# Investigation of the pathophysiology of progression in multiple sclerosis

Studies on cerebrospinal fluid biomarkers

Markus Axelsson

Department of Clinical Neuroscience and Rehabilitation Institute of Neuroscience and Physiology The Sahlgrenska Academy



### UNIVERSITY OF GOTHENBURG

Gothenburg 2013

Cover illustration: Reprinted with permission from Wolters Kluwer Health. From: Trapp BD, Ransohoff R, Rudick R. Axonal pathology in multiple sclerosis: relationship to neurologic disability; Curr Opin Neurol 1999 Jun 12(3):295-302.

Investigation of the pathophysiology of progression in multiple sclerosis – Studies on cerebrospinal fluid biomarkers

© Markus Axelsson 2013 markus.axelsson@neuro.gu.se ISBN 978-91-628-8654-7 http://hdl.handle.net/2077/32007 Printed in Gothenburg, Sweden 2013 Ale Tryckteam AB, Bohus

We work in the dark – we do what we can – we give what we have. Our doubt is our passion and our passion is our task. The rest is the madness of art.

Henry James (1843-1916)

# Abstract

Multiple Sclerosis (MS) is considered an autoimmune disease of the central nervous system (CNS). It usually starts with a relapsing remitting (RR) course that eventually transforms into progressive (P)MS, showing neurodegenerative features. The pathogenesis behind the transition from RRMS to PMS is essentially unknown.

The aim of this thesis was to investigate if biomarkers in the cerebrospinal fluid (CSF) of MS patients could provide new insights into the pathophysiology of MS progression, and if biomarker levels could reflect disease activity, disability progression, or therapeutic efficacy.

Three study designs were established. The first was cross sectional and comprised MS patients, healthy controls (HC) and control subjects with another inflammatory disease. The second used a long-term follow-up setting in which RRMS, PMS and HC were assessed twice 8-10 years apart. The third used immunomodulatory or immunosuppressive intervention (natalizumab, mitoxantrone or rituximab) and assessed MS patients pre- and 12-24 months post-treatment. CSF biomarkers were analyzed for i) axonal damage (neurofilament light, NFL), ii) astrogliosis (glial fibrillary acidic protein, GFAP), iii) amyloid precursor protein metabolism (BACE1 activity, and sAPP/A $\beta$  metabolites) iv) B-cell regulation (CXCL13) and v) intrathecal IgG synthesis (IgG index, oligoclonal IgG bands (OCB)).

Increased mean GFAP levels were found in all courses of MS with the highest levels in PMS, whereas the mean NFL level of this MS population was not different from that of HC (Paper I). At long-term follow-up GFAP levels correlated with disability and had prognostic value. In contrast, increased NFL levels were found in another MS population compared to HC (Paper IV). This discrepancy might be explained by differences in disease activities between the investigated populations and due to improved sensitivity of the NFL immunoassay. We found signs of downregulation of BACE1 activity (Paper II) and sAPP/A $\beta$  metabolism (Paper III) in MS. The levels of sAPP/A $\beta$  in MS were generally decreased compared to HC suggestive of impaired neuronal function in MS. Mass spectrometry studies indicated that the sAPP/A $\beta$  metabolism was changed in PMS compared to HC by formation of other decomposition products.

We demonstrated, in opposite to the general view, changed number and pattern of OCB in CSF over time, which correlated to CXCL13 levels (Paper V). Natalizumab treatment increased sAPP A $\beta$  metabolites towards HC levels. Immunosuppressive treatment (mitoxantrone, rituximab) reduced NFL and CXCL13 in PMS. Interestingly, significantly lower NFL levels were found prior to immunosuppression in PMS patients previously treated with interferon beta or glatiramer acetate, suggest-

ing an impact on axonal damage also with first line MS therapies. Immunosuppressive treatment did not influence the number or pattern of OCB (Paper V).

In conclusion, our studies present evidence that increased immune activity plays a critical role in PMS for axonal damage and seemed to influence sAPP/A $\beta$  metabolism. In PMS, the reduced NFL level following immunosuppressive treatment clearly supports a relationship between CNS inflammation and neurodegeneration. Biomarkers in CSF provide unique information about the pathophysiology in PMS, and may serve as complement to clinical and MRI measures for assessment of disease activity, progression, severity and therapeutic efficacy.

**Key words:** multiple sclerosis, cerebrospinal fluid, biomarker, disease progression, NFL, GFAP, CXCL13, BACE1, sAPP/Aβ, IgG, oligoclonal IgG bands, IgG index

# Populärvetenskaplig sammanfattning

Multipel Skleros (MS) är en av de vanligaste orsakerna till neurologiskt handikapp bland unga vuxna i västvärlden. I Sverige lever 17 500 personer med sjukdomen. Minst två tredjedelar av de drabbade är kvinnor. MS är en autoimmun sjukdom som drabbar det centrala nervsystemet (CNS). Vanligtvis debuterar sjukdomen med relapsing remitting (RR) MS där återkommande försämringsepisoder (skov) följs av hel eller delvis återhämtning (remission). Efter i genomsnitt 15-20 år övergår sjukdomen i sekundär progressiv (SP) MS där en gradvis försämring inträder. Vid MS föreligger inflammatoriska cellinfiltrat i avgränsade områden (plaque) i hjärna och ryggmärg. Där skadas nervfibrernas isolering (myelin), myelinbildande celler (oligodendrocyter), stödjeceller (astrocyter) och nervcellernas utskott (axon). I SPMS sjunker eller upphör skovfrekvensen och den neuro-axonala förstörelsen är utbredd med förtvining (atrofi) av både vit och grå substans i CNS. Man vet idag att det är förstörelsen av axon som framförallt orsakar sjukdomssymptomen och den progressiva neurologiska funktionsförlusten.

Sista decenniernas genombrott vad gäller behandling av MS har ingivit hopp om att kunna förbättra prognosen för många MS patienter. Framgångarna har dock i princip gällt RRMS och förståelsen för vilka mekanismer som är speciella för progressiv (P) MS är fortfarande bristfällig.

Tanken bakom avhandlingens studier var att med hjälp av biomarkörer i ryggmärgsvätska (CSF), hitta samband för att öka förståelsen av dessa mekanismer. Vi ville också karaktärisera biomarkörerna för att bestämma deras förmåga att avspegla sjukdomsaktivitet, progression och terapeutiska effekter vid PMS.

I CSF undersöktes nivåer av i) neurofilament (NFL), en axonskademarkör, ii) gliafibrillärt surt protein (GFAP), en markör för astrocytaktivering, iii) BACE1 aktivitet och amyloid precursor protein/betamyloid (sAPP/A $\beta$ ) metabolismen och iv) tecken på B-cells reglering av inflammation (CXCL13) och en ökad produktion av immunoglobuliner.

Studierna har byggts upp kring tre studiedesigner med tillhörande patient- och kontroll populationer. I den första användes en tvärsnittsanalys, i den andra gjordes en långtidsuppföljning, och i den tredje studerades effekten på biomarkörer under immunomodulerande eller immunosuppressiv terapi (natalizumab, mitoxantrone eller rituximab). Patienterna som långtidsuppföljdes och de som behandlades undersöktes vid två tillfällen med intervall på 8-10 år respektive 1-2 år.

CSF biomarkörerna visar att inflammation utgör en betydande del av sjukdomsmekanismen också vid PMS och att den går att påverka med läkemedel som dämpar immunaktiviteten. NFL nivåerna var stegrade vid PMS. Behandling med cellgiftet mitoxantrone och det immundämpande medlet rituximab sänkte NFL nivåerna. Vi såg en tydlig koppling mellan immunrespons (CXCL13 nivåer och inflammation synlig på magnetkamera(MRI)) och axonskada (NFL).

Under hela MS förloppet sågs tecken på aktivering av hjärnans stödjeceller med ökat läckage av GFAP till CSF. Detta var mest uttalat vid hög sjukdomsaktivitet som ett tecken på ett akut astrocytsvar men sågs också vid långvarig sjukdom som ett troligt mått på den sammanlagda spridda plaque bildningen i CNS. GFAP nivåerna visade ett samband med klinisk funktionsnedsättning och visade prognostiskt värde men påverkades inte av terapi.

BACE1 aktivitet och sAPP/Aβ metabolismen har tidigare framförallt studerats vid Alzheimer's sjukdom, men har sista åren också studeras vid inflammatoriska sjukdomar. I våra studier fann vi att BACE1 aktivitet och sAPP/Aβ nivåerna sjönk vid MS generellt, sannolikt som tecken på pågående inflammation och möjligen störd nerv funktion. Vid RRMS ökade nivåerna mot de normala efter natalizumab behandling vilket inte skedde efter mitoxantronebehandling av SPMS. Genom utvidgade studier med mass spectrometri sågs tecken på att metabolismen inte bara är sänkt utan också ändrad vid PMS genom att andra nedbrytningsprodukter bildas.

En generell uppfattning är att immunglobolinproduktionen av oligoklonala IgG band i CSF är oförändrade över tid. Våra observationer motsäger detta och visar en koppling till att OCB bildningen är relaterad till B-cells aktiviteten. Antalet band och mönstret av OCB ändrades över tid men inte av behandling med mitoxantrone.

Våra studier av biomarkörer i CSF talar för att inflammatorisk aktivitet har betydelse för degenerativa processer såsom axonal skada och ändrad sAPP/A $\beta$  metabolism. Vi visar att detta samband även finns vid PMS och att immunhämmande behandling kan påverka dessa processer. Några av dessa biomarkörer kan komma att få betydelse för värdering av sjukdomsaktivitet (NFL, CXCL13), progression och svårighetsgrad (GFAP) och monitorering av terapi (NFL, CXCL13, sAPP/A $\beta$ metaboliter) vid progressiv MS.

