Additionally, alterations in body composition with reduced total body water and lean body mass lead to a relative increase in body fat and altered volume of distribution for water-soluble and lipid-soluble drugs.<sup>118</sup> Albumin levels may not be decreasing with age per se but are decreased in acute illness and malnutrition, and may affect protein binding of drugs. Pharmacodynamic age-related effects often result in increased sensitivity to drugs. Altogether these alterations may lead to toxic drug effects, particularly for drugs with a narrow therapeutic window.<sup>118</sup> In addition, the interindividual variability increases with age making effects of drugs less predictable and comorbidities may in turn affect organ function.<sup>118, 120</sup> Although the pharmacokinetic effects of aging in PLHIV have not been studied it is reasonable to believe that studies on HIV-negative individuals can be applied on PLHIV, with the addition of possible organ damage related to HIV infection.<sup>120</sup>

### 1.7.3 INFLAMMATION AND IMMUNE ACTIVATION

The grade of inflammation and immune activation caused by HIV decline during ART. But despite viral suppression a low grade of chronic inflammation persists which is elevated compared to HIV negative individuals.<sup>121-123</sup> As stated above this inflammation is suggested to be a contributor to the increased risk of non-communicable diseases seen in PLHIV.<sup>109-111</sup> Different mechanisms have been proposed to give rise to the persistent inflammation such as low level residual viremia, co-infections (e.g. cytomegalovirus), lifestyle factors, and microbial translocation.<sup>124, 125</sup>

The choice of markers of inflammation and immune activation analysed in paper II were based on findings of earlier studies.<sup>126</sup> They are briefly described below.

Soluble (s) intercellular adhesion molecule-1 (ICAM-1) is present on endothelial cells and can be used as a marker of endothelial activation. It is upregulated after inflammatory stimuli and helps leukocytes migrate to the tissues.<sup>127</sup> High-sensitive C reactive protein (hsCRP) is an acute phase protein produced in the liver and is a marker of systemic inflammation.<sup>128</sup> Interleukin-6 (IL-6) is a proinflammatory cytokine that is involved in several physiologic processes, as the acute phase response.<sup>129</sup> It is secreted by a variety of cells, particularly monocytes/macrophages on stimulation.<sup>104</sup> C-X-C motif chemokine ligand 10 (CXCL10) is a chemokine induced by interferon gamma and secreted by various cells but predominantly by monocytes.<sup>130</sup> An important function is to recruit leukocytes to sites of infection.<sup>131</sup> s-cluster of differentiation 163 (sCD163) and sCD14 are markers of monocyte/macrophage activation.<sup>125, 132</sup> sCD14 is particularly associated with microbial translocation.<sup>125</sup> Both s-tumour necrosis factor receptor-II (sTNFRII) and sCD27 are soluble cytokine receptors belonging to the TNF superfamily and markers of T cell activation.<sup>126, 133, 134</sup> Matrix metalloproteinase-3 (MMP-3) is upregulated by cytokines and involved in tissue remodeling.<sup>135</sup> s-glycoprotein130 (sgp130) is involved in IL-6 signalling.<sup>136</sup>

## 1.8 HIV IN THE CENTRAL NERVOUS SYSTEM

Neurological symptoms were described in AIDS patients prior to the identification of HIV as the causative agent.<sup>137</sup> In the years after the discovery of HIV, the symptoms and neuropathology of a form of dementia affecting HIV-infected were described.<sup>138, 139</sup> If left untreated at least 20–30% of HIV-infected would develop HIV-associated dementia (HAD), initially called AIDS dementia complex.<sup>140, 141</sup> HAD is caused by HIV replication within the CNS, leading to HIV encephalitis. This is a disease unrelated to the opportunistic infections or malignancies such as cryptococcal meningitis, cerebral toxoplasmosis, progressive multifocal leukoencephalopathy caused by JC virus, or lymphoma that can affect the CNS in the late stages of HIV infection.<sup>142</sup>

Even though HIV infects the CNS shortly after disease transmission, HAD develops during the late immunocompromised stages.<sup>139</sup> HAD gives rise to a triad of cognitive, motor, and behavioural symptoms. Early symptoms include impaired memory and concentration, loss of balance, and social withdrawal which during weeks to months develop to severe dementia.<sup>139</sup>

Although ART prevents the development of HAD, PLHIV still experience milder forms of cognitive symptoms.<sup>143, 144</sup> To adjust to this new situation the neurocognitive complications of HIV are now brought together under the umbrella diagnosis termed HIV-associated neurocognitive disorder, HAND. During the pre-ART era no specific objective diagnostic markers for the diagnosis

of HAD were available. Indeed, the diagnosis relied on criteria based on symptoms, and the exclusion of other diagnoses. This issue still remains and in 2007 a set of criteria, the Frascati criteria, were defined to try to overcome some of the diagnostic difficulties and establish criteria useful for research purposes.

In addition to HAD, the Frascati criteria introduced two new subgroups of HAND, that included HIV-associated mild neurocognitive disorder (MND) and asymptomatic neurocognitive impairment (ANI). The basis of these criteria is neurocognitive testing of at least five domains. To fulfill the criteria of ANI an individual need to perform at least one standard deviation below adjusted normative scores in at least two cognitive domains. For the diagnosis of MND the criteria of ANI need to be fulfilled and in addition give rise to symptoms that interfere with daily functioning. For the most severe form, HAD, neurocognitive testing needs to show a result at least two standard deviations below adjusted normative scores in at least two domains, and the individual must experience a distinct interference with everyday functioning. For all diagnoses the presence of another condition that explains the symptoms is an exclusion criterion.<sup>145</sup>

Studies in the ART era (of note, not all included PLHIV were on ART) have found a high prevalence of HAND, primarily ANI.<sup>146, 147</sup> In addition, studies have suggested ANI to be a predictor of development of symptomatic HAND.<sup>144, 148</sup> Notably, the clinical relevance of ANI, and the results of these studies are controversial.<sup>149, 150</sup> Indeed, the use of neuropsychological testing as the basis of diagnosis is complicated since the result can be confounded by comorbidities and life style factors. Furthermore, the results cannot distinguish ongoing injury from an effect related to previous events. This has led to the search for biomarkers that can facilitate the diagnostics and to identify individuals at risk of progressive disease, which is discussed further below.

### 1.8.1 NEUROPATHOGENESIS – A TROJAN HORSE?

The neuropathogenesis of HIV is still not completely understood. The CNS is probably infected during the first weeks after transmission.<sup>151, 152</sup> It is believed that the virus infects circulating blood monocytes, which in turn migrate to the CNS through the blood-brain barrier and carry the virus like a Trojan horse. Other plausible mechanisms for CNS infection are via infected T cells, or through the transcytosis of free virions through endothelial cells.<sup>153-155</sup>

Monocytes further differentiate into perivascular macrophages, which are the main cell type for HIV infection in the CNS. Virus from these macrophages can in turn infect microglia. Additionally, astrocytes might be infected but do not give rise to viral replication. Infected cells can induce fusion with non-infected cells

expressing CD4- and co-receptors and create multinucleated giant cells. These cells can continue to replicate virus, and is a pathological hallmark of HIV encephalitis.<sup>154</sup>

Neurons, however, are not directly infected by HIV, which raises the question of, how CNS HIV infection can give rise to neurological symptoms. Two different pathways have been proposed. First, a direct neurotoxic effect of viral proteins has been suggested. Vpu, Vpr, gp120 and Tat have all been described as possibly neurotoxic, where at least gp120 and Tat are neurotoxic in vitro. However, it is not certain that the concentrations needed are present in vivo. Second, and more probable, the infection of macrophages and microglia induces secretion of inflammatory mediators and products that can have neurotoxic effects (i.e., quinolinic acid, tumor necrosis factor and arachidonic acid metabolites), induce activation of other cells, and attract additional T cells and macrophages further enhancing inflammation. This mechanism is further supported by the fact that few cells are infected, in comparison to the severity of the disease. In addition, several studies have found signs of macrophage activation in cerebrospinal fluid (CSF) of untreated PLHIV, and this has in turn been related to neuronal damage.<sup>153, 154</sup>

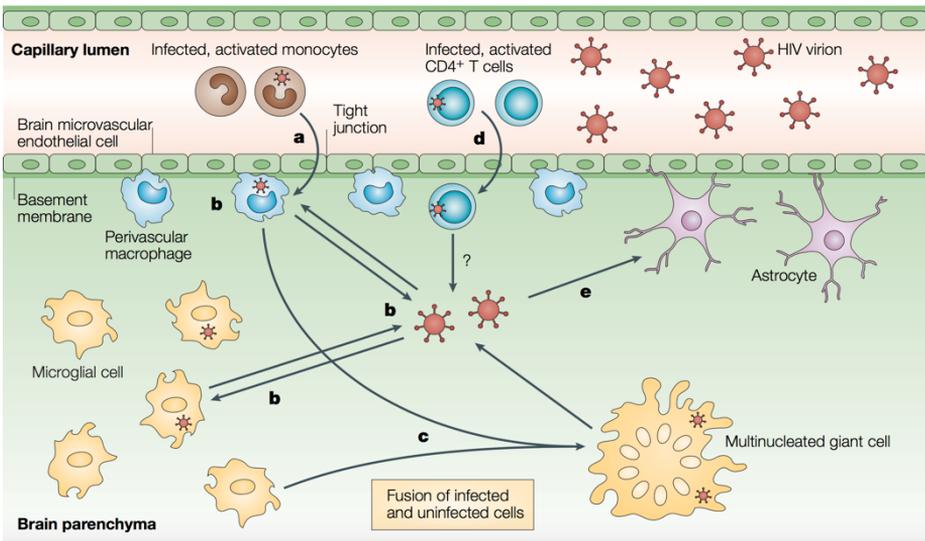


Figure 6. *Neuropathogenesis of HIV. a) HIV-infected circulating blood monocytes migrate across the blood-brain barrier. b) Monocytes differentiate into perivascular macrophages. Virus derived from perivascular macrophages in turn infect microglia. c) Infected cells fuse with uninfected cells creating multinucleated giant cells. d) Another possible entryway of HIV is through infected CD4<sup>+</sup> T cells. e) Astrocytes do not replicate virus, but their malfunction due to the inflammatory response caused by HIV, contribute to neuropathogenesis. Reprinted with permission from Springer Nature: Nature reviews Immunology. The Neuropathogenesis of AIDS, González-Scarano F et al., 2005.*

## 1.8.2 BIOMARKERS OF CNS INFECTION

It is inherently hard to obtain samples from the brain parenchyma. CSF is, by lumbar puncture, accessible and is used as a proxy for pathological processes in the brain. A number of different biomarkers measurable in the CSF, and in some cases in plasma, have proven to be useful in the study of the HIV CNS infection. This section will focus on the biomarkers most relevant to this thesis.

## 1.8.3 NEUROFILAMENT LIGHT PROTEIN

The cytoskeleton in neurons consists of three different components; the microtubules, the microfilaments, and the neurofilaments. In turn the neurofilament is constructed from three neurofilament subunits (the heavy, medium, and light chain), alfa-internexin, and peripherin. They are found in the cytoplasm of all parts of the neuron, but most abundantly in the axon, and particularly of large myelinated neurons. The neurofilament determines the caliber of the axon and hence the conduction velocity.<sup>156, 157</sup> When damage to a neuron occurs, neurofilament is released to the extracellular space. The mechanism for transport to CSF or blood is not fully known.<sup>157</sup>

The neurofilament light protein (NfL) has proven to be a sensitive and specific marker of axonal damage. However, it is not disease specific and elevated levels can be found in the CSF in several neurological disorders such as; multiple sclerosis, amyotrophic lateral sclerosis, cerebral infarction, and traumatic brain injury.<sup>158-160</sup> Furthermore, CSF NfL is elevated in different types of dementia; Alzheimer's disease, vascular dementia, and frontotemporal dementia.<sup>160</sup> In 2016, an assay able to detect NfL in plasma was developed, with a strong correlation between CSF NfL and plasma (P-)NfL.<sup>161</sup> The results of elevated CSF NfL has then been reproduced in P-NfL.<sup>160</sup> In addition, P-NfL may reflect axonal damage of peripheral neurons.<sup>160, 162, 163</sup>

NfL is a sensitive biomarker of neuronal damage in HIV infection. Untreated HIV-infected individuals with HAD or opportunistic CNS infection exhibit very high CSF NfL levels.<sup>164-166</sup> Interestingly, NfL may be increased in neuroasymptomatic individuals, corresponding to the level of immunodeficiency, with higher levels in the late stages.<sup>164-167</sup> In addition, NfL levels may anticipate the development of HAD 1–2 years prior to diagnosis.<sup>168</sup> After initiation of ART, NfL levels decrease and in most, but not all, cases normalise.<sup>165-167, 169, 170</sup> Notably, one study found that PLHIV on ART had CSF NfL levels corresponding to HIV negative individuals 3.9 years older.<sup>169</sup>

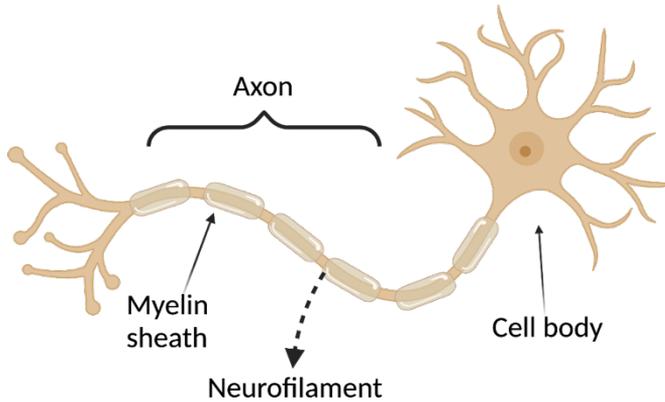


Figure 7. Schematic picture of a neuron.<sup>51</sup>

#### 1.8.4 NEOPTERIN

Neopterin is a biomarker of macrophage/monocyte activation. It is released from macrophages on stimulation from interferons, primarily interferon- $\gamma$ . Neopterin can be measured both in blood and CSF. In blood it has been found to predict HIV disease progression. In untreated HIV infection without neurological symptoms both blood and CSF neopterin rise with progression to immunodeficiency, with the highest levels in those with a CD4<sup>+</sup> cell count below 200 cells/mm<sup>3</sup>. In individuals with HAD, CSF neopterin levels exceed the levels found in patients with neuroasymptomatic immunodeficiency, while a concomitant rise in blood neopterin is absent.<sup>171</sup> ART effectively reduce neopterin levels both in blood and in CSF but despite years of treatment elevated levels of neopterin may persist.<sup>170</sup>

#### 1.9 B VITAMINS & HOMOCYSTEINE

The history of the B vitamins in medicine dates back to the 19<sup>th</sup> century. Then a fatal form of anemia with macrocytic red blood cells, related to atrophic gastritis, was described. It was named pernicious anemia. In addition to anemia, concomitant neurological symptoms were reported. In the mid 1920s it was discovered that raw liver could cure pernicious anemia, and in 1948 vitamin B<sub>12</sub>, the compound that gave rise to the effect, was isolated and the chemical structure subsequently described.<sup>172</sup> Parallel to this discovery, the physician Lucy Wills investigated macrocytic anemia in pregnant women in Bombay and discovered

that yeast extract could cure the anemia. In the 1940s it was discovered that the active agent was folate, and it was subsequently industrially synthesised. After the isolation of both enzymes it was found that their deficiency gave rise to the same pathologic picture in the bone marrow, and later it was discovered that the vitamins are indeed metabolically closely related.<sup>173</sup>

Homocysteine, a non-essential amino acid, was first described in 1932. However, it was not until the 1960s that the implication in disease was made, when a number of genetic diseases caused by different enzyme defects affecting homocysteine metabolism were described. These diseases gave rise to homocysteinuria and high blood levels of homocysteine, and among other symptoms mental disturbance and arteriosclerotic changes.<sup>172</sup> This was the start of the field of research investigating the role of homocysteine in disease. Increased levels of homocysteine has since been related to over a hundred different conditions, predominantly cardiovascular disease and diseases of the CNS.<sup>174</sup>

Homocysteine is involved in two different metabolic pathways, the remethylation pathway in the methionine cycle and the transsulfuration pathway. In the remethylation pathway homocysteine is methylated to methionine, through a reaction that is both B<sub>12</sub> and folate dependent. An adenosine group is then transferred to methionine, creating S-adenosylmethionine (SAM). SAM donates a methyl group, of importance to several reactions in cells, and is converted into S-adenosylhomocysteine (SAH). SAH is then converted through hydrolysis back to homocysteine. In the transsulfuration pathway homocysteine is metabolised to cysteine, in a reaction dependent of vitamin B<sub>6</sub>. Cysteine is in turn used in the synthesis of the antioxidant glutathione.<sup>175, 176</sup> Since folate is a cosubstrate in the homocysteine metabolism, the metabolism of homocysteine and folate are closely related, which will be discussed below. These different metabolic pathways are described in figure 8.

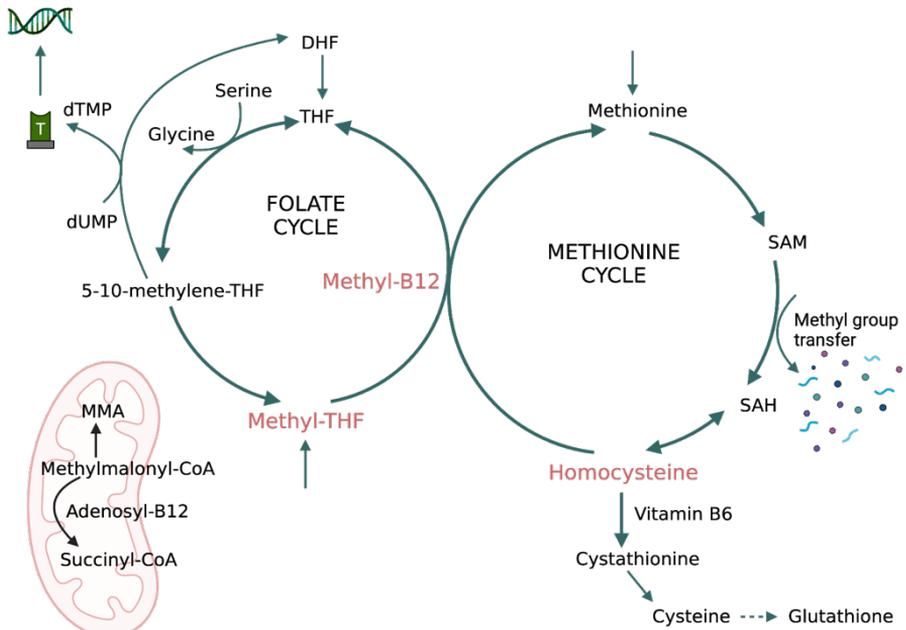


Figure 8. Homocysteine, folate, and vitamin B<sub>12</sub> metabolism.<sup>51</sup>

As mentioned above, one of the causes of high levels of homocysteine, hyperhomocysteinemia, is genetic defects of enzymes involved in the metabolism of homocysteine. Furthermore, deficiency of B<sub>12</sub>, folate, or B<sub>6</sub> hinders homocysteine metabolism and as a consequence increase the homocysteine levels. Levels of homocysteine also increase with age and are inversely related to renal function. In addition, several drugs such as trimethoprim, methotrexate, and anticonvulsants may increase homocysteine.<sup>176, 177</sup>

Homocysteine can be used as a marker of B<sub>12</sub> and folate deficiency, as it increases when B<sub>12</sub> and folate levels decrease. In addition, methylmalonic acid (MMA), described in the next section, can be used as a marker of B<sub>12</sub> deficiency. With the introduction of assays capable of determining homocysteine and MMA in blood, the concept of subclinical deficiency was introduced.<sup>172, 178-180</sup> It was now possible to detect metabolic signs of intracellular deficiency through raised levels of homocysteine and MMA in a subset of individuals without clinical symptoms, and vitamin levels not traditionally regarded as deficient.<sup>178, 179</sup> Studies have followed that show that homocysteine and MMA concentrations start to increase at vitamin levels within the normal reference interval.<sup>181</sup> In the context of cognitive decline,

this has led to the discussion of which cut off levels are appropriate and whether subclinical deficiency may be of clinical relevance.<sup>175, 179, 180</sup>

There is still no consensus or gold standard for the diagnosis of B12 or folate deficiency.<sup>177, 178, 180, 181</sup> However, there is support for using of a combination of metabolic markers (i.e., homocysteine or MMA) and vitamin levels.<sup>177, 178, 180, 182</sup>

Closer descriptions of the two B vitamins of greatest importance for this thesis follow in the next sections.

### 1.9.1 VITAMIN B<sub>12</sub>

Vitamin B<sub>12</sub>, also called cobalamin, is an essential water-soluble vitamin found in animal source foods such as meat, egg, and dairy products. On the contrary, vitamin B<sub>12</sub> is not found in vegetables.<sup>178, 182, 183</sup>

Upon ingestion vitamin B<sub>12</sub> is released from animal proteins in the acid environment of the stomach, where it binds to haptocorrin to prevent its degradation. In the duodenum, vitamin B<sub>12</sub> is released from haptocorrin, and forms a complex with intrinsic factor (IF) which is produced by parietal cells in the stomach. The B<sub>12</sub>-IF complex is absorbed by receptor-mediated endocytosis in the distal ileum. In addition, a small fraction of vitamin B<sub>12</sub> is absorbed through passive diffusion throughout the intestine. Once absorbed in the enterocyte the B<sub>12</sub>-IF complex enters a lysosome where IF is degraded and vitamin B<sub>12</sub> is released. From the basolateral membrane of the enterocyte vitamin B<sub>12</sub> is released into the blood where it binds to one of two possible transport proteins. Approximately 20% of vitamin B<sub>12</sub> in the blood is bound to transcobalamin and constitutes the active form that delivers vitamin B<sub>12</sub> to the tissues. The remaining 80% is bound to haptocorrin and forms a complex that can only be taken up by hepatocytes where it is stored. A portion of the stored vitamin B<sub>12</sub> is secreted in the bile every day and may again be absorbed in the ileum.<sup>178, 182-184</sup>

Inside the cell, vitamin B<sub>12</sub> is converted into its two different biologically active forms. In the cytoplasm, vitamin B<sub>12</sub> is converted into methylcobalamin and is involved in the methionine cycle described in figure 8. In the mitochondria vitamin B<sub>12</sub> is converted into adenosylcobalamin, which is a cofactor for the enzyme methylmalonyl-CoA mutase in the conversion of methylmalonyl-CoA to succinyl-CoA.<sup>178, 183, 184</sup> In the absence of vitamin B<sub>12</sub>, MMA accumulates and can be measured as a marker of B<sub>12</sub> deficiency.

Causes for vitamin B<sub>12</sub> deficiency can be divided into low intake or malabsorption (see table 3). As described above B<sub>12</sub> deficiency can manifest as macrocytic anemia, through pathways related to folate metabolism, described in more detail

in the next section. It can, in addition to anemia, or as the sole manifestation, give rise to various neurologic symptoms such as polyneuropathy, paresthesia, cognitive impairment, and depression. The exact pathogenic mechanism is not clear, though it is known that deficiency can cause demyelination of central and peripheral neurons.<sup>178, 182, 185, 186</sup>

### 1.9.2 FOLATE

Folates is the common name of a family of closely related compounds of water-soluble vitamins. In contrast to vitamin B<sub>12</sub>, folate is found in many plants such as asparagus, broccoli, green leafy vegetables, and whole grain, but also in animal products such as liver. Folate is absorbed in the proximal part of the jejunum through specific folate transporters. Inside the intestinal cells absorbed forms of folate are converted to 5-methyl-tetrahydrofolate (5-methyl-THF). 5-methyl-THF is the form that circulates in blood where it is either unbound or protein-bound, mainly to albumin.<sup>187, 188</sup>

The folate cycle is described in figure 8. 5-methyl-THF is a cosubstrate for methionine synthase, the enzyme converting homocysteine to methionine, and is thus part of the methionine cycle. This reaction converts 5-methyl-THF to THF. In the next step THF accepts a carbon unit from serine forming 5,10-methylene-THF. 5,10-methylene-THF is required for the synthesis of thymidylate and purines, and subsequently important for the synthesis of DNA and RNA. The interruption of this step by the deficiency of folate gives rise to errors in DNA synthesis, the cause of the macrocytic anemia seen in folate deficiency. Even if sufficient levels of folate is available the conversion of 5-methyl-THF to THF is dependent of vitamin B<sub>12</sub> and hence vitamin B<sub>12</sub> deficiency hampers the folate cycle, a phenomenon called the folate trap, and through this pathway leads to macrocytic anemia. 5,10-methylene-THF converts to dihydrofolate which in turn is converted back to THF. Concomitantly 5,10-methylene-THF is converted to 5-methyl-THF in an irreversible reaction.<sup>178, 188, 189</sup>

Similar to B<sub>12</sub>, the classical manifestation of folate deficiency is macrocytic anemia. It may also give rise to neurological symptoms, such as cognitive impairment and depression.<sup>190</sup>

Table 3. Causes of vitamin B<sub>12</sub> and folate deficiency.

	B <sub>12</sub>	Folate
<b>Insufficient intake</b>	Dietary factors (e.g. malnutrition, vegan diet, alcoholism)	Dietary factors (e.g. malnutrition, alcoholism)
<b>Impaired absorption or metabolism</b>	Pernicious anemia (autoimmune gastritis) Chronic atrophic gastritis Gastrointestinal surgery Pancreatic disease Inherited disorders Medications (e.g. proton pump inhibitors, metformin) Bacterial or parasitic infection Inflammatory bowel disease	Gastrointestinal disorders (e.g. Crohn's and coeliac disease) High alcohol intake Gastrointestinal surgery Tobacco smoking Inherited disorders Antifolate medications (e.g. methotrexate, trimethoprim, anticonvulsants)
<b>Increased demand</b>		Pregnancy

### 1.9.3 HOMOCYSTEINE AND NEUROCOGNITIVE DISEASE

The development of cognitive impairment and dementia is multifactorial, where homocysteine may have a contributing role. In the early 1990s two articles were published presenting the hypothesis that homocysteine metabolism might play a role in the etiology of Alzheimer's disease.<sup>191</sup> This was followed by a large number of cross-sectional studies that reported a relationship between homocysteine, and cognitive impairment and dementia.<sup>191</sup> These results have been confirmed by both prospective observational studies and meta-analyses, reporting an increased risk of cognitive impairment or dementia in individuals with elevated homocysteine levels.<sup>192-195</sup> In several studies the association remains after adjusting for B vitamin status, arguing against vitamin deficiency as the sole mechanism.<sup>192, 196, 197</sup> It is possible that the relationship is influenced by another unidentified risk factor or that the risk is due to the combination of several pathways.<sup>198, 199</sup>

Several clinical trials have been performed to examine the effect of vitamin B supplementation on the levels of homocysteine and cognitive decline. These studies have unquestionably showed the homocysteine lowering effect of B

vitamins, but they have had contradictory results on the effect on cognition.<sup>200-205</sup> It has been argued that the lack of effect in many trials is due to inadequate trial design.<sup>174, 206</sup>



## Aims

## 2 AIMS

The general aim of this thesis was to describe HIV infection in the elderly, focusing on drug levels, side effects, drug-drug interactions, comorbidities and inflammation, and to investigate the role of homocysteine (as a marker of vitamin B deficiency) in neuronal injury in PLHIV.

The specific aims were:

- To examine drug levels, side effects, adherence, concomitant medications, drug-drug interactions, and comorbidities in PLHIV aged 65 years or older compared to a group of PLHIV aged 49 years or younger.
- To analyse levels of inflammatory markers in PLHIV aged 65 years or older in individuals with three different ART regimens.
- To explore the relationship of homocysteine levels and signs of neuronal injury in PLHIV.
- To investigate the effect of vitamin B supplementation on signs of neuronal injury in PLHIV with raised levels of homocysteine.

## Study population and design

### 3 STUDY POPULATION AND DESIGN

The four papers in this thesis are based on three studies. All PLHIV included in the studies attended the Department of Infectious Diseases at Sahlgrenska University Hospital, Gothenburg; South Älvsborg Hospital, Borås; Karolinska University Hospital, Huddinge; or Stockholm South General Hospital, Stockholm. The distribution of study participants is shown in figure 9.

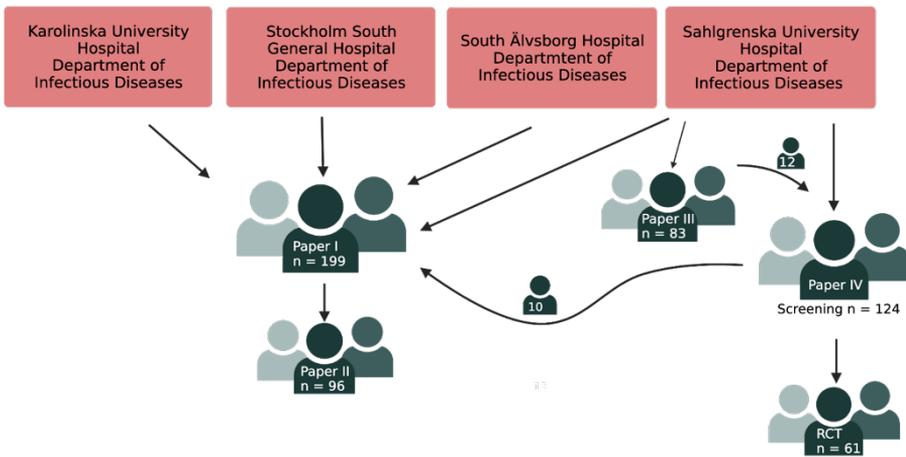


Figure 9. The distribution of study participants in the thesis papers.

#### 3.1 PAPER I & II

The basis of paper I and II is a multi-center cross-sectional study performed at four sites in Sweden: the Department of Infectious Diseases at Sahlgrenska University Hospital in Gothenburg; the Department of Infectious Diseases at South Älvsborg Hospital in Borås; the Department of Infectious Diseases at Karolinska University Hospital Huddinge in Stockholm; and the Department of Infectious Diseases at Stockholm South General Hospital in Stockholm. The aim was to study aspects of HIV infection in elderly individuals, defined as aged 65 years or older, compared to PLHIV under the age of 50. The inclusion criteria were age ( $\geq 65$  years for the study group and 18–49 years in the control group), HIV infection, and stable ART for at least 6 months, with a drug regimen containing atazanavir (ATV), darunavir (DRV), or efavirenz (EFV).

Inclusion began in November 2013. At that time 3568 PLHIV were receiving HIV care at the study sites. Two hundred fifty-six (7.2%) were 65 years or older, whereof 38 (14.8%) were treated with ATV, 56 (21.9%) with DRV, and 78 (30.4%) with EFV. Patients were offered to participate during ordinary routine clinical follow-up. If enrolled, a blood sample for analysis of drug concentrations (paper I), markers of inflammation (paper II), and data on concomitant medications, side effects, and adherence were collected. Further information on comorbidities and routine blood test results were recorded through systematic review of medical charts. PDDIs were analysed using drug-drug interaction databases.

From November 2013 to August 2015, 100 individuals were included in the study group and 99 in the control group. In paper I, 27 individuals were excluded from the drug concentration analysis, due to sample management (3) or dosing (24) issues. All enrolled individuals were included in the other analyses in paper I. Only individuals over the age of 65 years were included in paper II, whereof 4 were excluded due to ongoing bacterial or fungal infections (3) or treatment with two of the study drugs (1).

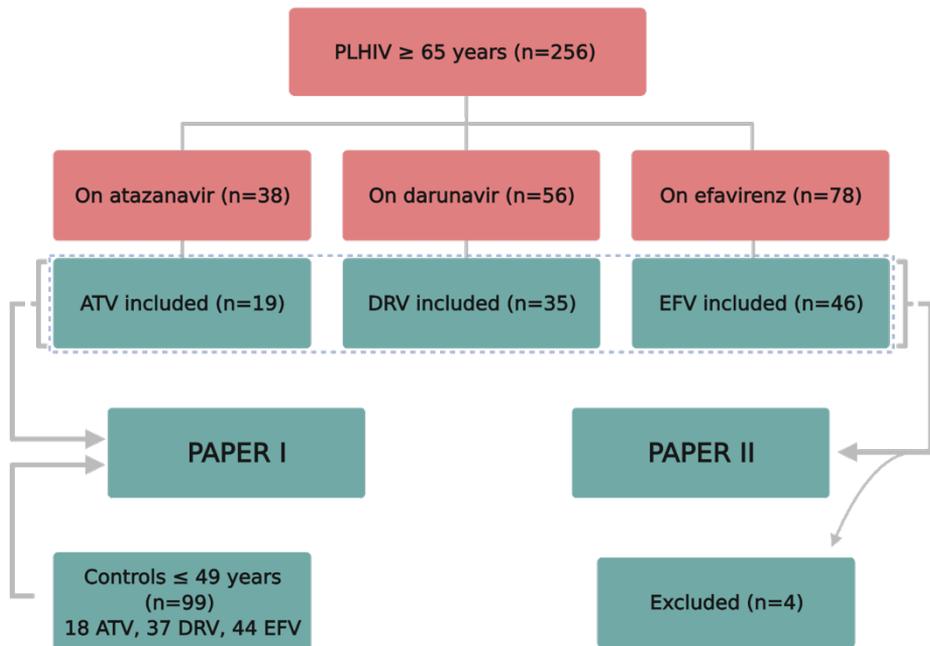


Figure 10. Study design of the multi-center cross-sectional study.<sup>51</sup>

### 3.2 PAPER III

Since 1985, a longitudinal prospective study of the effects of HIV on the CNS has been ongoing at the department of Infectious diseases at Sahlgrenska University Hospital in Gothenburg. The study includes blood and CSF sampling with lumbar punctures performed before and after treatment initiation, after change of ART regimen, and annually during follow-up. Currently, 648 PLHIV have been included in the longitudinal study. In 2014, at the time for the study described in paper III, 494 had been included.

The aim of the study was to investigate the relationship between P-homocysteine and signs of neural injury in PLHIV. Based on the inclusion criteria (untreated HIV infection, age  $18 \geq$  years, no opportunistic CNS infection) 83 HIV-infected individuals from the prospective study were randomly included in the retrospective study described in paper III. A paired CSF and serum sample was analysed, and in 22 individuals a second paired sample was analysed after ART initiation.