## List of original articles

### Paper I

**M. Axelsson**, C. Malmeström, S. Nilsson, S. Haghighi, L. Rosengren, J. Lycke *Glial fibrillary acidic protein: a potential biomarker for progression in multiple sclerosis. J Neurol 2011*; 258: 882-888

### Paper II

N. Mattsson, **M. Axelsson**, C Malmeström, G Wu, R Anckarsäter, S Sankaranarayan, U Andreasson, S Fredrikson, A. Gundersen, L Johnsen, T Fladby, A Tarkowski, E Trysberg, A Wallin, H Anckarsäter, J. Lycke, O Andersen, AJ. Simon, K Blennow, H Zetterberg *Reduced cerebrospinal fluid BACE1 activity in multiple sclerosis Mult Scler* 2009; 15: 448-454

### Paper III

K. Augutis<sup>®</sup>, M. Axelsson<sup>®</sup>, E. Portelius, G. Brinkmalm, U. Andreasson,
M. K Gustavsson, C. Malmeström, J. Lycke, K. Blennow, H. Zetterberg and
N. Mattsson
<sup>®</sup> contributed equally *Cerebrospinal fluid biomarkers of β-amyloid metabolism in multiple sclerosis Mult Scler* published online 15 October 2012

### Paper IV

**M. Axelsson**, C. Malmeström, M. Gunnarsson, H. Zetterberg, P. Sundström, J. Lycke<sup>®</sup>, A. Svenningsson<sup>®</sup> <sup>®</sup>contributed equally *Immunosuppressive therapy reduces axonal damage in progressive multiple sclerosis Manuscript-Submitted* 

### Paper V

**M. Axelsson**, N. Mattsson, C. Malmeström, H. Zetterberg, J. Lycke *The influence from disease duration, clinical course, and immunosuppressive therapy on the synthesis of intrathecal oligoclonal IgG bands in MS Manuscript-Submitted* 

# Contents

Lis	t of abl	breviations	12
1. I	ntrodu	ction	15
1.1	Genera	l background	15
1.2	MS epi	demiology	16
1.3	Aetiolo	egy of MS	17
	1.3.1		
	1.3.2	Environmental and lifestyle factors and MS risk	17
	1.3.3	Combination of genes and environmental or lifestyle factors	
1.4	Clinica	l course of MS	19
		stic criteria in MS	
1.6	Disease	e-modifying MS treatment	
	1.6.1	Interferon beta	
	1.6.2	Glatiramer acetate	
	1.6.3	Natalizumab	23
	1.6.4	Mitoxantrone	23
	1.6.5	Rituximab	24
2. I	mmuno	opathogenesis and pathology of MS	25
2.1	Genera	1 considerations	25
	2.1.1	MS lesion (plaque) formation	25
	2.1.2	Axonal degeneration	27
	2.1.3	Astrogliosis	27
	2.1.4	Suggested patterns of MS pathology (120)	
2.2	The hal	llmarks of grey matter pathology	
2.3	Mening	geal pathology	29
3. F	Pathop	hysiology of progressive MS	
	-	l considerations	
	3.1.1	Microglial activation	
	3.1.2	Altered ion homeostasis	
	3.1.3	Mitochondrial dysregulation	
4.	Measu	rements of disease activity and disability progression in MS	
		e rate	
		l scales of neurological disability, progression and severity	
	4.2.1	Expanded Disability Status Scale, EDSS	
	4.2.2	Multiple Sclerosis Severity Score, MSSS	
	4.2.3	Progression index	
	4.2.4	Multiple Sclerosis Functional Composite, MSFC	
4.3	MRI		
	4.3.1	T1-weighted images	
	4.3.2	Gadolinium-enhanced T1-weighted images	
	4.3.3	T2-weighted images	
	4.3.4	Fluid-Attenuated Inversion Recovery (FLAIR)	
	4.3.5	MRI as a surrogate marker for MS activity and progression	
4.4		mical biomarkers in MS	
	4.4.1	General considerations	

	4.4.2	Structural biomarkers	36
	4.4.3	Inflammatory biomarkers	38
	4.5.4	Biomarkers of the sAPP/A $\beta$ metabolism	39
5. /	Aims of	f the study	42
6. 9	Subject	s and methods	43
6.1	MS pop	pulations	43
	6.1.1	Patients included in the cross-sectional study of the sAPP/AB pathway (Paper II)	43
	6.1.2	MS patients with long-term follow-up (Papers I, II, III, and V)	44
	6.1.3	MS patients treated with immunosuppressive or second-line	
		immunomodulatory treatment (Papers III, IV, and V)	
6.2		l subjects	
	6.2.1	HC and SAS included in the cross-sectional study of the sAPP-A $\beta$ pathway (Paper II)	
	6.2.2	HC population with long-term follow-up (Papers I-V)	
	6.2.3	Control population with systemic lupus erythematosus (Paper II)	
		l assessments, MRI, and serum and CSF sampling	
6.4	-		
	6.4.1	Polyclonal NFL assay (Paper I)	
	6.4.2	Monoclonal NFL assay (Paper IV)	
	6.4.3	Glial fibrillary acidic protein assay (Papers I and IV)	
	6.4.4	CXCL13 (Papers IV and V)	
	6.4.5	Albumin ratio (Paper V)	
	6.4.6	IgG index, CSF-specific oligoclonal IgG bands (Paper V)	
	6.4.7	BACE1 (β-Secretase ) activity (Paper II)	
	6.4.8	<i>α</i> -sAPP and β-sAPP (Papers II and III)	
	6.4.9	A $\beta$ X-38, A $\beta$ X-40, and A $\beta$ X-42 (Papers II and III)	
	6.4.10		
	6.4.11	Immunoprecipitation and mass spectrometry (Paper III)	
		Liquid chromatography and tandem mass spectrometry (Paper III)	
6.5		CS	
	6.5.1	Multivariate analyses	
		i	
7.1	Paper I		54
	7.1.1	Increased CSF GFAP levels in MS patients	54
	7.1.2	CSF GFAP correlated with progression	54
	7.1.3	GFAP correlated with clinical disability and was closely related to neurological	
		disability	
	7.1.4	GFAP had predictive value	55
	7.1.5	NFL was not elevated in clinically stable MS but some patients showed signs of	
		subclinical disease activity	
7.2	Paper I	Ι	
	7.2.1	BACE1 distinguished MS from other inflammatory disease and controls	
	7.2.2	BACE1 activity correlated with APP metabolites that were altered in MS	
	7.2.3	BACE1 activity decreased towards progressive disease course	
7.3	-	II	
	7.3.1	The APP metabolites were decreased in MS patients	
	7.3.2	Natalizumab normalized the APP metabolite levels	
	7.3.3	Progressive MS patients exhibited altered sAPP/Aβ metabolism	60

7.4	Paper I	V	60
	7.4.1	NFL levels were reduced by immunosuppressive treatment in progressive MS patients	60
	7.4.2	First-line disease-modifying therapies seemed to influence the NFL levels in	
		progressive MS	
	7.4.3	NFL levels seemed to influence Gadolinium enhancement on MRI in progressive MS	62
	7.4.4	CXCL13 levels in CSF of progressive MS were normalized by immunosuppressive	
		treatment	63
	7.4.5	GFAP levels were increased in progressive MS patients but unaffected by	()
	746	immunosuppressive treatment	
75	7.4.6	NFL were correlated with CXCL 13, and GFAP in progressive MS	
1.5	Paper V 7.5.1	/ The OCB pattern changed over 8-10 years of follow-up	
	7.5.2	Immunosuppressive treatment did not affect the OCB	
	7.5.2	CXCL13 levels were decreased following immunosuppressive treatment of	04
	1.3.3	progressive MS	64
	7.5.4	Correlations between OCB, IgG synthesis, CXCL13, and clinical parameters	
ог		sion	
		l considerations regarding biomarkers in MS	
		ochemical biomarkers related to PMS disease activity	
0.2	8.2.1	Neurofilament light protein	
	8.2.2	Glial fibrillary acidic protein	
	8.2.3	CXCL13	
	8.2.4	BACE1 activity and sAPP/Aβ metabolites	
8.3	Bioche	mical biomarkers in CSF reflecting disability development, progression,	
		rity of PMS	68
	8.3.1	Neurofilament light protein	69
	8.3.2	Glial fibrillary acidic protein	69
	8.3.3	CXCL13	70
	8.3.4	Intrathecal IgG synthesis	
	8.3.5	BACE1 activity and sAPP/Aβ metabolites	
		ochemical biomarkers for discriminating between MS clinical courses	
8.5		rkers for determining therapeutic efficacy in MS	
	8.5.1	Neurofilament light protein	
	8.5.2	Glial fibrillary acidic protein	
	8.5.3	CXCL13 and intrathecal IgG synthesis	
	8.5.4	sAPP/A $\beta$ metabolites	
8.6		mical biomarkers for exploring the pathophysiology of progressive MS	
	8.6.1	Axonal degeneration	
	8.6.2	Astrogliosis The role of the B cell lineage in PMS (CXCL13, IgG index, and OCB)	
	8.6.3 8.6.4	BACE1 activity and sAPP/Aβ metabolism	
		ding remarks and future considerations	
		dgements	
Ret	ference	9S	79

### List of abbreviations

Αβ	Beta amyloid
APP	Amyloid precursor protein
BACE1	β-site APP cleaving enzyme
BBB	Blood-brain barrier
CD	Cluster of differentiation
CIS	Clinically isolated syndrome
CNS	Central nervous system
CSF	Cerebrospinal fluid
CXCL 13	Cys-X-Cys motif ligand 13
CV	Coefficient of variation
DIS	Dissemination in space
DIT	Dissemination in time
DMT	Disease modifying treatment
DNA	Deoxyribonucleic acid
EAE	Experimental autoimmune encephalomyelitis
EBV	Epstein Barr virus
EDSS	Expanded disability status scale
ELISA	Enzyme linked immunosorbent assay
FLAIR	Fluid attenuated inversion recovery
FS	Functional system
GA	Glatiramer acetate
$\mathrm{Gd}^+$	Gadolinium enhancement
GFAP	Glial fibrillary acidic protein
HC	Healthy controls
HLA	Human leukocyte antigen
HR	Hazard ratio
IFNB	Interferon beta
IL	Interleukin
Ig G	Immunoglobulin G
IP	Immunoprecipitation

kDa	kilo Dalton
LC-MS/MS	liquid chromatography
MAG	Myelin-associated glycoprotein
MALDI-TOFMS	matrix assisted laser desorption/ionization time-of-flight mass spectrometry
МНС	Major histocompatibility complex
MRI	Magnetic resonance imaging
MS	Multiple sclerosis
MSFC	Multiple sclerosis functional composite scale
MS-MS	Tandem mass spectrometry
MSSS	Multiple sclerosis severity score
Mx	Mitoxantrone
NAB	Neutralizing antibodies
NFH	Neurofilament-heavy chain
NFL	Neurofilament-light chain
NFM	Neurofilament-medium chain
NRG1	Neuregulin 1
Nz	Natalizumab
OCB	Oligoclonal IgG bands
OR	Odds ratio
PML	Progressive multifocal leukoencephalopathy
PMS	Progressive multiple sclerosis
PPMS	Primary progressive multiple sclerosis
RRMS	Relapsing remitting multiple sclerosis
sAPP	soluble amyloid precursor protein
SAS	Spinal anaesthesia subjects
SD	Standard deviation
SLE	Systemic lupus erythematosus
SPMS	Secondar progressive multiple sclerosis
Th 2 cells	T helper 2 cells
TNFα	Tumor necrosis factor a
VCAM-1	Vascular cell adhesion molecule 1

# 1. Introduction

### 1.1 General background

Multiple Sclerosis (MS) is an organ-specific autoimmune disease of the central nervous system (CNS). The aetiology and pathogenesis of MS remain largely unknown. There is evidence that it may begin as a primarily inflammatory disorder that subsequently takes on degenerative features; however the inflammation activity seems to vary widely between patients (1).