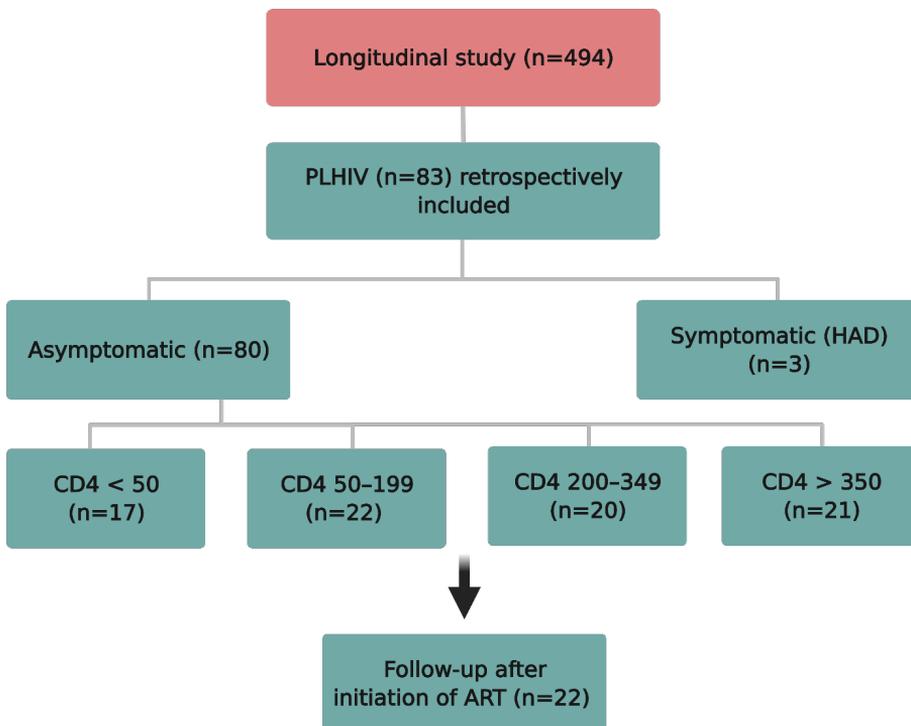


Figure 11. Study design of the retrospective study.<sup>51</sup>

### 3.3 PAPER IV

To further explore the association between P-homocysteine and NfL found in paper III we designed an open randomised controlled trial to study the effect of vitamin B supplementation on NfL, as a marker of neuronal injury. The inclusion criteria were stable ART for > 12 months, HIV RNA < 50 copies/ml, and age  $\geq$  18 years. The exclusion criteria were; treatment with trimethoprim-sulfamethoxazole or methotrexate, ongoing vitamin B<sub>12</sub>, B<sub>6</sub> or folic acid substitution, antiepileptic treatment, small intestine or ventricular resection, disturbed absorption in the small intestine, ongoing neurological or severe psychiatric disease, any malignant tumor in the history, severe ongoing infection or opportunistic infection, alcohol abuse, clinical depression, pregnancy, and significant vitamin B<sub>12</sub> or folate deficiency that require higher treatment doses. Individuals who met the criteria were screened, and those with P-homocysteine  $\geq$  12  $\mu$ mol/L were enrolled in the trial. The participants were randomised to either active treatment arm or control arm, with cross over from the control arm to the active treatment arm after 12 months. The trial continued for in total 24 months. The active treatment consisted of 1 tablet Triobe (cyanocobalamin 0.5 mg, folic acid 0.8 mg, and pyridoxine 3.0 mg) q.d. Based on power calculations it was decided to include 30 PLHIV in each arm.

All study participants were included at the department of Infectious diseases at Sahlgrenska University hospital in Gothenburg from April 2016 to June 2017. The last study visit was performed in June 2019.

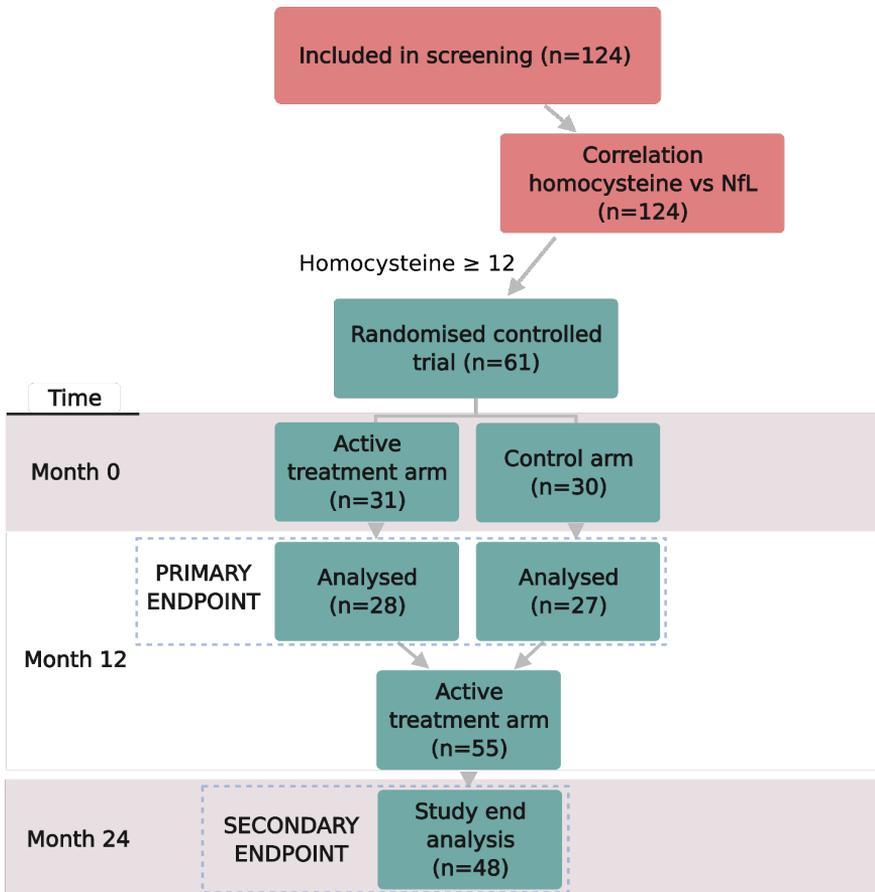


Figure 12. Study design of the randomised controlled trial.<sup>51</sup>

# Methods

## 4 METHODS

### 4.1 LABORATORY ASSAYS

Blood samples analysed for drug concentrations (paper I), markers of inflammation (paper II) and P-NfL (paper IV) were frozen to  $-70$  degrees directly after sampling until analysis. The additional blood tests analysed in paper IV were analysed directly after sampling. In paper III thawed CSF and blood samples were analysed.

If not otherwise stated the blood sample analyses were performed at the local laboratories according to local standards.

#### 4.1.1 DRUG CONCENTRATIONS

Steady-state drug levels of ATV, DRV, and EFV were analysed at the pharmacology analytical laboratory at Karolinska University Hospital, Huddinge in Stockholm, Sweden. The method used was a reverse-phase High Pressure Liquid Chromatography with ultraviolet detection.

#### 4.1.2 MARKERS OF INFLAMMATION

The following analyses were performed at the department of Clinical Microbiology at Sahlgrenska University Hospital, Gothenburg, Sweden.

Plasma concentrations of sCD14, sCD27, sCD163, sgp130, IL-6, sTNFR1I, CXCL-10, sICAM-1, and MMP-3 were measured using the premixed Magnetic Luminex Assay (R&D Systems, Minneapolis, Minnesota, USA) according to the manufacturer's instructions.

The Human high sensitivity C-reactive protein (hs-CRP) ELISA kit (Cusabio, Houston, Texas, USA) was used to analyse hsCRP levels.

#### 4.1.3 NEUROFILAMENT LIGHT PROTEIN

In paper III NfL levels were measured in CSF using a commercial Enzyme-linked immunosorbent assay (ELISA) (Uman Diagnostics, Umeå, Sweden). The age-related cut off levels for CSF NfL used were: 18–30 years:  $< 380$  ng/L; 30–39 years:  $< 560$  ng/L; 40–59 years:  $< 890$  ng/L;  $> 59$  years:  $< 1850$  ng/L.

Initially, only methods for the analysis of NfL in CSF were available, but a sensitive method for analysis in plasma has been developed.<sup>161</sup> This enabled us to use P-NfL in paper IV. P-NfL levels were analysed using an in-house Single molecule array (Simoa) method on an HD-1 Analyzer (Quanterix, Billerica, MA). The clinical age-related cut off levels for P-NfL used were: < 18 years: < 7 pg/mL; 18–50 years: < 10 pg/mL; 51–60 years: < 15 pg/mL; 61–70 years: < 20 pg/mL; and > 70 years: < 35 pg/mL.

All NfL analyses were performed at the Clinical Neurochemistry Laboratory at Sahlgrenska University Hospital, Mölndal, Sweden.

#### **4.1.4 HOMOCYSTEINE AND B VITAMINS**

In paper III P-homocysteine was measured using a stable isotope dilution liquid chromatography tandem mass spectrometry (LC-MS/MS) using a Quattro micro instrument (Waters Corporation, Milford, MA, USA). In paper IV the Roche Homocysteine Enzymatic Assay on a Cobas c501 instrument was used (Roche Diagnostics, Rotkreutz, Switzerland).

P-B<sub>12</sub> and S/P-folate (paper III and IV) were analysed on a Cobas e instrument (Roche Diagnostics, Rotkreutz, Switzerland).

## **4.2 DRUG-DRUG INTERACTIONS**

The Liverpool University HIV drug interactions database<sup>207</sup> was used for analysis of PDDIs between ART and concomitant medications. The database divides interactions into four classes: green (no interaction expected), yellow (potential interaction likely to be of weak intensity, additional monitoring or dose adjustment is unlikely to be required), orange (potential interaction that may require dose monitoring, alteration of drug dosage or timing of administration), and red (do not co-administer). If a specific comedication was not found in the Liverpool University HIV drug database the Swedish drug interactions database Janusmed<sup>208</sup> was used. PDDIs from the orange and red classes were included in the analysis in paper I.

## **4.3 NEUROCOGNITIVE TESTING**

In paper IV neurocognitive testing was performed with Cogstate (Cogstate Ltd, Melbourne, Australia). Cogstate is a computerised neuropsychological screening test that have been validated for HIV-associated neurocognitive impairment.<sup>209, 210</sup> In paper IV the testing consisted of five tasks testing five different cognitive domains: Detection (psychomotor function), Identification (attention), One card

learning (visual learning), One back test (working memory) and Groton Maze learning test (executive function). The five test results were combined to one total score (Cogstate combined z-score), which was used in the analysis.

## 4.4 STATISTICAL METHODS

Correlations were calculated using Pearson correlation coefficient (paper III & IV). Differences in continuous variables between groups were analysed using either parametric tests (independent T-sample, ANOVA) or nonparametric tests (Mann Whitney U-test, Kruskal Wallis test). In paper I differences in drug levels between groups were adjusted for time, using ANCOVA analysis. When differences were analysed within a group at different timepoints Paired T-test was used. Group comparisons of categorical variables were analysed with Chi-square test and Fisher's exact test. In paper III predictors of homocysteine and NfL were investigated with multiple linear regression analysis with forward selection.

Power analysis was performed in the cross-sectional study (paper I and II) and the randomised controlled trial (paper IV).

Variables were log-transformed when suitable to reduce skewness throughout the thesis.

A  $p$ -value  $< 0.05$  was considered significant throughout the thesis. All statistical analyses were performed using SPSS version 21, 25, or 27 (IBM SPSS Statistics, Armonk NY, USA) or Prism version 6.0, 8.0 or 9 (Graph-pad Software Inc., La Jolla, CA, USA).

## 4.5 ETHICS

All studies in this thesis were granted ethical approval by the Research Ethics Committee at Gothenburg University (paper I & II: Dnr: 642-13; paper III: Dnr: 82-88, and Ö588-10; paper IV: Dnr: 029-16). In addition, the randomised controlled trial described in paper III was approved by the national Swedish Medical Products Agency, and registered at [clinicaltrials.gov](https://clinicaltrials.gov) (NCT number: NCT02773147), and in the European Union Drug Regulation Authorities Clinical Trials Database (EudraCT number: 2015-004311-20).

All studies were performed according to the Helsinki declaration and all participants gave their written informed consent.

## HIV in the elderly

## 5 HIV IN THE ELDERLY

The aim of the cross-sectional study was to describe different aspects of HIV in the elderly. The rationale to conduct the study was a general lack of studies focusing on elderly PLHIV. The number of PLHIV  $\geq 65$  years of age is increasing, due to a higher number of survivors reaching higher ages but also incident cases in this age group. In many studies still PLHIV over the age of 50 is regarded elderly. Notably, the lack of studies in the old ages is not specific for the HIV research field.<sup>211</sup>

In our study, the term elderly was used to describe those over the age of 65 and it is in this meaning it will be used throughout this thesis. When there is no data available on elderly, data on those  $\geq 50$  years of age may be discussed.

There are specific aspects and challenges related to the care of elderly PLHIV, and one of them is testing. Older PLHIV are more often late presenters, i.e. diagnosed at a more advanced stage.<sup>212-214</sup> In many cases there has been at least one contact with healthcare prior to diagnosis where a missed opportunity for testing has occurred.<sup>214</sup> Testing is uncommon in the elderly, also in those with a higher risk of transmission.<sup>215</sup> One reason is supposedly a reluctance of healthcare providers to discuss HIV testing and sexual health with older individuals.<sup>216, 217</sup>

Older age at seroconversion is associated with an increased risk of progress to AIDS and mortality in untreated HIV infection.<sup>218, 219</sup> Also, in PLHIV on ART older age is related to poorer CD4<sup>+</sup> cell count recovery and disease progression.<sup>220-223</sup> In addition, late presenters have an increased risk of AIDS and mortality,<sup>220, 224</sup> further pointing out the importance of testing and early diagnosis in the elderly.

Another challenge is the physiologic changes related to aging, which may influence the pharmacokinetics and pharmacodynamics of drugs, described in section 1.7.2. To investigate the role of older age on drug levels we analysed the levels of EFV, ATV and DRV, at the time recommended as first line alternatives. We found significantly higher levels of DRV in the elderly compared to the controls. In addition, we also found higher ATV levels in the elderly, with a trend towards significance indicating that it might be a drug class effect. In contrast, we found no difference in EFV levels. In concert with our finding two studies reported higher concentrations of DRV in PLHIV aged  $> 60$  and  $> 65$  years respectively.<sup>225, 226</sup> To my knowledge no other study has compared levels of ATV in the elderly compared to a younger control group. Yet several studies,<sup>227, 228</sup> but

not all,<sup>229, 230</sup> have found an association between ATV concentrations and age. Studies specifically studying EFV in the elderly are also lacking, but in line with our finding studies have not found an association between EFV concentrations and age.<sup>228, 230</sup>

A recent study used mathematical modelling to calculate the effect of aging on the pharmacokinetics of 10 commonly used ART drugs. They found an increased exposure to drugs with age, but it was not considered clinically significant due to the wide therapeutic window of the study drugs.<sup>231</sup> Furthermore, the effect of age on ART pharmacokinetics was recently reviewed by Calcagno et al, who conclude that data in the elderly is scarce, and although exposure of ART may increase with age clinically relevant alterations are rare based on available data.<sup>120</sup> However, the elderly PLHIV may be more vulnerable to the specific toxicities and risks of different ART, such as the renal and bone effects associated with TDF and the possible increased risk of cardiovascular disease associated with PIs and ABC.

Supported by others,<sup>112, 232</sup> we found a higher frequency of concomitant medications and PDDIs in the elderly. PDDIs were common in the elderly in our study, with 59% having one or more amber or red flag PDDIs (data not presented in paper I). The same finding is described by others.<sup>233, 234</sup> Comparable to other studies,<sup>235</sup> we found red flag PDDIs in 8% of the elderly. All of them were related to PI use, not surprisingly since the PIs are notorious for PDDIs. At the time when the study was executed, DRV and ATV were the options at hand when a robust ART was preferred and in the presence of resistance. Today, with the arrival of dolutegravir and bictegravir as robust alternatives and with fewer drug interactions, these might be alternatives.

In our study 68% of the elderly met the criteria of polypharmacy, defined as 5 drugs or more including ART but not booster (data not reported in paper I). This is in concert with a previous finding of 70%, but somewhat lower than another study which reported 93%.<sup>233, 236</sup> Polypharmacy is associated with several disadvantageous events, e.g. increased risk of adverse events,<sup>237</sup> drug-drug interactions,<sup>237</sup> non-adherence,<sup>237, 238</sup> hospitalization,<sup>237</sup> and quality of life.<sup>239</sup> This highlights the need for proper prescription in the elderly. As a result, tools have been established to help reduce the use of inappropriate medicines, such as the screening tool of older people's prescriptions and screening tool to alert to right treatment (STOPP/START) and Beer's criteria.<sup>240, 241</sup> In our study 25% of the elderly fulfilled a STOPP criterion (data not presented in paper I). Indeed substantial, but lower than previous studies in elderly PLHIV who reported frequencies of 63% and 71%.<sup>234, 236</sup> Our result may underestimate the prevalence of STOPP criteria since the study was not designed for this and some data is lacking.

A pillar of successful treatment with virologic control and beneficial effects on morbidity and mortality, is good adherence to ART. Older PLHIV are in general more adherent than younger.<sup>242, 243</sup> Our study showed no difference in adherence between the elderly and the controls. Of note, the median age in the control groups were 43–46 and thus not very young. However, adherence may decline with more advanced age with the possible development of dementia and neurocognitive decline.<sup>242-244</sup> In addition, as described above, polypharmacy might have a deteriorating effect. Insufficient data exist on how to improve adherence in elderly PLHIV.<sup>245</sup> Nonetheless, several issues that are a concern for the elderly have been related to non-adherence, such as loneliness, poor social support, and depression.<sup>5</sup> Furthermore, pill burden and multiple daily doses have been related to poorer adherence.<sup>246</sup> On the other hand, treatment simplification with fixed-dose combinations and once-daily dosing has been related to better adherence, but notably with uncertain effect on clinical outcome.<sup>247</sup>

Elderly have an increased risk of adverse events. Although, we did not find any difference in self-reported side effects between the elderly and the controls. When analysing the elderly separately, we found a higher frequency of side effects in the DRV arm compared to ATV, and the EFV arm compared to ATV. Interestingly, we did not find a higher frequency of CNS side effects caused by EFV in the elderly, which has been reported to be related to age, comorbidities, and concomitant medications.<sup>248</sup> The use of self-reporting of side effects is both easy and may reveal side effects that are important to the patient. However, the result may have been influenced by side effects related to the ART backbone or other concomitant medications. A study focusing on side effects after the initiation of ART or after ART-switch would be interesting to investigate how side effects affect the elderly.

As described in chapter 1 PLHIV are at higher risk for comorbidities compared to HIV negative individuals. In addition, many comorbidities are related to aging. We found, as expected, a higher number of comorbidities in the elderly group compared to controls, and only 2 (2%) of the elderly had no comorbidity. In table 4 comorbidities affecting the elderly in our study are presented (data not shown in paper I). The three most common comorbidities were all related to aging; hypertension, dyslipidemia, and cardiovascular disease, in line with the findings of others.<sup>93, 94, 249</sup>

Table 4. Comorbidities in PLHIV  $\geq 65$  years of age included in our study.

Comorbidity	Number of individuals
Cardiovascular disease	20
Hypertension	47
Hyperlipidemia*	45
Diabetes mellitus	15
Malignancy <sup>+</sup>	16
Sleep disorder	13
Psychiatric disorder	11
Nervous system disease	10
Pulmonary disease	8
Chronic kidney disease	2
Inflammatory disease	1
Other	71

Other diseases with  $\geq 2$  individuals include: allergy, arthrosis, cardiac arrhythmia, chronic pain, chronic constipation, diverticulosis, eye diseases, gastro-esophageal reflux disease or gastritis, gout, haemophilia, harmful use of alcohol, hearing impairment, hyperplasia of prostate, irritable bowel syndrome, kidney stone, liver diseases, osteoporosis, polyp of colon, recurrent cystitis, recurrent herpes simplex, skin diseases, vitamin deficiencies

\*As diagnosis or on lipid lowering treatment

<sup>+</sup>current or in personal history

In addition to the increased risk of comorbidities related to natural aging, PLHIV have an increased risk independent of age. Possibly adding further to the risk, elderly PLHIV more often have been exposed to the early ART drugs with more toxicities.<sup>249, 250</sup>

As mentioned in section 1.7.3, PLHIV have, despite ART, a persisting low-level inflammation and immune activation.<sup>121-123, 126</sup> This has been proposed as one of the possible mechanisms underlying the elevated risk of comorbidities.<sup>104</sup> In our study we compared the influence of ART regimen on levels of inflammation markers in the elderly group. Among the ten analysed markers we found differences between the arms in IL-6, sICAM-1, and CXCL10 levels. The ATV group had lower IL-6 levels compared to the other arms. On the other hand, the EFV group had lower levels of sICAM-1. The DRV group had higher CXCL10 levels, but significantly different only when compared to the EFV arm. A potential possibility of mitigating the residual inflammation with the choice of ART is interesting. However, it is not possible to draw any conclusion of causality based

on these results and the clinical relevance is uncertain. Earlier studies have not focused on elderly PLHIV which reduces the comparability of the studies.<sup>251-253</sup> In addition, intervariability between methods and within the Luminex method further complicates comparison.

When analysing and evaluating data concerning elderly PLHIV, it is important to remember that the group is not homogenous. The group consist of on the one hand the survivors, who probably experienced periods of immunodeficiency and were exposed to early ART drugs with toxicities. On the other hand, the group includes those who were diagnosed at an older age and may have started ART early during the course of infection. In addition, studies performed at different times during the course of the pandemic have included untreated PLHIV and those who started ART at different times during infection due to the recommendations at the time, and often a combination of these groups. These differences need to be considered, and may limit the comparability between studies. The future will come with new aspects and the composition of the elderly group will continue to evolve over time.

When caring for the increasing number of elderly PLHIV we need to consider certain aspects that are particularly important in this group. The choice of ART regimen is a cornerstone in all HIV care, but the need of individually tailored drug regimens are particularly important in the elderly. In our choice of ART, we need to consider how to reduce the disadvantageous effects of polypharmacy and PDDIs. Also, we need to consider how to ensure adherence, including social aspects. Lastly, we need to diagnose comorbidities early and treat them correctly.

**Vitamin B  
metabolism in  
HIV infection**

## 6 VITAMIN B METABOLISM IN HIV INFECTION

An article in *Läkartidningen*<sup>254</sup> discussing and summarizing a trial of the effect of vitamin B therapy on the rate of brain shrinkage attracted our interest to the field of homocysteine, vitamin B metabolism, and neurocognition.

In 1985 Petito et al reported that the myelopathy of AIDS had a pathological resemblance to the myelopathy found in vitamin B deficiency.<sup>255</sup> Several researchers hypothesised and investigated disturbances of the methylation cycle as a possible mechanism in HIV-related neurodegeneration.<sup>256, 257</sup> Vitamin B<sub>12</sub> deficiency was reported as a common finding before the introduction of effective ART.<sup>258, 259</sup> However, studies investigating the association of B<sub>12</sub> levels and cognition yielded contradictory results.<sup>258, 260</sup> Homocysteine levels are higher in untreated PLHIV compared to HIV-negative individuals, a finding that is consistent in those on ART.<sup>261</sup>

In the retrospective study, presented in paper III, we report a novel finding of a correlation between P-homocysteine and CSF NfL in 83 individuals with untreated HIV infection. The correlation remained significant also after the initiation of ART (in a second sample of 22 individuals). The result was reproduced in paper IV where a correlation was found between P-homocysteine and P-NfL in 124 PLHIV on ART.

As described in section 1.9, elevated homocysteine is a marker of vitamin B<sub>12</sub> and/or folate deficiency. In the screening-cohort (n = 124) of the clinical trial one individual had vitamin B<sub>12</sub> deficiency and five had folate deficiency, (diagnosed as P-homocysteine > 15 µmol/L and P-B<sub>12</sub> < 140 pmol/L or P-folate < 7 nmol/L), and thus deficiency was rare. However, vitamin levels in the low-normal spectrum were frequent, 40% for P-B<sub>12</sub> (< 300) and 35% for P-folate (< 10). Furthermore, 27% had elevated homocysteine levels, suggesting that suboptimal vitamin levels may be common in this group.

From the screening cohort those with P-homocysteine ≥ 12 µmol/L were included in the randomised trial, as described in section 3.3. After 12 months of treatment with vitamin B<sub>12</sub>, folate, and B<sub>6</sub> the P-homocysteine levels had decreased, and the P-B<sub>12</sub> and P-folate levels had increased in the active treatment arm. However, this was not accompanied by a decrease in P-NfL when compared to the control group. Neither after 24 months of vitamin B supplementation a

P-NfL lowering effect was noted. This result speaks against suboptimal vitamin B<sub>12</sub> and folate as the cause of the association between homocysteine and NfL.

Both homocysteine and NfL increase with age.<sup>177, 262</sup> However, age was accounted for in the multiple regression analysis in paper III. Homocysteine inversely correlates to kidney function.<sup>177, 263, 264</sup> An earlier study of PLHIV found no correlation between kidney function measured by creatinine and P-NfL, but some later studies in PLHIV and non-HIV-infected individuals have found an association between P-NfL and creatinine.<sup>265-268</sup> However, it would not be probable that kidney function in the normal range, as is the case in paper III, would influence CSF NfL levels, speaking against kidney function as the cause of the association.

Immune activation is considered a pathogenic mechanism of HIV infection both systemically and in the CNS.<sup>50, 153, 171</sup> Despite continuous ART for several years signs of immune activation in the CNS, measured as neopterin, persist.<sup>170, 269</sup> Interestingly, in HIV-negative individuals serum neopterin levels have been associated with homocysteine levels.<sup>270</sup> In paper III we found a moderate correlation between neopterin and homocysteine in bivariate correlation but not in the multiple regression analysis, and thus did not support this as a common pathologic mechanism.

The metabolic syndrome has been connected with cognitive impairment both in the general population and in PLHIV.<sup>271-273</sup> Elevated homocysteine levels have in turn been related to the metabolic syndrome.<sup>274, 275</sup> In addition inflammation has been related to dementia and cognitive decline.<sup>276, 277</sup> As described above PLHIV have higher homocysteine levels and signs of inflammation compared to HIV negative individuals.<sup>121, 123, 261</sup> Furthermore, the metabolic syndrome is common among PLHIV, studies having reported a prevalence of 20–40%.<sup>272, 278</sup> We did not study the metabolic syndrome in the present study and hence a conclusion cannot be drawn from our data, but hypothetically the interactions of these factors may be a plausible explanation for the association between homocysteine levels and signs of neuroaxonal injury in PLHIV.



## Conclusion

## 7 CONCLUSION

We found higher steady-state concentrations of PIs in elderly PLHIV compared to younger controls. In addition, polypharmacy was common in the elderly and they had a higher risk of potential drug-drug interactions. Altogether this puts elderly PLHIV in risk of adverse drug events and warrants careful consideration when prescribing both antiviral and non-antiviral drugs.

Despite effective ART PLHIV show a persistent chronic low-grade inflammation. Elderly treated with atazanavir, darunavir, or efavirenz had different levels of IL-6, ICAM-1 and CXCL10. Further studies are needed to confirm if different ART may give rise to different inflammatory profiles.

An association was found between NfL, a marker of ongoing neuroaxonal injury, and homocysteine, a marker of vitamin B deficiency, in PLHIV with and without ART. Supplementation with B vitamins decreased homocysteine levels but had no effect on NfL levels. This suggest that suboptimal B vitamin levels are not the cause of the association. Further studies to investigate possible underlying causes are warranted.

## Future perspectives

## 8 FUTURE PERSPECTIVES

The worldwide recommendation to treat all irrespective of CD4<sup>+</sup> cell count as a result of the START study was a turning point in HIV medicine. Early treatment changes the focus from complications related directly to the virus to those of ART, comorbidities, and indirect effects of the virus.

As seen in our studies elderly PLHIV are at risk of drug related unfavorable events. There are today a couple of cohort studies focusing on older and elderly PLHIV,<sup>279</sup> but in the future, studies designed to study ART and the relationship to concomitant medications in the elderly are needed.

We need to learn how to best care for the growing number of elderly PLHIV. In the UK a combined HIV and geriatrics clinic has been developed that may lead the way.<sup>280</sup>

Still in the era of effective ART, PLHIV are at higher risk of developing comorbidities. Further studies are needed that investigate the mechanisms underlying the higher risk of comorbidities and how to mitigate them.

We still do not know all the effects of long-term ART and future studies focusing on organ dysfunction and comorbidities, are needed. Prospective studies are needed to see if residual inflammation is related to ART regimen.

Studies are needed to further elucidate the mechanisms that drive the development of and progression of HAND in those with effective ART. Our finding of a relationship between NfL and homocysteine warrants further investigation into the cause, including the role of inflammation and the metabolic syndrome in neuroaxonal injury in PLHIV.

The ever alluring goal is cure. In the history of HIV two individuals living with HIV have been medically cured. The first one was Timothy Brown, the so called “Berlin patient” who was reported to be cured of HIV in 2006. Timothy Brown received an allogeneic stem-cell transplantation due to acute myeloid leukemia with a donor who was homozygous for the CCR5Δ32 mutation. A second transplant was needed after relapse but led to complete remission of the leukemia and no virus could be detected during the follow-up period.<sup>281</sup> Timothy Brown died of leukemia in 2020. He experienced no reoccurrence of HIV during his lifetime.<sup>282</sup> The second case, Adam Castillejo, initially known as “the London patient”, was reported at the Conference on Retroviruses and Opportunistic Infections in 2019,

a conference I myself attended. As with Timothy Brown, Adam Castillejo received an allogenic stem-cell transplant with a donor homozygous for the CCR5 $\Delta$ 32 mutation due to treatment of lymphoma. He was transplanted in 2016, stopped ART in 2017 and have experienced no viral rebound.<sup>283</sup> Allogenic stem-cell transplantation is a dangerous treatment and not feasible to perform with the purpose to cure HIV but has rendered some hope and insights. Furthermore, in the case of two elite controllers the analysis of a large number of cells failed to identify any replication competent proviral DNA suggesting naturally achieved cure.<sup>284</sup>

As previously discussed, the latent reservoir constitute an obstacle to HIV cure and subsequently the reservoir has been in focus in the research for a cure. Different modes of action have been suggested and investigated such as; the “shock and kill” strategy which aims to activate latently infected cells who are then killed by cells of the immune system; inactivate or remove proviral DNA through gene-editing such as CRISPR/Cas9 ; or the “lock and block” theory that aims to hinder latently infected cells to reactivate.<sup>285, 286</sup> None of the strategies has of yet been successful but the search for a cure will continue.

Last but not least we cannot forget that a life with HIV today looks very different depending on where you live. Large inequalities in HIV care exist globally and the diagnosis is still often associated with stigma.



# Acknowledgements

## 9 ACKNOWLEDGEMENTS

Mitt varmaste tack går till alla de personer som lever med HIV som tagit sig tid för att delta i dessa studier. Utan er hade denna bit aldrig lagts till pusslet.

Till följande personer vill jag också ge ett särskilt tack:

Min huvudhandledare, Lars-Magnus Andersson. Som i slutet av 2013 frågade om jag ville göra hans HIV äldre projekt till mitt. Som sedan gett mig osvikligt stöd och hjälp längs vägen men också utrymme att prova mina egna vingar. Och vars arbete på ett inspirerande sätt alltid genomsyras av att sätta patienternas väl först.

Min bihandledare, Magnus Gisslén. Som delat med sig av sin stora kunskap och givit ständig uppmuntran. Som trots ett oändligt antal bollar i luften alltid haft tid för mina funderingar och vars dörr alltid stått öppen.

Alla medförfattare som delat med sig av sin kunskap och kloka synpunkter på mina manuskript.

Göteborgs universitet, som representeras av Johan Westin och Marie Studahl och fram tills nyligen Lars Hagberg. Som bidrar till ett öppet och utvecklande forsknings- och utbildningsklimat och ett gott samarbete mellan universitetet och sjukhuset.

Vår verksamhetschef Lars-Magnus Andersson, och vår tidigare verksamhetschef Rune Wejstål som gör infektionskliniken till en utvecklande plats att arbeta på och bidrar till ett stimulerande forskningsklimat. Aylin Yilmaz, min närmsta chef och nära kollega, som får allting att verka enkelt.

Ulrika, min omnipotenta kliniska handledare som lotsade mig igenom ST och var den som en gång anställde mig på infektionskliniken.

Helena, Mia, Kristina, Lissie och Marie, sjuksköterskor på HIV-mottagningen. För ovärderlig hjälp och stöd i omhändertagandet av patienter både i forskning och den kliniska verkligheten.

Eva Karlsson, på forskningslaboratoriet som med stenkoll och varsam hand handskats med alla prover.

Staffan Nilsson, för god hjälp när jag varit i statistisk nöd.

Alla kollegor på infektionskliniken på Östra sjukhuset som gör det värt att gå till jobbet varje dag.

Malin och Erik som i tid och otid svarat på alla mina praktiska frågor om att disputerat. Arvid för genomläsning och kloka synpunkter på avhandlingen.

Infektionskliniken på Norrlands Universitetssjukhus i Umeå, som lärde mig att bli doktor och befäste mitt intresse för infektionssjukdomar.

Thomas, Josefine, Linn, Malin, Birgitta, Gustaf och Anna mina nära vänner och IKÖ-gäng. Janne, Christoffer och Ida som lämnat IKÖ men inte gemenskapen.

Jenny, Lotta, Anna, Sofia som gjorde Göteborg till ett hem.

Jens, min första work husband.

Min västerbottniska familj, Lisa, Sara, Karin, Anna, Lene, Sonia, Fanny, Hans, Elias, Johan i vars sällskap jag blev vuxen och i vars sällskap jag ser fram emot att åldras.

Sten, Maria, Mattias, Jennie, Petter, Jenny, Vilhelm, Sigrid, Alfred, Miriam och Nore för att ni inneslutit mig i den varma gemenskap som är familjen Tyrberg.

Ida, min ständiga följeslagare. Min Sancho Panza till min Don Quixote. Min Howie till min Nick.

Mamma, Pappa, Fredrik för villkorslös kärlek, stöd och trygghet. För att jag har fått gå min egen väg även om jag i slutändan inte föll så långt ifrån päronträdet.

Farfar, som saknas mig, som under sitt liv besatt en aldrig sinande livsglädje.

Tobias, mitt hjärta. Mitt livs äventyr.