MS is initially dominated by focal white matter inflammatory infiltrates, with demyelinating lesions also appearing in the cortex and deep grey matter of the brain. Additionally, the CNS tissue shows diffuse and global changes, including signs of widespread inflammation (2), microglial activation (3), neuron and axon damage (4), oligodendrocyte depletion (5), and astrogliosis (6). Irreversible degeneration appears early in the disease process (7), and brain atrophy may already be observable at the clinical onset of MS (clinically isolated syndrome; CIS).

The initial clinical course is usually relapsing-remitting (RR) with transient episodes of neurological symptoms. In most cases, this eventually changes into a secondary progressive (SP) course characterized by continuous accumulation of neurological disability with or without superimposed relapses. Over time, the inflammatory activity decreases and CNS degeneration becomes more prominent. The role of inflammation in neurodegeneration and the pathophysiology behind secondary progression are essentially unknown. Further investigations are required to determine whether they are independent from each other, or if inflammation is responsible for secondary degeneration.

Magnetic resonance imaging (MRI) has become the dominant method for diagnosis, monitoring disease activity, and evaluating treatment effects. Although, MRI may differentiate between inflammatory activity and degenerative processes to some extent, this technique cannot be used to identify different pathophysiological processes of MS. Moreover, during the progressive phase of MS, the appearance of new lesions may be undetectable due to confluent lesion formation, and the methods for determining lesion volume and atrophy are laborious and commonly unavailable.

Biochemical biomarkers of body fluids, especially cerebrospinal fluid (CSF), have increasingly gained attention in studies of MS. For decades, selective detection of oligoclonal IgG bands in CSF has been used for diagnostic purposes (8). More recently, it has been found that several inflammatory and CNS parenchymal biomarkers seem to reflect important pathological processes in MS, with some being related to disease activity, disease severity, and clinical course (9).

As patients change from RRMS to SPMS, they exhibit several features that resemble those of known neurodegenerative diseases. They develop irreversible disability, are relatively unresponsive to immune modulatory or immunosuppressive treatment, and show brain and spinal cord atrophy. In the present thesis, we explored the pathophysiology behind this transformation by measuring the levels of different biomarkers in CSF. Our major objectives were to find biomarkers that are associated with clinical course, predict disease severity, and reflect the effects of therapeutic intervention.

### 1.2 MS epidemiology

MS is the main non-traumatic cause of neurological disability among young adults in Sweden. The Swedish MS prevalence is among the highest in the world (189/100 000), with approximately 17 500 diagnosed cases as of the end of 2008 (10). Previous calculations show a Swedish yearly MS incidence of between 3.9-5.2/100 000 (11, 12), but a recent nationwide study estimates this incidence to be twice as high (Ahlgren et al., unpublished data). MS distribution varies widely in different parts of the world; temperate climate zones are considered high-risk areas (13), and the risk seems to increase with the distance from the equator (14). There are exceptions to this latitude gradient, with prevalence being lower in northern Norway (73/100 000) (15) and higher on Sardinia (152/100 000) (16) than in surrounding areas at the same latitude. In Sweden, the MS prevalence increases by 1% for women and 1.5% for men per degree of latitude increase (10). Studies of populations with mixed genetic background at the same latitude have suggested that the risk of MS development depends on ethnicity (16). In the 1970s, it was found that people with African and Asian backgrounds, respectively, had 50% and 20% lower MS risks compared to Caucasians. However, a recent investigation showed reversed numbers, with an increased MS risk among African Americans compared to other groups (17).

MS with relapse onset affects women twice as often as men, while this female preponderance is less obvious in primary progressive MS (PPMS). The female/male ratio in Sweden is 2.35 (10). Several studies have found that RRMS occurs with an increased female/male ratio (18). The increased MS risk for women seems to be dependent of the year of birth and geographic location (19), with the highest female/ male ratio (4.55) observed among patients born in 1980-1989 in northern Europe. Socioeconomic factors (20) or migration from low- to high-risk areas (21) might affect the gender ratio. These observations appear to suggest on-going changes in MS incidence, but this has not been confirmed (10).

### 1.3 Aetiology of MS

Although the aetiology of MS is essentially unknown, there is accumulating evidence that both genetic and environmental factors influence MS risk.

### 1.3.1 Genes and MS

An increased risk is found in genetic relatives of MS patients, but the heredity of MS is complex. The background risk of MS is about 0.2% in Sweden (10). For a person with an affected relative, the risk of MS development is estimated to be 25-30% for a monozygotic twin, 3-5% for a dizygotic twin or sibling, 2% for a parent or child, and 1% for other second and third degree relatives (22, 23).

The search for MS risk genes was initially focused on genes regulating the immune system. Among all genes, the strongest association has been found with the HLA class II genotype DRB1\*15:03 (on the short arm of chromosome 6); this genotype is carried by 28-33% of northern Caucasian MS patients compared to 9-15% of heal-thy controls, having an OR of 3.08 (24). Protective genes have also been isolated among MS patients. HLA A\*02 is the most potent independent risk reducer, with an OR of 0.73 (25). HLA-C\*05 and HLA B\*44 have also been found to reduce MS risk, both independently and in combination with each other (26, 27).

Genome-wide association studies in vast multinational MS and matching control populations have detected at least 57 risk loci, the majority with OR in the range of 1.1-1.3 (28, 29). Notably, the majority of identified non-HLA genes have also been located in or near immune system-regulating genes. One-third of the MS-associated genes are also associated with other autoimmune diseases (30, 31).

### 1.3.2 Environmental and lifestyle factors and MS risk

Among the environmental and lifestyle factors that have been suspected to influence MS risk and prognosis, few have been convincingly associated with MS in repeated studies.

### 1.3.2.1 Infections and MS

Many infectious agents have been suspected to either trigger MS or maintain the disease as a chronic CNS infection. The proposal that infections are involved in MS aetiology and pathogenesis is based on the observation that people who migrate during adulthood from low to high MS prevalence areas or vice versa retain their original risk, but their children are at the risk determined by their new location (32, 33). It has been suggested that MS incidence rose after increased migration from high MS prevalence areas to isolated environments (Faroe, Sardinia, etc.), leading to "MS epidemics" (34-36).

### 1.3.2.2 Epstein-Barr virus

Epstein-Barr virus (EBV) is one of several human herpesviridae that, after an initial infection, hibernate in humans; EBV predominantly hibernates in B lymphocytes. Among adult humans, 90-95% are seropositive, compared to almost 100% of MS patients (37, 38). Individuals who are seronegative for EBV have an OR of only 0.06 for developing MS compared to EBV-seropositive persons (39). Furthermore, previous clinical mononucleosis increases the risk of developing MS by 2-3 times (40). An intriguing but unconfirmed finding is the detection of EBV-infected B cells in germ-like follicle structures of the meninges in SPMS patients (41).

### 1.3.2.3 Tobacco smoke

Tobacco smoke seems to influence the risk and prognosis of MS. The relative risk of MS development is higher among current smokers compared to individuals who have never smoked (OR, 1.6) (42). MS risk is also increased among children of smokers (OR, 2.12) (43) and for non-smoking adults who are exposed to tobacco smoke (OR, 1.3) (44). Compared to non-smokers, smokers have a higher risk of developing MS from CIS (HR, 1.8) (45), and have a higher lesion load and increased atrophy development seen on MRI (46).

### 1.3.2.4 Vitamin D and sun exposure

It is well documented that MS prevalence increases with higher latitude (14), and this association has been connected to low sunlight exposure and less endogenous vitamin D production (47). It has been suggested that UVB radiation has an independent protective effect on MS risk (48, 49). High serum level of 25-hydroxy vitamin D is a risk reducer for MS (OR, 0.59) (50), and vitamin D seems to suppress disease activity. It has been demonstrated that peroral vitamin D treatment leads to a lower relapse rate compared to in a placebo-treated group (51, 52), and supplemental vitamin D therapy has been suggested as a potential MS treatment.

### 1.3.3 Combination of genes and environmental or lifestyle factors

Like many other diseases, MS seems to emerge due to a combination of genetic vulnerability and harmful environment. Recent studies have shown that >80% of the isolated MS risk genes are regulated by vitamin D (53, 54), including HLA DRB1\*15:01. Among smokers, presence of the HLA DRB1\* 15:01 genotype and absence of the protective HLA A\*02 genotype gives an OR of 13.5 for MS development (55), compared to an OR of 4.9 in non-smokers with the same gene set. The current knowledge of risk genes is an important resource to promote better understanding and identification of pathological processes. This information may also be useful in establishing risk profiles for individual environmental factors and predicting treatment effects.