## References

## 10 REFERENCES

1. Pneumocystis pneumonia--Los Angeles. *MMWR Morb Mortal Wkly Rep.* Jun 5 1981;30(21):250-2.
2. Kaposi's sarcoma and Pneumocystis pneumonia among homosexual men--New York City and California. *MMWR Morb Mortal Wkly Rep.* Jul 3 1981;30(25):305-8.
3. Update on acquired immune deficiency syndrome (AIDS)--United States. *MMWR Morb Mortal Wkly Rep.* Sep 24 1982;31(37):507-8, 513-4.
4. Gallo RC, Montagnier L. The discovery of HIV as the cause of AIDS. *N Engl J Med.* Dec 11 2003;349(24):2283-5. doi:10.1056/NEJMp038194
5. Barré-Sinoussi F, Chermann JC, Rey F, et al. Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immune deficiency syndrome (AIDS). *Science.* May 20 1983;220(4599):868-71. doi:10.1126/science.6189183
6. Gallo RC, Sarin PS, Gelmann EP, et al. Isolation of human T-cell leukemia virus in acquired immune deficiency syndrome (AIDS). *Science.* May 20 1983;220(4599):865-7. doi:10.1126/science.6601823
7. Gallo RC, Salahuddin SZ, Popovic M, et al. Frequent detection and isolation of cytopathic retroviruses (HTLV-III) from patients with AIDS and at risk for AIDS. *Science.* May 4 1984;224(4648):500-3. doi:10.1126/science.6200936
8. Levy JA, Hoffman AD, Kramer SM, Landis JA, Shimabukuro JM, Oshiro LS. Isolation of lymphocytopathic retroviruses from San Francisco patients with AIDS. *Science.* Aug 24 1984;225(4664):840-2. doi:10.1126/science.6206563
9. Coffin J, Haase A, Levy JA, et al. What to call the AIDS virus? *Nature.* May 1-7 1986;321(6065):10. doi:10.1038/321010a0
10. Clumeck N, Mascart-Lemone F, de Maubeuge J, Brenez D, Marcelis L. Acquired immune deficiency syndrome in Black Africans. *Lancet.* Mar 19 1983;1(8325):642. doi:10.1016/s0140-6736(83)91808-1
11. Piot P, Quinn TC, Taelman H, et al. Acquired immunodeficiency syndrome in a heterosexual population in Zaire. *Lancet.* Jul 14 1984;2(8394):65-9. doi:10.1016/s0140-6736(84)90241-1
12. Serwadda D, Mugerwa RD, Sewankambo NK, et al. Slim disease: a new disease in Uganda and its association with HTLV-III infection. *Lancet.* Oct 19 1985;2(8460):849-52. doi:10.1016/s0140-6736(85)90122-9
13. Van de Perre P, Clumeck N, Carael M, et al. Female prostitutes: a risk group for infection with human T-cell lymphotropic virus type III. *Lancet.* Sep 7 1985;2(8454):524-7. doi:10.1016/s0140-6736(85)90462-3
14. Kreiss JK, Koech D, Plummer FA, et al. AIDS virus infection in Nairobi prostitutes. Spread of the epidemic to East Africa. *N Engl J Med.* Feb 13 1986;314(7):414-8. doi:10.1056/nejm198602133140704
15. Oleske J, Minnefor A, Cooper R, Jr., et al. Immune deficiency syndrome in children. *Jama.* May 6 1983;249(17):2345-9.
16. Peeters M, Honoré C, Huet T, et al. Isolation and partial characterization of an HIV-related virus occurring naturally in chimpanzees in Gabon. *Aids.* Oct 1989;3(10):625-30. doi:10.1097/00002030-198910000-00001
17. Huet T, Cheynier R, Meyerhans A, Roelants G, Wain-Hobson S. Genetic organization of a chimpanzee lentivirus related to HIV-1. *Nature.* May 24 1990;345(6273):356-9. doi:10.1038/345356a0

18. Keele BF, Van Heuverswyn F, Li Y, et al. Chimpanzee reservoirs of pandemic and nonpandemic HIV-1. *Science*. Jul 28 2006;313(5786):523-6. doi:10.1126/science.1126531
19. Sharp PM, Hahn BH. Origins of HIV and the AIDS pandemic. *Cold Spring Harb Perspect Med*. Sep 2011;1(1):a006841. doi:10.1101/cshperspect.a006841
20. Hahn BH, Shaw GM, De Cock KM, Sharp PM. AIDS as a zoonosis: scientific and public health implications. *Science*. Jan 28 2000;287(5453):607-14. doi:10.1126/science.287.5453.607
21. Zhu T, Korber BT, Nahmias AJ, Hooper E, Sharp PM, Ho DD. An African HIV-1 sequence from 1959 and implications for the origin of the epidemic. *Nature*. Feb 5 1998;391(6667):594-7. doi:10.1038/35400
22. Worobey M, Gemmel M, Teuwen DE, et al. Direct evidence of extensive diversity of HIV-1 in Kinshasa by 1960. *Nature*. Oct 2 2008;455(7213):661-4. doi:10.1038/nature07390
23. Nyamweya S, Hegedus A, Jaye A, Rowland-Jones S, Flanagan KL, Macallan DC. Comparing HIV-1 and HIV-2 infection: Lessons for viral immunopathogenesis. *Rev Med Virol*. Jul 2013;23(4):221-40. doi:10.1002/rmv.1739
24. Hirsch VM, Olmsted RA, Murphey-Corb M, Purcell RH, Johnson PR. An African primate lentivirus (SIVsm) closely related to HIV-2. *Nature*. Jun 1 1989;339(6223):389-92. doi:10.1038/339389a0
25. Fanales-Belasio E, Raimondo M, Suligoi B, Buttò S. HIV virology and pathogenetic mechanisms of infection: a brief overview. *Ann Ist Super Sanita*. 2010;46(1):5-14. doi:10.4415/ann\_10\_01\_02
26. Levy JA. The multifaceted retrovirus. *Cancer Res*. Nov 1986;46(11):5457-68.
27. Sierra S, Kupfer B, Kaiser R. Basics of the virology of HIV-1 and its replication. *J Clin Virol*. Dec 2005;34(4):233-44. doi:10.1016/j.jcv.2005.09.004
28. Adapted from “HIV-1 Genome and Structure”, by BioRender.com (2021). Retrieved from <https://app.biorender.com/biorender-templates>.
29. Bbosa N, Kaleebu P, Ssemwanga D. HIV subtype diversity worldwide. *Curr Opin HIV AIDS*. May 2019;14(3):153-160. doi:10.1097/coh.0000000000000534
30. Simon V, Ho DD, Abdool Karim Q. HIV/AIDS epidemiology, pathogenesis, prevention, and treatment. *Lancet*. Aug 5 2006;368(9534):489-504. doi:10.1016/s0140-6736(06)69157-5
31. Levy JA. HIV pathogenesis: 25 years of progress and persistent challenges. *Aids*. Jan 14 2009;23(2):147-60. doi:10.1097/QAD.0b013e3283217f9f
32. Hladik F, McElrath MJ. Setting the stage: host invasion by HIV. *Nat Rev Immunol*. Jun 2008;8(6):447-57. doi:10.1038/nri2302
33. Berger EA, Murphy PM, Farber JM. *Chemokine receptors as HIV-1 coreceptors: Roles in viral entry, tropism, and disease*. 1999.
34. Shen Q, Wu C, Freniere C, Tripler TN, Xiong Y. Nuclear import of hiv-1. Review. *Viruses*. 2021;13(11)2242. doi:10.3390/v13112242
35. Arhel N. Revisiting HIV-1 uncoating. *Retrovirology*. Nov 17 2010;7:96. doi:10.1186/1742-4690-7-96
36. Burdick RC, Li C, Munshi M, et al. HIV-1 uncoats in the nucleus near sites of integration. *Proc Natl Acad Sci U S A*. Mar 10 2020;117(10):5486-5493. doi:10.1073/pnas.1920631117

37. Zila V, Margiotta E, Turoňová B, et al. Cone-shaped HIV-1 capsids are transported through intact nuclear pores. *Cell*. Feb 18 2021;184(4):1032-1046.e18. doi:10.1016/j.cell.2021.01.025
38. Freed EO. HIV-1 assembly, release and maturation. *Nat Rev Microbiol*. Aug 2015;13(8):484-96. doi:10.1038/nrmicro3490
39. Adapted from “HIV Replication Cycle”, by BioRender.com (2021). Retrieved from <https://app.biorender.com/biorender-templates>.
40. Zhuang J, Jetzt AE, Sun G, et al. Human immunodeficiency virus type 1 recombination: rate, fidelity, and putative hot spots. *J Virol*. Nov 2002;76(22):11273-82. doi:10.1128/jvi.76.22.11273-11282.2002
41. Berger EA, Doms RW, Fenyo EM, et al. A new classification for HIV-1 [6]. Letter. *Nature*. 1998;391(6664):240. doi:10.1038/34571
42. Pope M, Haase AT. Transmission, acute HIV-1 infection and the quest for strategies to prevent infection. *Nat Med*. Jul 2003;9(7):847-52. doi:10.1038/nm0703-847
43. Liu R, Paxton WA, Choe S, et al. Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell*. Aug 9 1996;86(3):367-77. doi:10.1016/s0092-8674(00)80110-5
44. Connor RI, Sheridan KE, Ceradini D, Choe S, Landau NR. Change in coreceptor use correlates with disease progression in HIV-1-infected individuals. *J Exp Med*. Feb 17 1997;185(4):621-8. doi:10.1084/jem.185.4.621
45. Moir S, Chun TW, Fauci AS. Pathogenic mechanisms of HIV disease. *Annu Rev Pathol*. 2011;6:223-48. doi:10.1146/annurev-pathol-011110-130254
46. Tindall B, Barker S, Donovan B, et al. Characterization of the Acute Clinical Illness Associated With Human Immunodeficiency Virus Infection. Article. *Archives of Internal Medicine*. 1988;148(4):945-949. doi:10.1001/archinte.1988.00380040185026
47. Borrow P, Lewicki H, Hahn BH, Shaw GM, Oldstone MBA. Virus-specific CD8+ cytotoxic T-lymphocyte activity associated with control of viremia in primary human immunodeficiency virus type 1 infection. Note. *Journal of Virology*. 1994;68(9):6103-6110. doi:10.1128/jvi.68.9.6103-6110.1994
48. Mellors JW, Rinaldo CR, Jr., Gupta P, White RM, Todd JA, Kingsley LA. Prognosis in HIV-1 infection predicted by the quantity of virus in plasma. *Science*. May 24 1996;272(5265):1167-70. doi:10.1126/science.272.5265.1167
49. Deeks SG, Walker BD. Human immunodeficiency virus controllers: mechanisms of durable virus control in the absence of antiretroviral therapy. *Immunity*. Sep 2007;27(3):406-16. doi:10.1016/j.immuni.2007.08.010
50. Sodora DL, Silvestri G. Immune activation and AIDS pathogenesis. *Aids*. Feb 19 2008;22(4):439-46. doi:10.1097/QAD.0b013e3282f2db7
51. Created with BioRender.com.
52. Fischl MA, Richman DD, Grieco MH, et al. The efficacy of azidothymidine (AZT) in the treatment of patients with AIDS and AIDS-related complex. A double-blind, placebo-controlled trial. *N Engl J Med*. Jul 23 1987;317(4):185-91. doi:10.1056/nejm198707233170401
53. Volberding PA, Lagakos SW, Grimes JM, et al. The duration of zidovudine benefit in persons with asymptomatic HIV infection. Prolonged evaluation of protocol 019 of the AIDS Clinical Trials Group. *Jama*. Aug 10 1994;272(6):437-42.

54. Hammer SM, Squires KE, Hughes MD, et al. A controlled trial of two nucleoside analogues plus indinavir in persons with human immunodeficiency virus infection and CD4 cell counts of 200 per cubic millimeter or less. AIDS Clinical Trials Group 320 Study Team. *N Engl J Med.* Sep 11 1997;337(11):725-33. doi:10.1056/nejm199709113371101
55. Palella FJ, Jr., Delaney KM, Moorman AC, et al. Declining morbidity and mortality among patients with advanced human immunodeficiency virus infection. HIV Outpatient Study Investigators. *N Engl J Med.* Mar 26 1998;338(13):853-60. doi:10.1056/nejm199803263381301
56. Gulick RM, Mellors JW, Havlir D, et al. Treatment with indinavir, zidovudine, and lamivudine in adults with human immunodeficiency virus infection and prior antiretroviral therapy. *N Engl J Med.* Sep 11 1997;337(11):734-9. doi:10.1056/nejm199709113371102
57. Perelson AS, Essunger P, Cao Y, et al. Decay characteristics of HIV-1-infected compartments during combination therapy. *Nature.* May 8 1997;387(6629):188-91. doi:10.1038/387188a0
58. Finzi D, Blankson J, Siliciano JD, et al. Latent infection of CD4+ T cells provides a mechanism for lifelong persistence of HIV-1, even in patients on effective combination therapy. *Nat Med.* May 1999;5(5):512-7. doi:10.1038/8394
59. Friis-Møller N, Sabin CA, Weber R, et al. Combination antiretroviral therapy and the risk of myocardial infarction. *N Engl J Med.* Nov 20 2003;349(21):1993-2003. doi:10.1056/NEJMoa030218
60. Carr A, Cooper DA. Adverse effects of antiretroviral therapy. *Lancet.* Oct 21 2000;356(9239):1423-30. doi:10.1016/s0140-6736(00)02854-3
61. Hirschel B. Planned interruptions of anti-HIV treatment. *Lancet Infect Dis.* Aug 2001;1(1):53-9. doi:10.1016/s1473-3099(01)00022-6
62. El-Sadr WM, Lundgren J, Neaton JD, et al. CD4+ count-guided interruption of antiretroviral treatment. *N Engl J Med.* Nov 30 2006;355(22):2283-96. doi:10.1056/NEJMoa062360
63. Lundgren JD, Babiker AG, Gordin F, et al. Initiation of Antiretroviral Therapy in Early Asymptomatic HIV Infection. *N Engl J Med.* Aug 27 2015;373(9):795-807. doi:10.1056/NEJMoa1506816
64. Adapted from “HIV Sites for Therapeutic Intervention”, by BioRender.com (2021). Retrieved from <https://app.biorender.com/biorender-templates>.
65. Quinn TC, Wawer MJ, Sewankambo N, et al. Viral load and heterosexual transmission of human immunodeficiency virus type 1. Rakai Project Study Group. *N Engl J Med.* Mar 30 2000;342(13):921-9. doi:10.1056/nejm200003303421303
66. Connor EM, Sperling RS, Gelber R, et al. Reduction of maternal-infant transmission of human immunodeficiency virus type 1 with zidovudine treatment. Pediatric AIDS Clinical Trials Group Protocol 076 Study Group. *N Engl J Med.* Nov 3 1994;331(18):1173-80. doi:10.1056/nejm199411033311801
67. Donnell D, Baeten JM, Kiarie J, et al. Heterosexual HIV-1 transmission after initiation of antiretroviral therapy: a prospective cohort analysis. *Lancet.* Jun 12 2010;375(9731):2092-8. doi:10.1016/s0140-6736(10)60705-2
68. Cohen MS, Chen YQ, McCauley M, et al. Antiretroviral Therapy for the Prevention of HIV-1 Transmission. *N Engl J Med.* Sep 1 2016;375(9):830-9. doi:10.1056/NEJMoa1600693

69. Cohen MS, Chen YQ, McCauley M, et al. Prevention of HIV-1 infection with early antiretroviral therapy. *N Engl J Med*. Aug 11 2011;365(6):493-505. doi:10.1056/NEJMoa1105243
70. Rodger AJ, Cambiano V, Bruun T, et al. Sexual Activity Without Condoms and Risk of HIV Transmission in Serodifferent Couples When the HIV-Positive Partner Is Using Suppressive Antiretroviral Therapy. *Jama*. Jul 12 2016;316(2):171-81. doi:10.1001/jama.2016.5148
71. Bavinton BR, Pinto AN, Phanuphak N, et al. Viral suppression and HIV transmission in serodiscordant male couples: an international, prospective, observational, cohort study. *Lancet HIV*. Aug 2018;5(8):e438-e447. doi:10.1016/s2352-3018(18)30132-2
72. Rodger AJ, Cambiano V, Bruun T, et al. Risk of HIV transmission through condomless sex in serodifferent gay couples with the HIV-positive partner taking suppressive antiretroviral therapy (PARTNER): final results of a multicentre, prospective, observational study. *Lancet*. Jun 15 2019;393(10189):2428-2438. doi:10.1016/s0140-6736(19)30418-0
73. Grant RM, Lama JR, Anderson PL, et al. Preexposure chemoprophylaxis for HIV prevention in men who have sex with men. *N Engl J Med*. Dec 30 2010;363(27):2587-99. doi:10.1056/NEJMoa1011205
74. Baeten JM, Donnell D, Ndase P, et al. Antiretroviral prophylaxis for HIV prevention in heterosexual men and women. *N Engl J Med*. Aug 2 2012;367(5):399-410. doi:10.1056/NEJMoa1108524
75. Molina JM, Capitant C, Spire B, et al. On-Demand Preexposure Prophylaxis in Men at High Risk for HIV-1 Infection. *N Engl J Med*. Dec 3 2015;373(23):2237-46. doi:10.1056/NEJMoa1506273
76. Chun TW, Finzi D, Margolick J, Chadwick K, Schwartz D, Siliciano RF. In vivo fate of HIV-1-infected T cells: quantitative analysis of the transition to stable latency. *Nat Med*. Dec 1995;1(12):1284-90. doi:10.1038/nm1295-1284
77. Siliciano RF, Greene WC. HIV latency. *Cold Spring Harb Perspect Med*. Sep 2011;1(1):a007096. doi:10.1101/cshperspect.a007096
78. Chun TW, Stuyver L, Mizell SB, et al. Presence of an inducible HIV-1 latent reservoir during highly active antiretroviral therapy. *Proc Natl Acad Sci U S A*. Nov 25 1997;94(24):13193-7. doi:10.1073/pnas.94.24.13193
79. Wong JK, Hezareh M, Günthard HF, et al. Recovery of replication-competent HIV despite prolonged suppression of plasma viremia. *Science*. Nov 14 1997;278(5341):1291-5. doi:10.1126/science.278.5341.1291
80. Finzi D, Hermankova M, Pierson T, et al. Identification of a reservoir for HIV-1 in patients on highly active antiretroviral therapy. *Science*. Nov 14 1997;278(5341):1295-300. doi:10.1126/science.278.5341.1295
81. Dufour C, Gantner P, Fromentin R, Chomont N. The multifaceted nature of HIV latency. *J Clin Invest*. Jul 1 2020;130(7):3381-3390. doi:10.1172/jci136227
82. Chun TW, Carruth L, Finzi D, et al. Quantification of latent tissue reservoirs and total body viral load in HIV-1 infection. *Nature*. May 8 1997;387(6629):183-8. doi:10.1038/387183a0
83. Siliciano JD, Kajdas J, Finzi D, et al. Long-term follow-up studies confirm the stability of the latent reservoir for HIV-1 in resting CD4+ T cells. *Nat Med*. Jun 2003;9(6):727-8. doi:10.1038/nm880