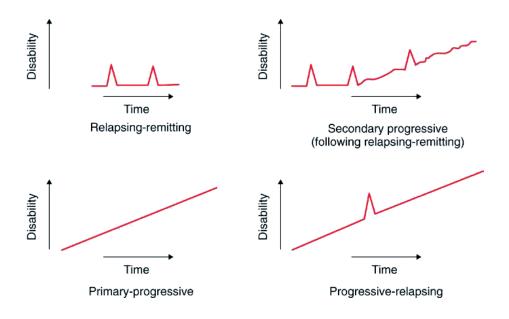
### 1.4 Clinical course of MS

In 80-90% of cases, the clinical onset of MS is transient and followed by complete or partial recovery from symptoms (23) *(Figure 1)*. The symptoms may have monoor multifocal origin in the CNS, most commonly involving the optic nerve, the spinal cord, or the brain stem. New relapses usually occur with an annual rate of 0.5-1. Over time, the recovery from each relapse becomes less complete, and persistent symptoms accumulate. In an untreated MS population, a majority of RRMS cases turn into SPMS after a median time of approximately 15-20 years from disease onset (23, 56). SPMS patients may initially have relapses that are superimposed over the on-going clinically progressive process. In most cases, clinical progression consists of spastic para- or tetraparesis, cerebellar ataxia, or spastic hemiparesis, and the symptoms gradually become more complex and severe with increasing decline of neurological functions. In 10-15% of patients, the course is progressive from onset; these patients are designated PPMS.

After clinical onset and before there is evidence for the chronic and disseminated disease of MS, the term CIS is used (57). A majority of patients with CIS will eventually be diagnosed with definitive MS once dissemination in space and time is fulfilled (*Figure 2*).

The term benign MS is often defined as a case with an EDSS of  $\leq 2.0-3.0$  after a disease duration of 10-15 years (58). Studies of benign MS show the frequent presence of non-motor symptoms, like fatigue, pain, and cognitive impairment (59). The term is retrospective and it is not possible to predict the individual clinical course in MS; it has been proposed that a better definition is needed (60).

It is debated whether different MS disease courses reflect different pathogenic mechanisms. RRMS and SPMS are per definition parts of the same disease, and the majority of untreated RRMS patients will proceed to develop SPMS. On the other hand, PPMS has a different clinical presentation and gender distribution; however, little evidence supports different mechanisms behind this process. PPMS and SPMS involve the same age of onset and similar disability development (61). The set of risk genes (HLA and non-HLA genes) also seems to be the same for both subtypes (28).

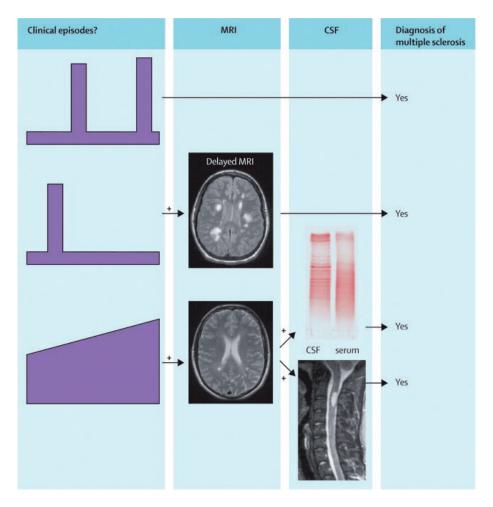


#### Figure 1

Clinical subtypes of MS. The figure shows the two main types of onset, relapsing or progressive, and totally four subgroups depending on the further course of disease. (Adapted from (64) reprinted with permission from with permission from Wolters Kluwer Health.)

### 1.5 Diagnostic criteria in MS

The diagnostic workup of MS is currently based on MRI results rather than clinical measurements; however, MS diagnosis still requires evidence of CNS white matter lesions that are disseminated in time (DIT) and space (DIS) and not better explained by any other diagnosis. In parallel with improved radiological and biochemical methods and the unification of clinical evaluations among neurologists, the MS diagnostic criteria have been repeatedly modified and revised. The progression from the criteria of Schumacher (8) and Poser (62) to the current revised criteria of McDonald (63) represents an on-going process of simplifying the diagnostic workup to allow early diagnosis without losing diagnostic sensitivity and specificity. This evolution has included almost complete elimination of the influence of other paraclinical methods, such as visual evoked potentials or detection of oligoclonal IgG bands in CSF. Evidence of DIT and DIS can be obtained by a single or repeated MRI, or by observation of new clinical attacks.



### Figure 2

Localization of lesions and demonstration of dissemination in space (different locations in CNS) and time have been core concepts in diagnosing MS. Clinical and paraclinical investigations are usually applied. (From (23), reprinted with permission from Elsevier).

In contrast with previous MS diagnostic criteria, the criteria of McDonald include PPMS. The diagnosis of PPMS requires continuous worsening of CNS symptoms over at least one year combined with paraclinical signs on MRI, or signs of inflammation in CSF, such as increased IgG index and/or selective formation of oligoclonal IgG bands (OCB). SPMS diagnosis is based on evidence of continued worsening for at least six months, without being associated with relapses (64), although some SPMS patients have relapses superimposed on their progression. The transition

from RRMS to SPMS can be difficult to assess, especially in RRMS patients with remaining disability following severe relapses and in SPMS patients who continue to have relapses.

### 1.6 Disease-modifying MS treatment

### 1.6.1 Interferon beta

In the mid-1990s, interferon beta-1b (IFNB-1b) and interferon beta 1a (INFB-1a) were approved as the first disease-modifying therapies for MS. INFB-1a is identical to endogenous synthesized interferon beta, while INFB-1b lacks glycosylation and differs by one amino acid substitution and one amino acid deletion. These IFNBs are now available as different products that are administered either subcutaneously or as intramuscular injections, with a frequency ranging from every other day to once weekly.

IFNBs belong to a group of cytokines that acts on target cells through the IFN-alpha/ beta receptor (65). They activate a cascade of genes that induce the synthesis of proteins with immunomodulatory properties, but the exact function remains obscure. IFNBs agonize the production of anti-inflammatory cytokines (e.g. IL-4 and 10), promoting a shift toward a Th2 response (66); they also seem to inhibit production of pro-inflammatory cytokines (e.g. IL-12, IL-17, IL-23, osteopontin, IFN-gamma, and TNF-alpha) (67, 68). IFNB also appears to stimulate T-cell apoptosis by downregulating anti-apoptotic proteins (69). The mechanisms are complex and far from completely understood.

IFNB treatment of RRMS reduces the relapse rate by 30%, and reduces MRI lesions and disability development. In CIS, trials have demonstrated benefits of early treatment, although the long-term effect is uncertain (70). IFNB treatment of progressive MS, with few exceptions, has failed to reduce neurological disability (71, 72); however, it significantly affects relapse rate and lesion development (71, 73, 74).

The most common adverse effects include flu-like symptoms and skin reactions (75), which are the major causes of treatment discontinuation. Neutralizing antibodies (NAB) against IFNB may develop in 5-35% of patients, depending on the INFB product (76), and a high NAB titre can reduce or completely eliminate the effects of the drug (77).

### 1.6.2 Glatiramer acetate

Glatiramer acetate (GA) is a combination of four amino acids (L-alanine, L-glutamic acid, L-lysine, and L-tyrosine) that are randomly polymerized into peptides. These

peptides have immunomodulatory effects that are poorly understood. GA attaches to the MHC II complex and seems to disrupt antigen presentation, which might block MBP-reactive T cells causing a shift to an anti-inflammatory Th2 state (78, 79). Since GA does not pass the blood-brain barrier (BBB), this process occurs in the periphery and activated T cells migrate into the CNS and induce anti-inflammatory effects (80). GA has been shown to affect the antigen-presenting properties of Bcells. Although anti-GA antibodies are frequently seen, they do not seem to have an inhibitory effect (81, 82).

Treatment of RRMS patients with GA seems to have a clinical effect of the same magnitude as INFB treatment; head-to-head studies have not shown IFNB to be superior to GA (83-85). In progressive MS, GA treatment induced no significant clinical effects but on MRI lesion development (86, 87).

GA is generally well tolerated, but injection reactions occur in a majority of patients (75). One specific injection-related adverse effect is the immediate post-injection systemic reaction (IPISR) that includes dyspnoea, palpitation, flushing, and anxiety occurring for between 30 seconds and 30 minutes (88).

### 1.6.3 Natalizumab

Natalizumab (Nz) is a humanized monoclonal antibody directed against the  $\alpha 4\beta 1$ integrin molecule on mononuclear leukocytes. By blocking the interaction of  $\alpha 4\beta 1$ integrin with the endothelial vascular cell adhesion molecule-1 (VCAM-1) ligand, Nz inhibits leukocyte migration across the BBB. Pivotal clinical trials have shown that Nz treatment decreases relapse rates by approximately 70% versus placebo, and clinical and MRI measurements showed that 37% of Nz-treated patients were disease free, compared to 8% of placebo-treated patients (89). No randomized placebo-controlled clinical trial has yet been performed in progressive MS patients.

The major problem with Nz treatment in MS is the appearance of progressive multifocal leukoencephalopathy (PML), which is caused by an opportunistic polyoma virus (JC virus) infection colonizing the kidneys and bone marrow. In the absence of appropriate immune defence of the CNS, mutated JC virus has the ability to cause a massive CNS infection. Although the effect of Nz can be reversed, PML often causes lasting neurological disability and, in about 20% of cases, death (90, 91). Strategies to minimize the risks of PML include selection of patients without previous immune suppressive treatment and those that are negative for JC virus antibodies.

### 1.6.4 Mitoxantrone

Mitoxantrone (Mx) is a synthetic DNA-intercalating anthracenedione derivate that affects B cells, T helper cells, and cytotoxic T cells, and can both suppress and mo-

dulate the immune system (92). Mx does not cross an intact BBB. It is the only drug approved for use in SPMS (in the USA, but not Europe). Two randomized controlled studies have shown efficacy in all stages of MS, except PPMS, with effects on both relapse rate and EDSS progression (93, 94).

However, Mx exerts dose-dependent toxicity on many organs, leading to increased risk of acute myeloid leukaemia, cardiac dysfunction in about 1% (95), and increased risk of serious infections. The cumulative lifetime dose should not exceed  $100 \text{ mg/m}^2$  for MS treatment (96).

### 1.6.5 Rituximab

Rituximab is a chimeric human/mouse anti-CD20 antibody. The Fab domain binds to the CD20 antigen on B lymphocytes, and the Fc domain recruits the immune system to mediate cell death (97). This antibody-dependent cytotoxicity is induced by either apoptosis or complement-dependent cytotoxicity (98-100). Because CD20 expression is unique to B cells, the beneficial effects of rituximab in MS support a role of B cells in MS pathogenesis. It is unclear how much of the effect of rituximab is generated outside the CNS, but determination of the BBB penetration shows a CSF/plasma ratio of 1/1000, which might be sufficient to also eliminate B cells from the CSF compartment (101).

Although rituximab is not registered for MS treatment, a number of open-label studies and case reports have shown clinical and radiological beneficial effects (102); it is used off-label and considered a potent agent for MS therapy. One phase II study of rituximab treatment in patients with RRMS showed significantly reduced relapse rate and lesion formation (103). In contrast, a phase III study in patients with PPMS showed non-significant differences; however, reduced disease progression was observed in a subgroup of patients younger than 51 years with Gd-enhancing (Gd<sup>+</sup>) lesions (104).

## 2. Immunopathogenesis and pathology of MS

### 2.1 General considerations

MS is considered to be an autoimmune disease characterized by focal lesions disseminated throughout all parts of the CNS, and involving both grey and white matter. Widespread diffuse pathology is also seen in normal-appearing tissue. In the early phase of MS, inflammatory activity dominates, with a high rate of lesion formation; subsequently, this activity declines and neurodegeneration takes over (2). However, there is accumulating evidence that signs of neurodegeneration—including neuroaxonal loss, astrogliosis, and CNS atrophy—are already evident at disease onset (105). It remains to be determined whether neurodegeneration in MS is secondary to destructive immune activity or essentially a parallel and primary event.

### 2.1.1 MS lesion (plaque) formation

MS lesion or plaque formation typically involves focal inflammation with bloodbrain barrier (BBB) breakdown and immune cell infiltration. Inflammation seems to be initiated in two steps. CD8+ T cells and activated microglia initially dominate, causing myelin destruction (106, 107). This destruction is followed by infiltration of activated macrophages, B cells, and T cells, and local secretion of pro-inflammatory cytokines and chemokines and their receptors (108, 109). Active lesions often give rise to BBB disruption, which can be detected by Gd<sup>+</sup> enhancement on MRI (110, 111). As inflammation declines, MS lesions may either progress to a chronic inactive stage that is characterized by astrogliosis and insufficient remyelination, or show sufficiently remyelinated axons that appear as "shadow plaques" on MRI. Some lesions, designated as chronic active plaques, show preserved immune activity of lower intensity (Figure 3); these lesions slowly expand at the border, while activity ceases at the centre (3). MS progression involves decreased active lesion formation, along with increased expanding chronic lesions. In progressive MS, inflammation seems to become more compartmentalized and the integrity of the BBB is maintained, allowing only low levels of protein exchange (111) (Figure 4).

MULTIPLE SCLEROSIS LESION

MULTIPLE SCLEROSIS LESION

otin

1.7.1

# HULTIPLE SCLEROSIS LESION

### Figure 3

Schematic, MRI and microscopic images of MS lesions in different stages. Reprinted with permission from http://multiple-sclerosis-research.blogspot.se/

### 2.1.2 Axonal degeneration

Axonal damage occurs in both early and late stages of MS, predominantly in active plaques (early and chronic), and correlates to the activity of lymphocytes and activated microglia (112). Ferguson et al. described accumulation of amyloid precursor protein (APP; a marker for axonal dysfunction or injury) all over active lesions and at the border of chronic active lesions (113). Axonal degeneration is also seen in non-lesion matter. Diffuse axonal injury and destruction are associated with wide-spread and diffuse low-grade inflammation, microglial activation, astrocytic gliosis, and mild demyelination (114), illustrating an active neurodegenerative process. The quantity of diffuse injury increases over time, and is more pronounced in progressive MS. Axonal loss outside of plaques could also be due to Wallerian degeneration, in which proximal axonal damage causes distal axonal degeneration. However, the extent of diffuse white matter injury does not correlate with the amount of focal white matter lesions, and only weakly correlates with cortical demyelination (2, 115).

### 2.1.3 Astrogliosis

Reactive gliosis with or without scar formation is a general feature of any kind of CNS damage and is a prominent feature in MS pathology (6). In areas outside of the plaques, the picture is diffuse with widespread areas of hypertrophic astrocytes with up regulated GFAP expression (116). In more severely affected areas, the astrocytes proliferate outside of their normal tissue architecture. In plaques, in addition to astroglial activation, dense and compact glial scars are formed (117); recent studies suggest that these scars act as neuroprotective barriers that stop inflammatory cells and predominantly form along the plaque borders (118). Glial scars interact with other cell types, and their extra cellular matrix contains substances that inhibit cellular migration (119).

### 2.1.4 Suggested patterns of MS pathology (120)

Investigation of the heterogeneity of MS pathology has led to the suggestion that there are four different and distinct immunopathogenetic patterns; these findings have been considered proof that there are different types of MS that may respond differently to treatments (121, 122). Pattern I shows T cells and macrophages around blood vessels, preserving oligodendrocytes but with no complement activation. Pattern II is like pattern I, but with complement activation. Pattern III shows diffuse inflammation, distal oligodendrogliopathy, microglial activation, and a loss of myelin-associated glycoprotein (MAG); contrary to patterns I and II, pattern III has no association with blood vessels. Pattern IV includes sharp bordered lesions, and oligodendrocyte degeneration with a rim of normal-appearing white matter; no complement activation or MAG loss is detected. These observations are based on biopsy or autopsy materials from severely disabled patients, and the results have not been confirmed. It remains to be clarified whether these patterns represent different disease subgroups or different stages of the same disease.

a RRMS	RPMS	SPMS	PPMS
	Age and	disease duratio	n
	Path	ological change	s
<ul> <li>Blood–brain bar</li> <li>New active CNS</li> </ul>		CNS Menin Slow Subpi	ed inflammation igeal inflammatory aggregates expansion of pre-existing lesions al cortical demyelination e white matter injury atrophy
b RRMS	RPMS	SPMS	PPMS
RRIVIS	ILE WIS	36,00	11110
RRINO		disease duratio	
CIVITA			
CMINA	Age and		n

### Figure 4

Schematic presentation of the evolution of structural pathology and disease mechanisms during the course of MS. a) Pathological features associated with conversion of RRMS (pink) to PMS (green). b) Changes in disease mechanisms associated with conversion of RRMS (pink) to PMS (green). The bars indicate the the extent of these differences in relation to increasing age and disease duration. Although no pathological or mechanistic feature is exclusive either for RRMS or PMS, major quantitative differences in their occurrence are evident between these stages. Abbreviations: iNOS, inducible nitric oxide synthase, RPMS relapsing progressive MS. From (131). Reprinted with permission from Nature Publishing Group.

### 2.2 The hallmarks of grey matter pathology

In MS, involvement of the deep grey matter and cortex is seen as either demyelination or retrograde degeneration from white matter lesions. Grey matter pathology and white matter pathology share many common features, but there are also fundamental differences. The cortex exhibits both focal and diffuse pathology, including atrophy (123), as is also observed in early stages of MS (124). Focal changes predominantly appear early on, and are often dominated by intense inflammation with perivascular infiltrates and large amounts of lymphocytes throughout the tissue, accompanied by activated macrophages and microglia (125). Losses of neurons, axons, and synapses are more pronounced in early-stage cortical plaques (2, 125). In general, cortical lesions are more commonly seen during the progressive phase of MS and in the subpial cortical layers (2), often in the vicinity of ectopic B-cell follicles of the meninges (126). The distance from the white matter seems to determine the content of inflammatory cells in grey matter lesions, with the subpial lesions dominated by activated microglia, apoptosis, and neuronal atrophy (127). Similar to white matter pathology in MS, global tissue loss is widespread in normal-appearing cortex, contributing to atrophy development (128).

### 2.3 Meningeal pathology

Topographically associated with subpial cortical lesions, meningeal B-cell folliclelike structures have been characterized in progressive MS (129, 130). Although it has not been confirmed, the B cells in these structures have been identified as immune reactive for Epstein-Barr virus (EBV) (41). These tertiary inflammatory germinal centres are predominantly found in SPMS (126), and are suggested to have a role in the pathogenesis of progressive MS. It is noteworthy that these findings indicate that MS can no longer be considered a disease affecting only nervous tissue.

# 3. Pathophysiology of progressive MS

### 3.1 General considerations

There are three main hypotheses explaining the pathophysiology of progressive MS and its relation to RRMS (131):

1. MS is a primary neurodegenerative disease. Dysfunctional and dying cells trigger an early immune response. Neurodegeneration may be modified or amplified by the reactive inflammation.

2. MS is a primary inflammatory disease that, over progressive stages, changes in anatomical location and intensity, making it untreatable using existing immune therapies.

3. MS develops through a combination of different pathological mechanisms, starting as an inflammatory disease and subsequently involving neurodegenerative processes. Delayed damage or neuronal death occurs when the reserve capacity is used and the anatomical structures are destroyed.

One general consideration is that axonal loss accumulates over the course of MS, eventually reaching a threshold at which the disease shifts to slow progression (132). It is likely that this process involves the imbalance between tissue injury and repair (133) and the consumption of compensatory mechanisms (132).

### 3.1.1 Microglial activation

Tissue injury in progressive MS is associated with chronically activated microglia (3), and activated microglia are found in normal-appearing white matter (134). Similar microglia involvement is seen in other neuroinflammatory and neurodegenerative diseases (135). Microglia are known to generate oxidative bursts and to induce demyelination and axonal damage; however, they also have neuroprotective functions (135).

### 3.1.2 Altered ion homeostasis

In normally myelinated axons, voltage-gated Na<sup>+</sup> channels are highly concentrated at the nodes of Ranvier. Loss of myelin is a major structural change during progressive MS. A number of studies have reported changes of ion homeostasis in demyelinated neurons, and redistribution of ion channels along the axon as a functional compensation. Altered expressions of voltage-gated Ca<sup>2+</sup> channels (136), glutamate receptor (137), and Na<sup>+</sup> channels (138, 139) have been observed, which can lead to intra-axonal Ca<sup>2+</sup> accumulation and eventually axonal death (131). This delayed process can be induced by inflammation-mediated demyelination and axonal injury that occurred several years earlier.