84. UNAIDS/WHO estimates. Updated July 2021. .  
[https://cdn.who.int/media/docs/default-source/hq-hiv-hepatitis-and-stis-library/2021\\_global\\_summary\\_web\\_v32.pdf?sfvrsn=4b8815ad\\_37](https://cdn.who.int/media/docs/default-source/hq-hiv-hepatitis-and-stis-library/2021_global_summary_web_v32.pdf?sfvrsn=4b8815ad_37). Accessed Dec 22nd 2021
85. Centers for Disease Control and Prevention. HIV Surveillance Report, 2019; vol. 32. <http://www.cdc.gov/hiv/library/reports/hiv-surveillance.html>. Published May 2021. Accessed November 19 2021.
86. Smit M, Brinkman K, Geerlings S, et al. Future challenges for clinical care of an ageing population infected with HIV: a modelling study. *Lancet Infect Dis*. Jul 2015;15(7):810-8. doi:10.1016/s1473-3099(15)00056-0
87. Sabin CA. Do people with HIV infection have a normal life expectancy in the era of combination antiretroviral therapy? *BMC Med*. Nov 27 2013;11:251. doi:10.1186/1741-7015-11-251
88. Gueler A, Moser A, Calmy A, et al. Life expectancy in HIV-positive persons in Switzerland: matched comparison with general population. *Aids*. Jan 28 2017;31(3):427-436. doi:10.1097/qad.0000000000001335
89. Lohse N, Obel N. Update of Survival for Persons With HIV Infection in Denmark. *Ann Intern Med*. Nov 15 2016;165(10):749-750. doi:10.7326/116-0091
90. Teeraananchai S, Kerr SJ, Amin J, Ruxrungtham K, Law MG. Life expectancy of HIV-positive people after starting combination antiretroviral therapy: a meta-analysis. *HIV Med*. Apr 2017;18(4):256-266. doi:10.1111/hiv.12421
91. Jani C, Patel K, Walker A, et al. Trends of HIV Mortality between 2001 and 2018: An Observational Analysis. *Trop Med Infect Dis*. Sep 24 2021;6(4)doi:10.3390/tropicalmed6040173
92. Hasse B, Ledergerber B, Furrer H, et al. Morbidity and aging in HIV-infected persons: the Swiss HIV cohort study. *Clin Infect Dis*. Dec 2011;53(11):1130-9. doi:10.1093/cid/cir626
93. Demontès M, Duvernay SE, Allavena C, et al. Multimorbidity in elderly persons according to the year of diagnosis of human immunodeficiency virus infection: A cross-sectional Dat'AIDS cohort study. Article. *Clinical Infectious Diseases*. 2020;71(11):2880-2888. doi:10.1093/cid/ciz1171
94. Kong AM, Pozen A, Anastos K, Kelvin EA, Nash D. Non-HIV Comorbid Conditions and Polypharmacy Among People Living with HIV Age 65 or Older Compared with HIV-Negative Individuals Age 65 or Older in the United States: A Retrospective Claims-Based Analysis. *AIDS Patient Care STDS*. Mar 2019;33(3):93-103. doi:10.1089/apc.2018.0190
95. Guaraldi G, Orlando G, Zona S, et al. Premature age-related comorbidities among HIV-infected persons compared with the general population. *Clin Infect Dis*. Dec 2011;53(11):1120-6. doi:10.1093/cid/cir627
96. Schouten J, Wit FW, Stolte IG, et al. Cross-sectional comparison of the prevalence of age-associated comorbidities and their risk factors between HIV-infected and uninfected individuals: the AGEhIV cohort study. *Clin Infect Dis*. Dec 15 2014;59(12):1787-97. doi:10.1093/cid/ciu701
97. Pourcher V, Gourmelen J, Bureau I, Boue S. Comorbidities in people living with HIV: An epidemiologic and economic analysis using a claims database in France. Article. *PLoS ONE*. 2020;15(12 December):e0243529. doi:10.1371/journal.pone.0243529

98. Jespersen NA, Axelsen F, Dollerup J, Nørgaard M, Larsen CS. The burden of non-communicable diseases and mortality in people living with HIV (PLHIV) in the pre-, early- and late-HAART era. *HIV Med.* Jul 2021;22(6):478-490. doi:10.1111/hiv.13077
99. Freiberg MS, Chang CC, Kuller LH, et al. HIV infection and the risk of acute myocardial infarction. *JAMA Intern Med.* Apr 22 2013;173(8):614-22. doi:10.1001/jamainternmed.2013.3728
100. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. *J Clin Endocrinol Metab.* Jul 2007;92(7):2506-12. doi:10.1210/jc.2006-2190
101. Paisible AL, Chang CC, So-Armah KA, et al. HIV infection, cardiovascular disease risk factor profile, and risk for acute myocardial infarction. *J Acquir Immune Defic Syndr.* Feb 1 2015;68(2):209-16. doi:10.1097/qai.0000000000000419
102. Brown TT, Qaqish RB. Antiretroviral therapy and the prevalence of osteopenia and osteoporosis: a meta-analytic review. *Aids.* Nov 14 2006;20(17):2165-74. doi:10.1097/QAD.0b013e32801022eb
103. Bedimo RJ, McGinnis KA, Dunlap M, Rodriguez-Barradas MC, Justice AC. Incidence of non-AIDS-defining malignancies in HIV-infected versus noninfected patients in the HAART era: impact of immunosuppression. *J Acquir Immune Defic Syndr.* Oct 1 2009;52(2):203-8. doi:10.1097/QAI.0b013e3181b033ab
104. Deeks SG, Tracy R, Douek DC. Systemic effects of inflammation on health during chronic HIV infection. *Immunity.* Oct 17 2013;39(4):633-45. doi:10.1016/j.immuni.2013.10.001
105. Frazier EL, Sutton MY, Brooks JT, Shouse RL, Weiser J. Trends in cigarette smoking among adults with HIV compared with the general adult population, United States - 2009-2014. *Prev Med.* Jun 2018;111:231-234. doi:10.1016/j.ypmed.2018.03.007
106. Johnston PI, Wright SW, Orr M, et al. Worldwide relative smoking prevalence among people living with and without HIV. *Aids.* May 1 2021;35(6):957-970. doi:10.1097/qad.0000000000002815
107. Vos AG, Venter WDF. Cardiovascular toxicity of contemporary antiretroviral therapy. *Curr Opin HIV AIDS.* Nov 1 2021;16(6):286-291. doi:10.1097/coh.0000000000000702
108. Sabin CA, Worm SW, Weber R, et al. Use of nucleoside reverse transcriptase inhibitors and risk of myocardial infarction in HIV-infected patients enrolled in the D:A:D study: A multi-cohort collaboration. Article. *The Lancet.* 2008;371(9622):1417-1426. doi:10.1016/S0140-6736(08)60423-7
109. Tenorio AR, Zheng Y, Bosch RJ, et al. Soluble markers of inflammation and coagulation but not T-cell activation predict non-AIDS-defining morbid events during suppressive antiretroviral treatment. *J Infect Dis.* Oct 15 2014;210(8):1248-59. doi:10.1093/infdis/jiu254
110. Tien PC, Choi AI, Zolopa AR, et al. Inflammation and mortality in HIV-infected adults: analysis of the FRAM study cohort. *J Acquir Immune Defic Syndr.* Nov 2010;55(3):316-22. doi:10.1097/QAI.0b013e3181e66216
111. Kuller LH, Tracy R, Belloso W, et al. Inflammatory and coagulation biomarkers and mortality in patients with HIV infection. *PLoS Med.* Oct 21 2008;5(10):e203. doi:10.1371/journal.pmed.0050203

112. Tseng A, Szadkowski L, Walmsley S, Salit I, Raboud J. Association of age with polypharmacy and risk of drug interactions with antiretroviral medications in HIV-positive patients. *Ann Pharmacother.* Nov 2013;47(11):1429-39. doi:10.1177/1060028013504075
113. López-Centeno B, Badenes-Olmedo C, Mataix-Sanjuan Á, et al. Polypharmacy and Drug-Drug Interactions in People Living With Human Immunodeficiency Virus in the Region of Madrid, Spain: A Population-Based Study. *Clin Infect Dis.* Jul 11 2020;71(2):353-362. doi:10.1093/cid/ciz811
114. *EACS Guidelines 11.0.* 2021. Accessed December 27, 2021. [https://www.eacsociety.org/media/final2021eacsguidelinesv11.0\\_oct2021.pdf](https://www.eacsociety.org/media/final2021eacsguidelinesv11.0_oct2021.pdf)
115. Panel on Antiretroviral Guidelines for Adults and Adolescents. *Guidelines for the Use of Antiretroviral Agents in Adults and Adolescents with HIV.* Department of Health and Human Services. Accessed December 27, 2021. <https://clinicalinfo.hiv.gov/sites/default/files/guidelines/documents/AdultandAdolescentGL.pdf>
116. *BHIVA guidelines for the routine investigation and monitoring of adult HIV-1-positive individuals 2016 (2019 interim update).* Accessed December 27, 2021. <https://www.bhiva.org/monitoring-guidelines>
117. Richterman A, Sax PE. Antiretroviral therapy in older people with HIV. *Curr Opin HIV AIDS.* Mar 2020;15(2):118-125. doi:10.1097/coh.0000000000000614
118. Mangoni AA, Jackson SH. Age-related changes in pharmacokinetics and pharmacodynamics: basic principles and practical applications. *Br J Clin Pharmacol.* Jan 2004;57(1):6-14. doi:10.1046/j.1365-2125.2003.02007.x
119. Stader F, Siccardi M, Battegay M, Kinzig H, Penny MA, Marzolini C. Repository Describing an Aging Population to Inform Physiologically Based Pharmacokinetic Models Considering Anatomical, Physiological, and Biological Age-Dependent Changes. *Clin Pharmacokinet.* Apr 2019;58(4):483-501. doi:10.1007/s40262-018-0709-7
120. Calcagno A, Trunfio M, D'Avolio A, Di Perri G, Bonora S. The impact of age on antiretroviral drug pharmacokinetics in the treatment of adults living with HIV. *Expert Opin Drug Metab Toxicol.* Jun 2021;17(6):665-676. doi:10.1080/17425255.2021.1915285
121. Bastard JP, Fellahi S, Couffignal C, et al. Increased systemic immune activation and inflammatory profile of long-term HIV-infected ART-controlled patients is related to personal factors, but not to markers of HIV infection severity. *J Antimicrob Chemother.* 2015;70(6):1816-24. doi:10.1093/jac/dkv036
122. French MA, King MS, Tschampa JM, da Silva BA, Landay AL. Serum immune activation markers are persistently increased in patients with HIV infection after 6 years of antiretroviral therapy despite suppression of viral replication and reconstitution of CD4+ T cells. *J Infect Dis.* Oct 15 2009;200(8):1212-5. doi:10.1086/605890
123. Neuhaus J, Jacobs DR, Jr., Baker JV, et al. Markers of inflammation, coagulation, and renal function are elevated in adults with HIV infection. *J Infect Dis.* Jun 15 2010;201(12):1788-95. doi:10.1086/652749
124. Gabuzda D, Jamieson BD, Collman RG, et al. Pathogenesis of Aging and Age-related Comorbidities in People with HIV: Highlights from the HIV ACTION Workshop. *Pathog Immun.* 2020;5(1):143-174. doi:10.20411/pai.v5i1.365

125. Hattab S, Guihot A, Guiguet M, et al. Comparative impact of antiretroviral drugs on markers of inflammation and immune activation during the first two years of effective therapy for HIV-1 infection: an observational study. *BMC Infect Dis*. Mar 4 2014;14:122. doi:10.1186/1471-2334-14-122
126. Wada NI, Jacobson LP, Margolick JB, et al. The effect of HAART-induced HIV suppression on circulating markers of inflammation and immune activation. *Aids*. Feb 20 2015;29(4):463-71. doi:10.1097/qad.0000000000000545
127. Yu X, Shang H, Jiang Y. ICAM-1 in HIV infection and underlying mechanisms. *Cytokine*. Jan 2020;125:154830. doi:10.1016/j.cyto.2019.154830
128. Sproston NR, Ashworth JJ. Role of C-Reactive Protein at Sites of Inflammation and Infection. *Front Immunol*. 2018;9:754. doi:10.3389/fimmu.2018.00754
129. Borges Á H, O'Connor JL, Phillips AN, et al. Factors Associated With Plasma IL-6 Levels During HIV Infection. *J Infect Dis*. Aug 15 2015;212(4):585-95. doi:10.1093/infdis/jiv123
130. Lei J, Yin X, Shang H, Jiang Y. IP-10 is highly involved in HIV infection. *Cytokine*. Mar 2019;115:97-103. doi:10.1016/j.cyto.2018.11.018
131. Liu M, Guo S, Hibbert JM, et al. CXCL10/IP-10 in infectious diseases pathogenesis and potential therapeutic implications. *Cytokine Growth Factor Rev*. Jun 2011;22(3):121-30. doi:10.1016/j.cytogfr.2011.06.001
132. Burdo TH, Lentz MR, Autissier P, et al. Soluble CD163 made by monocyte/macrophages is a novel marker of HIV activity in early and chronic infection prior to and after anti-retroviral therapy. *J Infect Dis*. Jul 1 2011;204(1):154-63. doi:10.1093/infdis/jir214
133. Steele AK, Lee EJ, Vestal B, et al. Contribution of intestinal barrier damage, microbial translocation and HIV-1 infection status to an inflammaging signature. *PLoS One*. 2014;9(5):e97171. doi:10.1371/journal.pone.0097171
134. Mehta AK, Gracias DT, Croft M. TNF activity and T cells. *Cytokine*. Jan 2018;101:14-18. doi:10.1016/j.cyto.2016.08.003
135. Skiles JW, Gonnella NC, Jeng AY. The design, structure, and therapeutic application of matrix metalloproteinase inhibitors. *Curr Med Chem*. Mar 2001;8(4):425-74. doi:10.2174/0929867013373417
136. Jostock T, Müllberg J, Ozbek S, et al. Soluble gp130 is the natural inhibitor of soluble interleukin-6 receptor transsignaling responses. *Eur J Biochem*. Jan 2001;268(1):160-7. doi:10.1046/j.1432-1327.2001.01867.x
137. Snider WD, Simpson DM, Nielsen S, Gold JW, Metroka CE, Posner JB. Neurological complications of acquired immune deficiency syndrome: analysis of 50 patients. *Ann Neurol*. Oct 1983;14(4):403-18. doi:10.1002/ana.410140404
138. Navia BA, Cho ES, Petito CK, Price RW. The AIDS dementia complex: II. Neuropathology. *Ann Neurol*. Jun 1986;19(6):525-35. doi:10.1002/ana.410190603
139. Navia BA, Jordan BD, Price RW. The AIDS dementia complex: I. Clinical features. *Ann Neurol*. Jun 1986;19(6):517-24. doi:10.1002/ana.410190602
140. McArthur JC, Hoover DR, Bacellar H, et al. Dementia in AIDS patients: Incidence and risk factors. Article. *Neurology*. 1993;43(11):2245-2252. doi:10.1212/wnl.43.11.2245
141. Portegies P, de Gans J, Lange JM, et al. Declining incidence of AIDS dementia complex after introduction of zidovudine treatment. *Bmj*. Sep 30 1989;299(6703):819-21. doi:10.1136/bmj.299.6703.819

142. Levy RM, Bredesen DE, Rosenblum ML. Neurological manifestations of the acquired immunodeficiency syndrome (AIDS): Experience at UCSF and review of the literature. Review. *Journal of Neurosurgery*. 1985;62(4):475-495. doi:10.3171/jns.1985.62.4.0475
143. d'Arminio Monforte A, Cinque P, Mocroft A, et al. Changing incidence of central nervous system diseases in the EuroSIDA cohort. *Ann Neurol*. Mar 2004;55(3):320-8. doi:10.1002/ana.10827
144. Heaton RK, Franklin DR, Ellis RJ, et al. HIV-associated neurocognitive disorders before and during the era of combination antiretroviral therapy: differences in rates, nature, and predictors. *J Neurovirol*. Feb 2011;17(1):3-16. doi:10.1007/s13365-010-0006-1
145. Antinori A, Arendt G, Becker JT, et al. Updated research nosology for HIV-associated neurocognitive disorders. *Neurology*. Oct 30 2007;69(18):1789-99. doi:10.1212/WNL.0000287431.88658.8b
146. Heaton RK, Clifford DB, Franklin DR, Jr., et al. HIV-associated neurocognitive disorders persist in the era of potent antiretroviral therapy: CHARTER Study. *Neurology*. Dec 7 2010;75(23):2087-96. doi:10.1212/WNL.0b013e318200d727
147. Sacktor N, Skolasky RL, Seaberg E, et al. Prevalence of HIV-associated neurocognitive disorders in the Multicenter AIDS Cohort Study. Article. *Neurology*. 2016;86(4):334-340. doi:10.1212/WNL.0000000000002277
148. Rourke SB, Bekele T, Rachlis A, et al. Asymptomatic neurocognitive impairment is a risk for symptomatic decline over a 3-year study period. *Aids*. Jan 1 2021;35(1):63-72. doi:10.1097/qad.0000000000002709
149. Gisslén M, Price RW, Nilsson S. The definition of HIV-associated neurocognitive disorders: are we overestimating the real prevalence? *BMC Infect Dis*. Dec 28 2011;11:356. doi:10.1186/1471-2334-11-356
150. Nightingale S, Dreyer AJ, Saylor D, Gisslén M, Winston A, Joska JA. Moving on from HAND: why we need new criteria for cognitive impairment in people with HIV and a proposed way forward. *Clin Infect Dis*. Apr 27 2021;doi:10.1093/cid/ciab366
151. Davis LE, Hjelle BL, Miller VE, et al. Early viral brain invasion in iatrogenic human immunodeficiency virus infection. Article. *Neurology*. 1992;42(9):1736-1739. doi:10.1212/wnl.42.9.1736
152. Valcour V, Chalermchai T, Sailasuta N, et al. Central nervous system viral invasion and inflammation during acute HIV infection. Article. *Journal of Infectious Diseases*. 2012;206(2):275-282. doi:10.1093/infdis/jis326
153. Spudich S, González-Scarano F. HIV-1-related central nervous system disease: Current issues in pathogenesis, diagnosis, and treatment. Article. *Cold Spring Harbor Perspectives in Medicine*. 2012;2(6)a007120. doi:10.1101/cshperspect.a007120
154. González-Scarano F, Martín-García J. The neuropathogenesis of AIDS. Review. *Nature Reviews Immunology*. 2005;5(1):69-81. doi:10.1038/nri1527
155. Haase AT. Pathogenesis of lentivirus infections. *Nature*. Jul 10-16 1986;322(6075):130-6. doi:10.1038/322130a0
156. Hoffman PN, Cleveland DW, Griffin JW, Landes PW, Cowan NJ, Price DL. Neurofilament gene expression: a major determinant of axonal caliber. *Proc Natl Acad Sci U S A*. May 1987;84(10):3472-6. doi:10.1073/pnas.84.10.3472