### 3.1.3 Mitochondrial dysregulation

Mitochondrial injury is observed in the demyelinated axons of MS lesions (140, 141). Axons have excessive energy demands when they are not supported by myelin, and mitochondrial function is critical during axonal injury. Increased mitochondrial number and size are seen in axons in active plaques, which normalize after remyelination (142-144). Mitochondrial damage and loss in nerve cell bodies could accelerate axonal death and induce a state of "virtual hypoxia" (140). Redistribution of mitochondria is observed within damaged nerve cells, as well as increased numbers of defective mitochondria (145). One major cause of mitochondrial dysfunction is oxidative stress induced by inflammation (145). In active MS lesions, it is likely due to increased production of enzymes and oxygen free radicals (146, 147). Thincalibre axons are more severely affected than thick ones because they have less mitochondria relative to their axonal surface area (140). The mitochondrial damage also affects oligodendrocytes and their ability to remyelinate injured axons (148, 149). Moreover, the release of high levels of extracellular  $Fe^{2+}$  during inflammation and tissue damage may lead to additional oxidative stress on mitochondria and axons (150, 151).

# 4. Measurements of disease activity and disability progression in MS

### 4.1 Relapse rate

Measuring the annual relapse frequency is a standard method for evaluating MS disease activity in clinical routine and clinical trials. A relapse is defined as a patient-reported or objectively observed event that is typical of an acute inflammatory demyelinating event in the CNS, current or historical, with duration of at least 24 hours, and in the absence of fever or infection (63). The average annual relapse rate is about 0.5-1 in an untreated population (89). A long observation time or large patient groups are needed to confirm altered activity.

# 4.2 Clinical scales of neurological disability, progression and severity

Clinical scales are used to score the progression of neurological deficit over time in clinical routine and clinical trials. A number of scales have been tested, with the aim of finding objective measurements of disability development in MS. Such measures should be reliable, MS specific, validated, and easy to use and interpret in clinical practice as well as in clinical research. The scales we used in this thesis are widely accepted and validated.

### 4.2.1 Expanded Disability Status Scale, EDSS

The currently dominant scale for clinical scoring is the Expanded Disability Status Score (EDSS), which gives patients a score from 0 to 10. Derived from the DSS (152), the EDSS is based on the evaluation of seven functional systems (FS) by targeted neurological examination. A synthesis of these evaluations gives an EDSS of up to 3.5 for which individual FS scores (with some exceptions) count equally. For scores in the range 4.0 to 6.5, walking ability is weighted as just as important as the FS scores. Scores of over 6.5 are given as an evaluation of the patient's independence and autonomy in ambulation. A score of 10 indicates death by MS. Obviously, this system leads to non-linear score development with a possibility of overlooking the deficit development, especially at higher scores. EDSS is an ordinal scale, which is only suitable for non-parametric statistics. Furthermore, the intra-rater repeatability is reportedly low (153), especially in mental and sensory FS where anamnestic information is necessary. Agreement between raters has been tested, and is only achieved with acceptance of differences  $\leq 1.5$  EDSS score (154). When using this scale, it should be noted that two EDSS steps are needed to confirm

clinical change (155), underlining the limitations of the EDSS in clinical studies or for therapy revisions. A major strength of the EDSS is its worldwide use and ease of use and interpretation.

### 4.2.2 Multiple Sclerosis Severity Score, MSSS

The Multiple Sclerosis Severity Score (MSSS) represents an attempt to determine the disability progression rate in MS; it is claimed that this scale measures MS severity and has predictive properties. Individual disease duration was combined with EDSS in 9892 patients from 11 countries (156) to establish the general EDSS development over the course of MS. By combining disease duration on the y-axis and the EDSS score on the x-axis, the MSSS score can be found in the matrix. Scores range from 0.01 to 9.99. Obviously the scores are non-linear and can only be calculated with non-parametric statistics. Patients with EDSS 10 (death) were not included in the investigation, underrepresenting severe cases of MS; MSSS scores were also not included for patients with disease duration exceeding 30 years. Thus, MSSS is based on a large—but in some aspects, historical—population. It could be questioned whether this population is still relevant for measuring MS severity.

### 4.2.3 Progression index

The progression index is calculated by EDSS/disease duration (157, 158). The main concept is the same as in MSSS, but the progression index has less predictive value. It has no limitations for use with patients with disease duration of above 30 years, but it has been scarcely used in previous studies.

### 4.2.4 Multiple Sclerosis Functional Composite, MSFC

The MSFC was established in 1999 as a complement to EDSS for overcoming the weaknesses of the widely used scale (159). The main idea was to use quantitative measurements and compare them with those derived from a large control population. The MSFC is based on three specific tests that examine walking speed in a short range (timed 25-foot walk; T25W), fine motor function and coordination in the arms (nine-hole peg test; 9HPT), and cognitive function (paced serial addition test, 3 sec; PASAT3). The test results are converted to a z-score and normalized to a control population. The z-score describes the number of standard deviations of a patient score in comparison with a reference population. This score has relatively high inter-rater agreement (160). Some studies claim that, compared to the EDSS, the T25W part of the MSFC is a better prognostic tool (161, 162), while other studies have reported that the T25W is equal or even inferior (163-165). The MSFC is time consuming and requires specific tools. It is mainly used in clinical treatment trials as a complement to the EDSS.

### 4.3 MRI

MRI has become the most important MS diagnostic tool (see MS diagnosis, page 20). The use of MRI in MS has increased along with the growing possibilities and demands for accurate diagnosis, assessment of disease activity and progression, and evaluation of therapeutic efficacy. Much of our current knowledge concerning disease activity, neurodegeneration, atrophy development, and grey matter involvement has been attained using MRI research. In phase II clinical trials, MRI measurements are used as surrogate markers for therapeutic outcome, and phase III clinical trials include several MRI-based secondary and tertiary objectives.

### 4.3.1 T1-weighted images

A T1 image is created by measuring the time that it takes protons to return to the magnetic field axis. MS lesions may appear as hypointense areas ("black holes"). These are preceded by  $Gd^+$  lesions, reflecting destructive inflammation. The degree of hypointensity correlates with the degree of pathological severity (166) and persistent hypodense areas reflect irreversible tissue destruction with axonal loss (166). T1 lesions correlate more strongly with clinical deficits than pathology seen on other sequences (167).

### 4.3.2 Gadolinium-enhanced T1-weighted images

A T1 image with Gd<sup>+</sup> lesions reflects a damaged BBB (168). BBB disruption appears during acute inflammation and may be detected for up to five weeks. Contrast enhancement is often connected to clinical symptoms, i.e. relapses (169). This sequence is a fundamental part of the diagnostic criteria, as it provides evidence of dissemination in space and time (63).

### 4.3.3 T2-weighted images

A T2 image is created by measuring the proton dephasing following a transverse pulse, and is useful for describing the anatomy and composition of the central nervous system. Lesions detected on T2-weighted images can reflect a wide diversity of pathological processes, such as inflammation, demyelination, and glial scar formation. This is the sequence that visualizes the expansion of MS in the CNS. T2 lesions appear as focal hyperdense areas with typical location (periventricular, infratentorial, or juxtacortical) and appearance (ovoid and >3 mm in diameter). T2 lesions are the main diagnostic source for establishing dissemination in time and space (63).

### 4.3.4. Fluid-Attenuated Inversion Recovery (FLAIR)

Fluid-Attenuated Inversion Recovery (FLAIR) produces T2-weighted images.

Using an inversion-recovery technique, the inversion time (TI; the time between inversion and excitation pulses) is carefully chosen for the CSF signal (170). In MS, FLAIR imaging is more sensitive for detecting lesions close to the ventricles than T2-weighted lesions, since these may not be distinguishable from the signal of CSF (171). FLAIR can also detect juxtacortical and cortical lesions (172, 173), but the appearance of infratentorial lesions may be false positive (174).

### 4.3.5. MRI as a surrogate marker for MS activity and progression

In CIS, the appearance of one or more T2 lesions predicts conversion to definitive MS (175), and the number of T2 lesions at disease onset may predict disability development (176). The disease activity measured by MRI is much higher than that estimated clinically. Monthly MRI performance shows approximately 10-fold more new lesions compared with the number of new relapses (177, 178). However, increasing lesion load is not a common feature of PMS. Disability progression is better correlated with atrophy measurements, such as brain parenchymal fraction on T1-weighted images (179). The annual rate of whole brain atrophy is approximately 0.5-1% in MS patients, compared to 0.2-0.5% in healthy individuals (180-182). Significant brain volume reduction is also evident early in the disease, involving both grey and white matter (183). Atrophy of the grey matter and spinal cord seem to predict disability progression (184, 185). Brain atrophy measurements have been proposed for predicting outcome following neuroprotective therapies in MS trials (186).

### 4.4 Biochemical biomarkers in MS

### 4.4.1 General considerations

Biomarkers are physical, functional, or biochemical indicators of physiological or disease processes; they can have diagnostic properties, provide information about the risk of disease development, reflect disease activity or disease severity, and have predictive or prognostic properties. They are also used to investigate responses to therapies, or discern adverse events and drug interactions. Additionally, biomarkers may be used to explore important pathophysiological mechanisms in disease progression. In clinical trials, biomarkers can act as surrogate endpoints, i.e. substitutes for clinical endpoints. New biomarker development involves many challenges, and the use of biomarkers carries the risk that a biomarker may not measure what it is supposed to and instead reflect other processes.

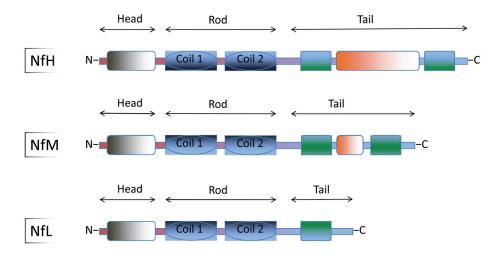
In MS, MRI measurements (discussed above) are the most frequently used biomarkers. However, CSF biomarkers are the most widely studied among the biochemical biomarkers in diseases affecting the CNS in general, and specifically in MS. MS is considered a CNS-specific disease and thus CSF is topographically near the disease process. However, the impact on the CSF differs depending on what CNS areas are affected. Frontal, parietal, or occipital regions of the cortex are considered CSF-distant, and pathological processes in these areas have less impact on CSF composition (187). The BBB creates an environment that is relatively isolated and partially protected from pathological processes affecting the rest of the body. However, about 80% of the protein content and all immune cells in the lumbar CSF are blood derived, and immune cells regularly migrate across the BBB in both directions. Under physiological conditions, blood-derived proteins enter the CSF compartment via passive diffusion and show a specific CSF-to-blood ratio (188), and these conditions change during some pathological processes.

Lumbar puncture side effects, including perceived discomfort of the patient, postpunctional headache, and minor risk for CNS infections or haematoma, makes CSF less attractive compared to blood. The development of blood-derived biomarkers for monitoring is therefore sought. Indeed, some serum and plasma biomarkers reflecting different immune mechanisms, show association with MS course or disease activity like osteopontin, LIGHT and metalloproteinases (189-192). However, structural biomarkers have been investigated in blood and the results to date have been inconclusive. For example, Eikelenboom et al. (2011) found no significant correlation between neurofilament heavy in blood and in CSF (193). Other studies have revealed diagnostic biomarkers for demyelinating diseases; aquaporin 4 antibodies have been included in the diagnostic criteria for neuromyelitis optica (194); and recently, KIR 4.1 antibodies were discovered in 50% of MS patients (195).

### 4.4.2 Structural biomarkers

### 4.4.2.1 Neurofilament protein

The neurofilament protein is the major component of the axonal cytoskeleton and is only found in nerve cells. Its function is to maintain the axonal structure, and it is essential in physiological processes like axonal transport (196). The neurofilament protein can be divided into three subunits: the 61-kDa neurofilament light (NFL) protein, the 103-kDa neurofilament medium (NFM) protein, and the 111-kDa neurofilament heavy (NFH) protein. The three subunits each have a common head and rod region, but differ in the tail region (196, 197) (*Figure 5*).



Schematic representation of the different neurofilament subunits. NFH: neurofilament heavy chain, NFM : neurofilament medium chain, NFL: neurofilament light chain, N: N-terminus, C: C-terminus. (199) Reprinted with permission from Sagepub.

These three subunits are assembled to interfilament structures of 8-10 nm. Neurofilaments are highly phosphorylated and the degree determines the three-dimensional structure and axonal diameter (198). During axonal damage, neurofilament leaks out into the surrounding tissue and further disperses into the CSF and blood. The pathological process of how the intermediate filaments are dissolved is not fully understood, and the further pathways of the three different subunits are uncertain but are clearly not identical (199, 200). While NFM remains poorly studied, the other two are increasingly valued as biomarkers of axonal damage in several neurological diseases. NFL levels in CSF are increased in many CNS diseases, such as herpes simplex virus encephalitis and tick borne encephalitis (201), cerebral vasculitis (202), atypical Parkinson disorders (PSP, MSA-C, MSA-P and CBD) (203), and ALS (204, 205). A number of studies have shown elevated neurofilament levels in MS. NFL levels are particularly elevated in active RRMS, early in the disease and during relapse but have been elevated in all courses of the disease (206-209). NFH is also elevated in patients with SPMS, and is related to disease progression and to brain atrophy on MRI (210, 211).

Neurofilament has also been experimentally measured in brain autopsy samples to verify axonal loss (112), and only been detected in blood (NFH) where massive axonal damage has occurred (212). The differing degrees of phosphorylation of the same protein in different situations and the similarities between the subunits in the

head region (213) create challenges in creating reliable assays that are free from cross reactions (214).

## 4.4.2.2 Glial fibrillary acidic protein (GFAP)

Glial fibrillary acidic protein (GFAP) is the major intermediate filament of the astrocyte cytoskeleton. It is thought to maintain the mechanical strength and outer shape of the astrocyte, and is essential for the processes of reactive astrogliosis and glial scar formation (215). Although GFAP has been isolated in various tissues, it is generally considered a CNS-specific protein (216, 217). GFAP consists of three domains: a head, rod, and tail. It can form homodimers as well as heterodimers with other proteins (218), it has a molecular weight of 43-49 kDa, and the proteins are arranged in fibre bundles of 8-9 nm (219). GFAP was originally isolated and characterized in MS lesions with severe astrogliosis (220). Elevated CSF GFAP levels are observed in pathological CNS conditions, such as dementia (221) and normal pressure hydrocephalus (222). In MS, augmented CSF GFAP levels are related to relapse (223) and a correlation has been observed between GFAP levels and disability in SPMS patients (208). Increased serum levels of GFAP are seen in patients with severe head injury (224) and are related to outcome in subarachnoid haemorrhage (225).

#### 4.4.2.3 Albumin ratio

Albumin ratio (CSF/plasma albumin) is used as a measure of BBB function (226). Albumin is a liver-produced, 67-kDa heavy plasma protein that passively diffuses over the BBB. The ratio is positively correlated to age, and is also elevated in several pathological processes (227). Most MS patients have albumin ratio values below the upper reference level (228); thus, this measure is not used as an independent marker in MS. However, the BBB affects the measurements of other biochemical markers, such as intrathecal IgG production (229).

## 4.4.3 Inflammatory biomarkers

#### 4.4.3.1 CXCL13

Cytokines are small proteins (~25 kDa) that function as mediators for intercellular communication. CXCL13 is 8-10 kDa, and is the 13th identified member of the cys-x-cys (CXC) motif ligand cytokine sub-family of the 17 members described in humans. This protein is also called ANGIE, ANGIE2, "B-cell chemoattractant", BCA-1, BLC, and BLR1L.

CXCL13 is selectively chemotactic for B cells in the B1 and B2 subsets, and stimulates the chemokine receptor CXCR5 (230). CXCL13 is produced by a number of cell types; it seems to control the organization of B cells within follicles in lymphoid tissue (231) and to indirectly influence immunoglobulin synthesis. From T-cell lymphomas, it is known that T cells can express CXCL13, reflecting their germinal centre origin (232). Elevated CXCL13 levels in CSF are seen in infectious diseases, such as Lyme borreliosis (233) and viral encephalitis (234). In MS, elevated levels of CSF CXCL13 reportedly predict conversion from CIS to definite MS (235). The levels are elevated during relapse, and correlate to IgG synthesis and the appearance of OCB (234, 236).

## 4.4.3.2 Oligoclonal IgG bands (OCB)

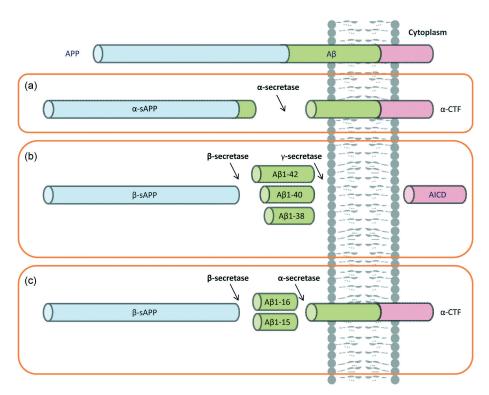
IgG in OCB produced in the CNS is characteristic of MS (237) and found in almost all patients (>95%) (228, 238, 239). In physiological processes, OCB are produced by clonally expanded plasma cells and plasma blasts (240, 241). Although target antigens for CSF antibodies in MS patients have been isolated, the congruence and pathological relevance of these antibodies remain obscure. Until recently, OCB were used as a diagnostic marker for all types of MS; however, the latest revision of the diagnostic criteria only uses OCB as a diagnostic marker for PPMS (63).

## 4.4.3.3 IgG Index

IgG produced in the CNS is the oldest identified biomarker in MS (242). To distinguish IgG produced outside the BBB, the IgG index is calculated as follows: (CSF IgG (mg/L)/serum IgG (mg/L))/(CSF albumin (mg/L)/serum albumin (mg/L)), thus compensating for the BBB function. An elevated IgG index is seen in 70-90% of MS patients, and is usually normal in OCB-negative patients (243-245). Use of the IgG index has been limited since its sensitivity is lower than that of OCB. In spite of its close connection to OCB, the IgG index is more closely related to immune activity (124, 234, 246) and treatment effects (247); therefore, the IgG index has been valued as a marker of immune suppression and treatment effect (247).

## 4.5.4 Biomarkers of the sAPP/Aβ metabolism

Amyloid precursor protein (APP), its degradation products, and the involved enzymes have been most extensively studied in Alzheimer's disease, where the CSF levels provide diagnostic information (248). The *figure 6* below describes the three known APP processing pathways, which are dependent on the involvement of three secretases:  $\alpha$ ,  $\beta$ , and  $\gamma$ .



APP processing pathways.

(a)  $\alpha$ -Secretase cuts APP within the A $\beta$  domain, inhibiting A $\beta$  formation and releasing  $\alpha$ -sAPP. (b) APP may be sequentially processed by BACE1 at the N-terminal portion of A $\beta$  and by  $\gamma$ -secretase at the C-terminal. In fact,  $\gamma$ -secretase may cut APP at different positions, producing A $\beta$  peptides with different C-terminal amino acids. Other enzymes are involved in the production of A $\beta$  peptides having other N-terminal amino acids (for example: A $\beta$ X-42, A $\beta$ X-40 and A $\beta$ X-38). (c) APP may undergo combined  $\alpha$ - and  $\beta$ -secretase processing, producing a range of short A $\beta$  peptides. A $\beta$ : amyloid beta; APP: amyloid precursor protein; BACE1:  $\beta$ -site APP cleaving enzyme 1; C-terminal: the acid end of an amino-acid molecule; CTF: C-terminal fragment; N-terminal: the amino end of an amino-acid molecule. Paper III Reprinted with permission from Sagepub.

#### 4.5.4.1 β-site APP cleaving enzyme, BACE1 (β-secretase)

As shown in the above figure, BACE1 ( $\beta$ -secretase) cleaves APP, resulting in the release of N-terminal  $\beta$ -cleaved soluble APP ( $\beta$ -sAPP). The C-terminal fragment is further processed by  $\gamma$ -secretase to yield different A $\beta$  peptides and the APP intracellular domain (AICD), or by  $\alpha$ -secretase to form shorter fragments. APP can also undergo  $\alpha$ -secretase-mediated cleavage, resulting in  $\alpha$ -sAPP release (249). Studies in BACE1-knockout mice suggest that this enzyme has various functions besides

its role in APP metabolism, including cell differentiation, immunoregulation, and a physiological role in cleaving neuregulin 1 (NRG1) (250, 251). NRG1 is an axonalexpressed membrane-bound factor required for CNS myelination (252). BACE1mediated release of the soluble ectodomain of NRG1 is required for the activation of myelination by oligodendrocytes (250).