157. Gafson AR, Barthélemy NR, Bomont P, et al. Neurofilaments: neurobiological foundations for biomarker applications. *Brain*. Jul 1 2020;143(7):1975-1998. doi:10.1093/brain/awaa098
158. Rosengren LE, Karlsson JE, Karlsson JO, Persson LI, Wikkelso C. Patients with amyotrophic lateral sclerosis and other neurodegenerative diseases have increased levels of neurofilament protein in CSF. *J Neurochem*. Nov 1996;67(5):2013-8. doi:10.1046/j.1471-4159.1996.67052013.x
159. Norgren N, Rosengren L, Stigbrand T. Elevated neurofilament levels in neurological diseases. *Brain Res*. Oct 10 2003;987(1):25-31. doi:10.1016/s0006-8993(03)03219-0
160. Khalil M, Teunissen CE, Otto M, et al. Neurofilaments as biomarkers in neurological disorders. *Nat Rev Neurol*. Oct 2018;14(10):577-589. doi:10.1038/s41582-018-0058-z
161. Gisslén M, Price RW, Andreasson U, et al. Plasma Concentration of the Neurofilament Light Protein (NFL) is a Biomarker of CNS Injury in HIV Infection: A Cross-Sectional Study. *EBioMedicine*. Jan 2016;3:135-140. doi:10.1016/j.ebiom.2015.11.036
162. Millere E, Rots D, Simrén J, et al. Plasma neurofilament light chain as a potential biomarker in Charcot-Marie-Tooth disease. *Eur J Neurol*. Mar 2021;28(3):974-981. doi:10.1111/ene.14689
163. Trojanowski JQ, Walkenstein N, Lee VM. Expression of neurofilament subunits in neurons of the central and peripheral nervous system: an immunohistochemical study with monoclonal antibodies. *J Neurosci*. Mar 1986;6(3):650-60. doi:10.1523/jneurosci.06-03-00650.1986
164. Hagberg L, Fuchs D, Rosengren L, Gisslén M. Intrathecal immune activation is associated with cerebrospinal fluid markers of neuronal destruction in AIDS patients. *J Neuroimmunol*. Jan 3 2000;102(1):51-5. doi:10.1016/s0165-5728(99)00150-2
165. Abdulle S, Mellgren A, Brew BJ, et al. CSF neurofilament protein (NFL) -- a marker of active HIV-related neurodegeneration. *J Neurol*. Aug 2007;254(8):1026-32. doi:10.1007/s00415-006-0481-8
166. Mellgren A, Price RW, Hagberg L, Rosengren L, Brew BJ, Gisslén M. Antiretroviral treatment reduces increased CSF neurofilament protein (NFL) in HIV-1 infection. *Neurology*. Oct 9 2007;69(15):1536-41. doi:10.1212/01.wnl.0000277635.05973.55
167. Peterson J, Gisslen M, Zetterberg H, et al. Cerebrospinal fluid (CSF) neuronal biomarkers across the spectrum of HIV infection: hierarchy of injury and detection. *PLoS One*. 2014;9(12):e116081. doi:10.1371/journal.pone.0116081
168. Gisslen M, Hagberg L, Brew BJ, Cinque P, Price RW, Rosengren L. Elevated cerebrospinal fluid neurofilament light protein concentrations predict the development of AIDS dementia complex. *J Infect Dis*. Jun 15 2007;195(12):1774-8. doi:10.1086/518043
169. Jessen Krut J, Mellberg T, Price RW, et al. Biomarker evidence of axonal injury in neuroasymptomatic HIV-1 patients. *PLoS One*. 2014;9(2):e88591. doi:10.1371/journal.pone.0088591
170. Ulfhammer G, Edén A, Mellgren Å, et al. Persistent central nervous system immune activation following more than 10 years of effective HIV antiretroviral

- treatment. *Aids*. Sep 24 2018;32(15):2171-2178. doi:10.1097/qad.0000000000001950
171. Hagberg L, Cinque P, Gisslen M, et al. Cerebrospinal fluid neopterin: an informative biomarker of central nervous system immune activation in HIV-1 infection. *AIDS Res Ther*. Jun 3 2010;7:15. doi:10.1186/1742-6405-7-15
172. McCaddon A. Homocysteine and cognition—a historical perspective. *J Alzheimers Dis*. Aug 2006;9(4):361-80. doi:10.3233/jad-2006-9402
173. Hoffbrand AV, Weir DG. The history of folic acid. *Br J Haematol*. Jun 2001;113(3):579-89. doi:10.1046/j.1365-2141.2001.02822.x
174. Smith AD, Refsum H. Homocysteine - from disease biomarker to disease prevention. *J Intern Med*. Oct 2021;290(4):826-854. doi:10.1111/joim.13279
175. Selhub J. Homocysteine metabolism. *Annu Rev Nutr*. 1999;19:217-46. doi:10.1146/annurev.nutr.19.1.217
176. Kim J, Kim H, Roh H, Kwon Y. Causes of hyperhomocysteinemia and its pathological significance. *Arch Pharm Res*. Apr 2018;41(4):372-383. doi:10.1007/s12272-018-1016-4
177. Refsum H, Smith AD, Ueland PM, et al. Facts and recommendations about total homocysteine determinations: an expert opinion. *Clin Chem*. Jan 2004;50(1):3-32. doi:10.1373/clinchem.2003.021634
178. Green R, Allen LH, Bjørke-Monsen AL, et al. Vitamin B(12) deficiency. *Nat Rev Dis Primers*. Jun 29 2017;3:17040. doi:10.1038/nrdp.2017.40
179. Smith AD, Refsum H. Do we need to reconsider the desirable blood level of vitamin B12? *J Intern Med*. Feb 2012;271(2):179-82. doi:10.1111/j.1365-2796.2011.02485.x
180. Carmel R. Biomarkers of cobalamin (vitamin B-12) status in the epidemiologic setting: a critical overview of context, applications, and performance characteristics of cobalamin, methylmalonic acid, and holotranscobalamin II. *Am J Clin Nutr*. Jul 2011;94(1):348s-358s. doi:10.3945/ajcn.111.013441
181. Selhub J, Jacques PF, Dallal G, Choumenkovitch S, Rogers G. The use of blood concentrations of vitamins and their respective functional indicators to define folate and vitamin B12 status. *Food Nutr Bull*. Jun 2008;29(2 Suppl):S67-73. doi:10.1177/15648265080292s110
182. Stabler SP. Vitamin B12 deficiency. *N Engl J Med*. May 23 2013;368(21):2041-2. doi:10.1056/NEJMc1304350
183. Obeid R, Heil SG, Verhoeven MMA, van den Heuvel E, de Groot L, Eussen S. Vitamin B12 Intake From Animal Foods, Biomarkers, and Health Aspects. *Front Nutr*. 2019;6:93. doi:10.3389/fnut.2019.00093
184. Allen LH, Miller JW, De Groot L, et al. Biomarkers of Nutrition for Development (BOND): Vitamin B-12 Review. Review. *Journal of Nutrition*. 2018;148:1995S-2027S. doi:10.1093/jn/nxy201
185. Heaton EB, Savage DG, Brust JC, Garrett TJ, Lindenbaum J. Neurologic aspects of cobalamin deficiency. *Medicine (Baltimore)*. Jul 1991;70(4):229-45. doi:10.1097/00005792-199107000-00001
186. Lindenbaum J, Heaton EB, Savage DG, et al. Neuropsychiatric disorders caused by cobalamin deficiency in the absence of anemia or macrocytosis. *N Engl J Med*. Jun 30 1988;318(26):1720-8. doi:10.1056/nejm198806303182604
187. Warzyszyńska JE, Kim Y-IJ. Folate in Human Health and Disease. *eLS*. 2014.

188. West AA, Caudill MA, Bailey LB. Chapter 14 - Folate. In: Marriott BP, Birt DF, Stallings VA, Yates AA, eds. *Present Knowledge in Nutrition (Eleventh Edition)*. Academic Press; 2020:239-255.
189. Carmel R. Folate metabolism. In: Carmel R, ed. *Homocysteine in Health and Disease*. Cambridge University Press; 2001:113-134.
190. Reynolds EH. The neurology of folic acid deficiency. *Handb Clin Neurol*. 2014;120:927-43. doi:10.1016/b978-0-7020-4087-0.00061-9
191. Smith AD. The worldwide challenge of the dementias: a role for B vitamins and homocysteine? *Food Nutr Bull*. Jun 2008;29(2 Suppl):S143-72. doi:10.1177/15648265080292s119
192. Seshadri S, Beiser A, Selhub J, et al. Plasma homocysteine as a risk factor for dementia and Alzheimer's disease. *N Engl J Med*. Feb 14 2002;346(7):476-83. doi:10.1056/NEJMoa011613
193. Zylberstein DE, Lissner L, Björkelund C, et al. Midlife homocysteine and late-life dementia in women. A prospective population study. *Neurobiol Aging*. Mar 2011;32(3):380-6. doi:10.1016/j.neurobiolaging.2009.02.024
194. Xu W, Tan L, Wang HF, et al. Meta-analysis of modifiable risk factors for Alzheimer's disease. *J Neurol Neurosurg Psychiatry*. Dec 2015;86(12):1299-306. doi:10.1136/jnnp-2015-310548
195. Nie T, Lu T, Xie L, Huang P, Lu Y, Jiang M. Hyperhomocysteinemia and risk of cognitive decline: a meta-analysis of prospective cohort studies. *Eur Neurol*. 2014;72(3-4):241-8. doi:10.1159/000363054
196. McCaddon A, Hudson P, Davies G, Hughes A, Williams JH, Wilkinson C. Homocysteine and cognitive decline in healthy elderly. *Dement Geriatr Cogn Disord*. Sep-Oct 2001;12(5):309-13. doi:10.1159/000051275
197. Dufouil C, Alperovitch A, Ducros V, Tzourio C. Homocysteine, white matter hyperintensities, and cognition in healthy elderly people. *Ann Neurol*. Feb 2003;53(2):214-21. doi:10.1002/ana.10440
198. Smith AD, Refsum H. Homocysteine, B Vitamins, and Cognitive Impairment. *Annu Rev Nutr*. Jul 17 2016;36:211-39. doi:10.1146/annurev-nutr-071715-050947
199. Smith AD, Refsum H, Bottiglieri T, et al. Homocysteine and Dementia: An International Consensus Statement. *J Alzheimers Dis*. 2018;62(2):561-570. doi:10.3233/JAD-171042
200. Durga J, van Boxtel MP, Schouten EG, et al. Effect of 3-year folic acid supplementation on cognitive function in older adults in the FACIT trial: a randomised, double blind, controlled trial. *Lancet*. Jan 20 2007;369(9557):208-16. doi:10.1016/S0140-6736(07)60109-3
201. de Jager CA, Oulhaj A, Jacoby R, Refsum H, Smith AD. Cognitive and clinical outcomes of homocysteine-lowering B-vitamin treatment in mild cognitive impairment: a randomized controlled trial. *Int J Geriatr Psychiatry*. Jun 2012;27(6):592-600. doi:10.1002/gps.2758
202. McMahon JA, Green TJ, Skeaff CM, Knight RG, Mann JI, Williams SM. A controlled trial of homocysteine lowering and cognitive performance. *N Engl J Med*. Jun 29 2006;354(26):2764-72. doi:10.1056/NEJMoa054025
203. Hankey GJ, Ford AH, Yi Q, et al. Effect of B vitamins and lowering homocysteine on cognitive impairment in patients with previous stroke or transient ischemic attack: a prespecified secondary analysis of a randomized,

- placebo-controlled trial and meta-analysis. *Stroke*. Aug 2013;44(8):2232-9. doi:10.1161/strokeaha.113.001886
204. Ford AH, Almeida OP. Effect of Vitamin B Supplementation on Cognitive Function in the Elderly: A Systematic Review and Meta-Analysis. *Drugs Aging*. May 2019;36(5):419-434. doi:10.1007/s40266-019-00649-w
205. Clarke R, Bennett D, Parish S, et al. Effects of homocysteine lowering with B vitamins on cognitive aging: meta-analysis of 11 trials with cognitive data on 22,000 individuals. *Am J Clin Nutr*. Aug 2014;100(2):657-66. doi:10.3945/ajcn.113.076349
206. McCaddon A, Miller JW. Assessing the association between homocysteine and cognition: reflections on Bradford Hill, meta-analyses, and causality. *Nutr Rev*. Oct 2015;73(10):723-35. doi:10.1093/nutrit/nuv022
207. University of Liverpool. HIV drug interactions. [www.hiv-druginteractions.org](http://www.hiv-druginteractions.org) Last review June 27 2019. Accessed June 27 2019.
208. Region Stockholm. Janusmed. Version 2.3.8. [www.janusmed.sll.se](http://www.janusmed.sll.se) Accessed 26 June 2019
209. Bloch M, Kamminga J, Jayewardene A, et al. A Screening Strategy for HIV-Associated Neurocognitive Disorders That Accurately Identifies Patients Requiring Neurological Review. *Clin Infect Dis*. Sep 1 2016;63(5):687-693. doi:10.1093/cid/ciw399
210. Cysique LA, Maruff P, Darby D, Brew BJ. The assessment of cognitive function in advanced HIV-1 infection and AIDS dementia complex using a new computerised cognitive test battery. *Arch Clin Neuropsychol*. Feb 2006;21(2):185-94. doi:10.1016/j.acn.2005.07.011
211. Watts G. Why the exclusion of older people from clinical research must stop. *Bmj*. May 21 2012;344:e3445. doi:10.1136/bmj.e3445
212. Mothe B, Perez I, Domingo P, et al. HIV-1 infection in subjects older than 70: A multicenter cross-sectional assessment in Catalonia, Spain. Article. *Current HIV Research*. 2009;7(6):597-600. doi:10.2174/157016209789973691
213. Mensforth S, Goodall L, Bodasing N, Coultas J. Late diagnosis among our ageing HIV population: a cohort study. *J Int AIDS Soc*. 2014;17(4 Suppl 3):19692. doi:10.7448/ias.17.4.19692
214. van den Bogaart L, Ranzani A, Oreni L, et al. Overlooked cases of HIV infection: An Italian tale of missed diagnostic opportunities. Article. *European Journal of Internal Medicine*. 2020;73:30-35. doi:10.1016/j.ejim.2019.09.006
215. Oraka E, Mason S, Xia M. Too old to test? Prevalence and correlates of HIV testing among sexually active older adults. *J Gerontol Soc Work*. May-Jun 2018;61(4):460-470. doi:10.1080/01634372.2018.1454565
216. Arya M, Patel S, Kumar D, et al. Why Physicians Don't Ask: Interpersonal and Intrapersonal Barriers to HIV Testing-Making a Case for a Patient-Initiated Campaign. *J Int Assoc Provid AIDS Care*. Jul 2016;15(4):306-12. doi:10.1177/2325957414557268
217. Tillman JL, Mark HD. HIV and STI testing in older adults: an integrative review. *J Clin Nurs*. Aug 2015;24(15-16):2074-95. doi:10.1111/jocn.12797
218. Time from HIV-1 seroconversion to AIDS and death before widespread use of highly-active antiretroviral therapy: a collaborative re-analysis. Collaborative Group on AIDS Incubation and HIV Survival including the CASCADE EU

- Concerted Action. Concerted Action on SeroConversion to AIDS and Death in Europe. *Lancet*. Apr 1 2000;355(9210):1131-7.
219. Babiker AG, Peto T, Porter K, Walker AS, Darbyshire JH. Age as a determinant of survival in HIV infection. *J Clin Epidemiol*. Dec 2001;54 Suppl 1:S16-21. doi:10.1016/s0895-4356(01)00456-5
220. Egger M, May M, Chêne G, et al. Prognosis of HIV-1-infected patients starting highly active antiretroviral therapy: a collaborative analysis of prospective studies. *Lancet*. Jul 13 2002;360(9327):119-29. doi:10.1016/s0140-6736(02)09411-4
221. Nogueras M, Navarro G, Antón E, et al. Epidemiological and clinical features, response to HAART, and survival in HIV-infected patients diagnosed at the age of 50 or more. *BMC Infect Dis*. Nov 6 2006;6:159. doi:10.1186/1471-2334-6-159
222. Sabin CA, Smith CJ, d'Arminio Monforte A, et al. Response to combination antiretroviral therapy: variation by age. *Aids*. Jul 31 2008;22(12):1463-73. doi:10.1097/QAD.0b013e3282f88d02
223. Manfredi R, Chiodo F. A case-control study of virological and immunological effects of highly active antiretroviral therapy in HIV-infected patients with advanced age. *Aids*. Jul 7 2000;14(10):1475-7. doi:10.1097/00002030-200007070-00034
224. Smith RD, Delpech VC, Brown AE, Rice BD. HIV transmission and high rates of late diagnoses among adults aged 50 years and over. *Aids*. Aug 24 2010;24(13):2109-15. doi:10.1097/QAD.0b013e32833c7b9c
225. Calza L, Colangeli V, Magistrelli E, et al. Plasma trough concentrations of darunavir/ritonavir and raltegravir in older patients with HIV-1 infection. *HIV Med*. Aug 2017;18(7):474-481. doi:10.1111/hiv.12478
226. Courlet P, Stader F, Guidi M, et al. Pharmacokinetic profiles of boosted darunavir, dolutegravir and lamivudine in aging people living with HIV. *Aids*. Jan 1 2020;34(1):103-108. doi:10.1097/qad.0000000000002372
227. Avihingsanon A, Kerr SJ, Punyawudho B, et al. Short communication: Aging not gender is associated with high atazanavir plasma concentrations in Asian HIV-infected patients. *AIDS Res Hum Retroviruses*. Dec 2013;29(12):1541-6. doi:10.1089/aid.2013.0069
228. Winston A, Jose S, Gibbons S, et al. Effects of age on antiretroviral plasma drug concentration in HIV-infected subjects undergoing routine therapeutic drug monitoring. *J Antimicrob Chemother*. Jun 2013;68(6):1354-9. doi:10.1093/jac/dkt029
229. Luz AJ, Poeta J, Linden R, Antunes MV, Caminha LI, Sprinz E. Related factors to atazanavir plasma levels in a cohort of HIV positive individuals with undetectable viral load. *Braz J Infect Dis*. Nov-Dec 2013;17(6):657-60. doi:10.1016/j.bjid.2013.04.002
230. Dumond JB, Adams JL, Prince HM, et al. Pharmacokinetics of two common antiretroviral regimens in older HIV-infected patients: a pilot study. *HIV Med*. Aug 2013;14(7):401-9. doi:10.1111/hiv.12017
231. Stader F, Courlet P, Kinvig H, et al. Effect of ageing on antiretroviral drug pharmacokinetics using clinical data combined with modelling and simulation. *Br J Clin Pharmacol*. Feb 2021;87(2):458-470. doi:10.1111/bcp.14402
232. Courlet P, Livio F, Guidi M, et al. Polypharmacy, Drug-Drug Interactions, and Inappropriate Drugs: New Challenges in the Aging Population With HIV. *Open Forum Infect Dis*. Dec 2019;6(12):ofz531. doi:10.1093/ofid/ofz531

233. Bastida C, Grau A, Márquez M, et al. Polypharmacy and potential drug-drug interactions in an HIV-infected elderly population. *Farm Hosp.* Sep 1 2017;41(5):618-624. Polifarmacia e interacciones farmacológicas potenciales en una población envejecida con infección por el VIH. doi:10.7399/fh.10778
234. Loste C, Moltó J, Pérez-Álvarez N, et al. Potential prescribing issues among older HIV-infected subjects in a Mediterranean cohort: Does the current prevalence give cause for concern? *Br J Clin Pharmacol.* Mar 2021;87(3):1310-1317. doi:10.1111/bcp.14513
235. Ruellan AL, Bourneau-Martin D, Joyau C, et al. Assessment of drug-drug interaction in an elderly human immunodeficiency virus population: Comparison of 3 expert databases. *Br J Clin Pharmacol.* Mar 2021;87(3):1194-1202. doi:10.1111/bcp.14491
236. Vinuesa-Hernando JM, Gimeno-Gracia M, Malo S, et al. Potentially inappropriate prescriptions and therapeutic complexity in older HIV patients with comorbidities. *Int J Clin Pharm.* Oct 2021;43(5):1245-1250. doi:10.1007/s11096-021-01242-1
237. Maher RL, Hanlon J, Hajjar ER. Clinical consequences of polypharmacy in elderly. *Expert Opin Drug Saf.* Jan 2014;13(1):57-65. doi:10.1517/14740338.2013.827660
238. Cantudo-Cuenca MR, Jiménez-Galán R, Almeida-Gonzalez CV, Morillo-Verdugo R. Concurrent use of comedications reduces adherence to antiretroviral therapy among HIV-infected patients. *J Manag Care Spec Pharm.* Aug 2014;20(8):844-50. doi:10.18553/jmcp.2014.20.8.844
239. Okoli C, de Los Rios P, Eremin A, Brough G, Young B, Short D. Relationship Between Polypharmacy and Quality of Life Among People in 24 Countries Living With HIV. *Prev Chronic Dis.* Mar 5 2020;17:E22. doi:10.5888/pcd17.190359
240. O'Mahony D, O'Sullivan D, Byrne S, O'Connor MN, Ryan C, Gallagher P. STOPP/START criteria for potentially inappropriate prescribing in older people: version 2. *Age Ageing.* Mar 2015;44(2):213-8. doi:10.1093/ageing/afu145
241. American Geriatrics Society 2019 Updated AGS Beers Criteria® for Potentially Inappropriate Medication Use in Older Adults. *J Am Geriatr Soc.* Apr 2019;67(4):674-694. doi:10.1111/jgs.15767
242. Barclay TR, Hinkin CH, Castellon SA, et al. Age-associated predictors of medication adherence in HIV-positive adults: health beliefs, self-efficacy, and neurocognitive status. *Health Psychol.* Jan 2007;26(1):40-9. doi:10.1037/0278-6133.26.1.40
243. Hinkin CH, Hardy DJ, Mason KI, et al. Medication adherence in HIV-infected adults: effect of patient age, cognitive status, and substance abuse. *Aids.* Jan 1 2004;18 Suppl 1(Suppl 1):S19-25. doi:10.1097/00002030-200418001-00004
244. Caballero J, Ownby RL, Jacobs RJ, Thomas JE, Schweizer MS. Association Between Cognitive Tests and Antiretroviral Medication Adherence in Older Adults With HIV. *Ann Pharmacother.* Feb 2019;53(2):151-158. doi:10.1177/1060028018798327
245. Mann SC, Castillo-Mancilla JR. HIV, aging, and adherence: an update and future directions. *Curr Opin HIV AIDS.* Mar 2020;15(2):134-141. doi:10.1097/coh.0000000000000615

246. O'Connor JL, Gardner EM, Mannheimer SB, et al. Factors associated with adherence amongst 5295 people receiving antiretroviral therapy as part of an international trial. *J Infect Dis.* Jul 2013;208(1):40-9. doi:10.1093/infdis/jis731
247. Elnaem MH, Irwan NA, Abubakar U, Syed Sulaiman SA, Elrggal ME, Cheema E. Impact of Medication Regimen Simplification on Medication Adherence and Clinical Outcomes in Patients with Long-Term Medical Conditions. *Patient Prefer Adherence.* 2020;14:2135-2145. doi:10.2147/ppa.S268499
248. Muche EA, Kiflu M, Ayalew MB. Patient Reported Central Nervous System Adverse Events of Efavirenz-Based Antiretroviral Therapy in People Living with HIV in Northwest Ethiopia. *HIV AIDS (Auckl).* 2020;12:601-609. doi:10.2147/hiv.S276111
249. Allavena C, Hanf M, Rey D, et al. Antiretroviral exposure and comorbidities in an aging HIV-infected population: The challenge of geriatric patients. *PLoS One.* 2018;13(9):e0203895. doi:10.1371/journal.pone.0203895
250. van Zoest RA, Wit FW, Kooij KW, et al. Higher Prevalence of Hypertension in HIV-1-Infected Patients on Combination Antiretroviral Therapy Is Associated With Changes in Body Composition and Prior Stavudine Exposure. *Clin Infect Dis.* Jul 15 2016;63(2):205-13. doi:10.1093/cid/ciw285
251. Kelesidis T, Tran TT, Stein JH, et al. Changes in Inflammation and Immune Activation With Atazanavir-, Raltegravir-, Darunavir-Based Initial Antiviral Therapy: ACTG 5260s. *Clin Infect Dis.* Aug 15 2015;61(4):651-60. doi:10.1093/cid/civ327
252. McComsey GA, Kitch D, Daar ES, et al. Inflammation markers after randomization to abacavir/lamivudine or tenofovir/emtricitabine with efavirenz or atazanavir/ritonavir. *Aids.* Jul 17 2012;26(11):1371-85. doi:10.1097/QAD.0b013e328354f4fb
253. Maggi P, Bellacosa C, Leone A, et al. Cardiovascular risk in advanced naïve HIV-infected patients starting antiretroviral therapy: Comparison of three different regimens - PREVALEAT II cohort. Article. *Atherosclerosis.* 2017;263:398-404. doi:10.1016/j.atherosclerosis.2017.05.004
254. Lökk J. B-vitaminer kan prövas vid kognitiv svikt. *Läkartidningen* 2013;2013;110:CDEY
255. Petito CK, Navia BA, Cho ES, Jordan BD, George DC, Price RW. Vacuolar myelopathy pathologically resembling subacute combined degeneration in patients with the acquired immunodeficiency syndrome. *N Engl J Med.* Apr 4 1985;312(14):874-9. doi:10.1056/nejm198504043121402
256. Tan SV, Guiloff RJ. Hypothesis on the pathogenesis of vacuolar myelopathy, dementia, and peripheral neuropathy in AIDS. *J Neurol Neurosurg Psychiatry.* Jul 1998;65(1):23-8. doi:10.1136/jnnp.65.1.23
257. McCaddon A, Regland B, Fear CF. Trypsin inhibition: a potential cause of cobalamin deficiency common to the pathogenesis of Alzheimer-type dementia and AIDS dementia complex? *Med Hypotheses.* Aug 1995;45(2):200-4. doi:10.1016/0306-9877(95)90069-1
258. Beach RS, Morgan R, Wilkie F, et al. Plasma vitamin B12 level as a potential cofactor in studies of human immunodeficiency virus type 1-related cognitive changes. *Archives of neurology.* May 1992;49(5):501-6. doi:10.1001/archneur.1992.00530290089016

259. Kiebertz KD, Giang DW, Schiffer RB, Vakil N. Abnormal vitamin B12 metabolism in human immunodeficiency virus infection. Association with neurological dysfunction. *Archives of neurology*. Mar 1991;48(3):312-4. doi:10.1001/archneur.1991.00530150082023
260. Robertson KR, Stern RA, Hall CD, et al. Vitamin B12 deficiency and nervous system disease in HIV infection. *Archives of neurology*. Aug 1993;50(8):807-11. doi:10.1001/archneur.1993.00540080018007
261. Deminice R, Silva TC, de Oliveira VH. Elevated homocysteine levels in human immunodeficiency virus-infected patients under antiretroviral therapy: A meta-analysis. *World J Virol*. May 12 2015;4(2):147-55. doi:10.5501/wjv.v4.i2.147
262. Yilmaz A, Blennow K, Hagberg L, et al. Neurofilament light chain protein as a marker of neuronal injury: review of its use in HIV-1 infection and reference values for HIV-negative controls. *Expert Rev Mol Diagn*. Aug 2017;17(8):761-770. doi:10.1080/14737159.2017.1341313
263. Brattström L, Lindgren A, Israelsson B, Andersson A, Hultberg B. Homocysteine and cysteine: determinants of plasma levels in middle-aged and elderly subjects. Article. *Journal of Internal Medicine*. 1994;236(6):633-641. doi:10.1111/j.1365-2796.1994.tb00856.x
264. Lewerin C, Ljungman S, Nilsson-Ehle H. Glomerular filtration rate as measured by serum cystatin C is an important determinant of plasma homocysteine and serum methylmalonic acid in the elderly. *J Intern Med*. Jan 2007;261(1):65-73. doi:10.1111/j.1365-2796.2006.01732.x
265. Alagaratnam J, De Francesco D, Zetterberg H, et al. Correlation between cerebrospinal fluid and plasma neurofilament light protein in treated HIV infection: results from the COBRA study. *J Neurovirol*. Dec 7 2021;doi:10.1007/s13365-021-01026-3
266. Korley FK, Goldstick J, Mastali M, et al. Serum NfL (Neurofilament Light Chain) Levels and Incident Stroke in Adults With Diabetes Mellitus. *Stroke*. Jul 2019;50(7):1669-1675. doi:10.1161/strokeaha.119.024941
267. Akamine S, Marutani N, Kanayama D, et al. Renal function is associated with blood neurofilament light chain level in older adults. *Sci Rep*. Nov 23 2020;10(1):20350. doi:10.1038/s41598-020-76990-7
268. Hermansson L, Yilmaz A, Axelsson M, et al. Cerebrospinal fluid levels of glial marker YKL-40 strongly associated with axonal injury in HIV infection. *J Neuroinflammation*. Jan 24 2019;16(1):16. doi:10.1186/s12974-019-1404-9
269. Yilmaz A, Yiannoutsos CT, Fuchs D, et al. Cerebrospinal fluid neopterin decay characteristics after initiation of antiretroviral therapy. *J Neuroinflammation*. May 10 2013;10:62. doi:10.1186/1742-2094-10-62
270. Schroecksnadel K, Frick B, Winkler C, Leblhuber F, Wirleitner B, Fuchs D. Hyperhomocysteinemia and immune activation. *Clin Chem Lab Med*. Nov 2003;41(11):1438-43. doi:10.1515/cclm.2003.221
271. Yaffe K, Kanaya A, Lindquist K, et al. The metabolic syndrome, inflammation, and risk of cognitive decline. Article. *Journal of the American Medical Association*. 2004;292(18):2237-2242. doi:10.1001/jama.292.18.2237
272. Yu B, Pasipanodya E, Montoya JL, et al. Metabolic Syndrome and Neurocognitive Deficits in HIV Infection. *J Acquir Immune Defic Syndr*. May 1 2019;81(1):95-101. doi:10.1097/qai.0000000000001964

273. Yates KF, Sweat V, Yau PL, Turchiano MM, Convit A. Impact of metabolic syndrome on cognition and brain: a selected review of the literature. *Arterioscler Thromb Vasc Biol.* Sep 2012;32(9):2060-7. doi:10.1161/atvbaha.112.252759
274. Catena C, Colussi G, Nait F, Capobianco F, Sechi LA. Elevated Homocysteine Levels Are Associated With the Metabolic Syndrome and Cardiovascular Events in Hypertensive Patients. *Am J Hypertens.* Jul 2015;28(7):943-50. doi:10.1093/ajh/hpu248
275. Hajer GR, van der Graaf Y, Olijhoek JK, Verhaar MC, Visseren FL. Levels of homocysteine are increased in metabolic syndrome patients but are not associated with an increased cardiovascular risk, in contrast to patients without the metabolic syndrome. *Heart.* Feb 2007;93(2):216-20. doi:10.1136/hrt.2006.093971
276. Shen XN, Niu LD, Wang YJ, et al. Inflammatory markers in Alzheimer's disease and mild cognitive impairment: a meta-analysis and systematic review of 170 studies. *J Neurol Neurosurg Psychiatry.* May 2019;90(5):590-598. doi:10.1136/jnnp-2018-319148
277. Su C, Zhao K, Xia H, Xu Y. Peripheral inflammatory biomarkers in Alzheimer's disease and mild cognitive impairment: a systematic review and meta-analysis. *Psychogeriatrics.* Jul 2019;19(4):300-309. doi:10.1111/psyg.12403
278. Bonfanti P, Giannattasio C, Ricci E, et al. HIV and metabolic syndrome: A comparison with the general population. Article. *Journal of Acquired Immune Deficiency Syndromes.* 2007;45(4):426-431. doi:10.1097/QAI.0b013e318074ef83
279. Milic J, Russwurm M, Cerezales Calvino A, Brañas F, Sánchez-Conde M, Guaraldi G. European cohorts of older HIV adults: POPPY, AGE(h)IV, GEPPPO, COBRA and FUNCFRAIL. *Eur Geriatr Med.* Apr 2019;10(2):247-257. doi:10.1007/s41999-019-00170-8
280. Levett T, Alford K, Roberts J, Adler Z, Wright J, Vera JH. Evaluation of a Combined HIV and Geriatrics Clinic for Older People Living with HIV: The Silver Clinic in Brighton, UK. *Geriatrics (Basel).* Oct 19 2020;5(4)doi:10.3390/geriatrics5040081
281. Hütter G, Nowak D, Mossner M, et al. Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. *N Engl J Med.* Feb 12 2009;360(7):692-8. doi:10.1056/NEJMoa0802905
282. Watts G. Timothy Ray Brown. *The Lancet.* 2020;396(10259):1327. doi:10.1016/S0140-6736(20)32151-6
283. Gupta RK, Peppas D, Hill AL, et al. Evidence for HIV-1 cure after CCR5Δ32/Δ32 allogeneic haemopoietic stem-cell transplantation 30 months post analytical treatment interruption: a case report. *Lancet HIV.* May 2020;7(5):e340-e347. doi:10.1016/s2352-3018(20)30069-2
284. Turk G, Seiger K, Lian X, et al. A Possible Sterilizing Cure of HIV-1 Infection Without Stem Cell Transplantation. *Ann Intern Med.* Nov 16 2021;doi:10.7326/121-0297
285. Vansant G, Bruggemans A, Janssens J, Debyser Z. Block-And-Lock Strategies to Cure HIV Infection. *Viruses.* Jan 10 2020;12(1)doi:10.3390/v12010084
286. Castro-Gonzalez S, Colomer-Lluch M, Serra-Moreno R. Barriers for HIV Cure: The Latent Reservoir. *AIDS Res Hum Retroviruses.* Sep 2018;34(9):739-759. doi:10.1089/aid.2018.0118