Increased BACE1 levels have been reported in Alzheimer's disease and in persons with mild cognitive impairment (253, 254), as well as in traumatic brain injury and ischemic brain injury (255, 256). Little is known about BACE1 activity related to MS.

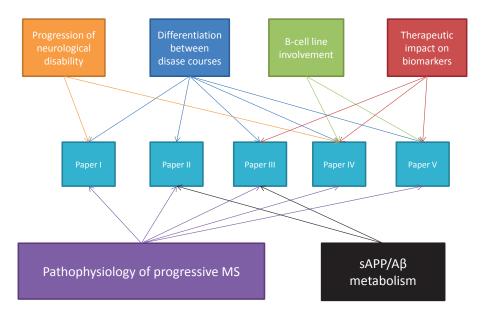
## 4.5.4.2 sAPP ( $\alpha$ -sAPP and $\beta$ -sAPP) and the A $\beta$ peptides

In Alzheimer's disease, the A $\beta$  1-42 levels are reduced, while  $\alpha$ -sAPP and  $\beta$ -sAPP are unaffected (257). Reduced CSF levels of A $\beta$  peptides as well as  $\alpha$ -sAPP and  $\beta$ -sAPP are observed in inflammatory conditions of the CNS, such as Lyme disease (258), opportunistic cerebral infections due to HIV (259), bacterial meningitis (260), and systemic lupus erythematosus (SLE) (261). Little is known about how APP metabolism in the CNS is affected in MS, and the CSF markers have not been systematically evaluated.

## 5. Aims of the study

The purpose of this project was to investigate if biomarkers in the CSF of MS patients

- can differentiate between different clinical courses of MS
- are associated with disease activity
- · are associated with progression of neurological disability in MS
- are useful for monitoring therapeutic efficacy in different stages of MS
- may provide new insights into the pathophysiology of MS progression.



## Investigation of biomarkers in MS Major purposes of the papers

## 6. Subjects and methods

## 6.1 MS populations

This thesis includes three major cohorts of MS patients, of which the majority were consecutively recruited at the Department of Neurology, Sahlgrenska University Hospital, Gothenburg, Sweden. The first cohort consisted of patients included in a cross-sectional investigation of the sAPP/Aβ pathway in MS (Paper II); patients in this cohort were also recruited from two other MS centres: Akershus University Hospital, Oslo, Norway and Karolinska University Hospital, Stockholm, Sweden. The second cohort comprised a sub-group of patients from the first cohort, who were additionally examined in long-term follow-up setting (Papers I, II, III, and V). The third cohort consisted of patients who were assessed before and after 12-24 months of immunosuppressive or second-line immunomodulatory treatment (Papers III, IV, and V); patients in this cohort were also recruited from two other MS centres: Umeå University Hospital, Umeå and Örebro University Hospital, Örebro, both in Sweden.

All MS patients recruited before 2001 were diagnosed according to the Poser criteria (62). Patients recruited thereafter were diagnosed according to the criteria of McDonald (262) and later revisions (63, 263). Informed consent was obtained from all subjects, and studies were approved by the regional ethical boards of Gothenburg and Uppsala.

## 6.1.1 Patients included in the cross-sectional study of the sAPP/A $\beta$ pathway (Paper II)

This cross-sectional study included 100 patients (33 men and 67 women) from three different populations (Gothenburg, Oslo, and Stockholm). The population from Gothenburg consisted of 66 patients (25 men and 41 women) and has been described previously (208). This group comprised 23 RRMS patients who were included at the time of an acute relapse, 18 RRMS patients in clinical remission, and 25 SPMS patients. The mean age was 39.5 years (range, 17-59 years), mean disease duration was 14.3 (range, 1-40 years), and median EDSS was 3.0 (range, 1.0-8.0).

The population from Oslo consisted of 21 patients (3 men and 18 women) with a mean age of 42 years (range, 27-65 years), mean disease duration was 5 years (range, 0.3-16 years), and median EDSS was 2.0 (range, 0-5.0). Fourteen had RRMS and 7 had PPMS.

The population from Stockholm constituted 13 patients (4 men and 9 women) with a mean age of 39 years (range, 25-52 years), mean disease duration of 3.2 years

(range, 1-14 years), and median EDSS of 1.0 (range, 0-4.5). Eleven had RRMS and two had SPMS.

## 6.1.2 MS patients with long-term follow-up (Papers I, II, III, and V)

Twenty-six patients (18 women and 8 men) were recruited from a previous study population of 66 MS patients (208). At their first assessment, the mean age was 41 years (range, 21-59 years) and mean disease duration was 11 years (range, 1-40 years). Each patient was assessed twice with a median interval of 9 years (range, 8-10 years). All had a clinically stable course and none had a relapse within 6 months prior to either assessment. At the first assessment, 16 patients were classified as RRMS and 10 as SPMS. At the second assessment, six additional patients were classified as SPMS. None of the patients were receiving disease-modifying treatment at the first assessment (five interferon beta and one glatiramer acetate). Their median EDSS was 3.25 (range, 1.0-8.0) at the first assessment and 5.25 (range, 0-9.5) at the second. From this long-term follow-up cohort, 25 patients were examined for Paper I, 23 patients for Paper II, 26 patients for Paper III, and 20 patients for Paper V.

## 6.1.3 MS patients treated with immunosuppressive or second-line immunomodulatory treatment (Papers III, IV, and V)

6.1.2.1 Patients treated with immunosuppressive therapies (Papers III, IV, and V) After publication of the OLYMPUS study (104), we modified the patient inclusion criteria for mitoxantrone treatment from the MIMS study (94), expanding the population to also include patients treated with rituximab. The cohort included 38 patients (19 women and 19 men); three had RRMS, 30 had SPMS, and five had PPMS. Fourteen patients had previously been treated with interferon beta and one with glatiramer acetate. At the pre-treatment assessment, the mean age was 49 years (range, 24-65 years), mean disease duration was 14 years (range, 2-29 years), and median EDSS was 6.0 (range, 3.0-8.0). After 12-24 months of immunosuppressive treatment (33 patients with mitoxantrone and 5 with rituximab), the median EDSS was 6.0 (range, 3.0-8.5). From this cohort, 20 mitoxantrone-treated patients (18 with SPMS and 2 with RRMS) were included in Paper III, and 10 of them (9 with SPMS, and 1 with RRMS) were re-examined post-treatment. In Paper IV, patients with RRMS (n = 3) were excluded, and Paper V included 22 mitoxantrone-treated patients (3 RRMS, 15 SPMS, 4 PPMS).

## 6.1.2.2 Second-line immunomodulatory treatment (Paper III)

Forty-one RRMS patients (14 men and 27 women) were assessed prior to natali-

zumab treatment. Five patients were previously untreated, whereas 36 had breakthrough disease with relapses or MRI activity while receiving other immunomodulatory treatment (23 with interferon beta, 8 mitoxantrone, 4 glatiramer, and 1 fingolimod). These patients comprised a subgroup of a previously described population (264). They were all regarded as highly active, and were scheduled to start monthly intravenous treatment with 300 mg natalizumab according to Swedish guidelines (96) At first assessment, their mean age was 36 years (range, 13-60 years) and mean disease duration was 8.5 (range, 0.5-26 years). At both assessments, their median EDSS was 3.5 (range, 0-6.5).

## 6.2 Control subjects

This thesis includes three major control populations; 82 healthy controls (HC), 32 spinal anaesthesia subjects (SAS), and 67 patients with systemic lupus erythematosus (SLE). Informed consent was obtained from all control subjects, and studies were approved by the regional ethical boards of Gothenburg and Uppsala.

The first cohort consisted of HC and SAS who were included in a cross-sectional investigation of the sAPP-A $\beta$  pathway in MS (Paper II). The majority were recruited at the Sahlgrenska University Hospital, Gothenburg, Sweden. HC were also recruited from the MS centre at Akershus University Hospital, Oslo, Norway. The second cohort was a sub-group of HC from the first cohort, who were further investigated in a long-term follow-up setting (Papers I-V). The third cohort consisted of patients with SLE (Paper II) recruited at the Department of Rheumatology, Sahlgrenska University Hospital, Sweden.

## 6.2.1 HC and SAS included in the cross-sectional study of the sAPP-A $\beta$ pathway (Paper II)

Our cross-sectional study included 114 HC and SAS (46 men and 68 women) with a mean age of 42 years (range, 18-82 years). The population consisted of 50 HC from the MS Centre, Gothenburg (208), 22 HC from the MS Centre, Oslo, and a mix of 42 HC and SAS from other departments at the Sahlgrenska University Hospital. The subjects included blood donors, students (recruited by advertisement), spouses of patients, and a group of neurologically healthy individuals undergoing lower limb surgery, which was previously described (265).

## 6.2.2 HC population with long-term follow-up (Papers I-V)

Twenty-eight HC (20 men and 8 women) were included from a previously described control population of 50 healthy blood donors and students (208). Their mean age

was 33 years (range, 18-53 years). They were subjected to lumbar puncture twice with a median interval of 9 years.

## 6.2.3 Control population with systemic lupus erythematosus (Paper II)

Sixty-seven SLE patients (10 men and 57 women) were included. Their mean age was 42 years (range, 17-75 years). They were recruited at the Department of Rheumatology, Sahlgrenska University Hospital, and were diagnosed according to the revised SLE criteria of The American Rheumatism Association (266). Twenty-six of these subjects were considered to fulfill the criteria for cerebral SLE (267, 268).

	MS patients				C	ontrol subject	s
	Long-term follow-up	Cross- sectional			Long-term follow-up	Cross- sectional	SLE
		oootionai	Mx or Rx*	Nz**		HC/SAS	
Paper I	25	0	0	0	28	0	0
Paper II	23	100	0	0	27	82/32	67
Paper III	26	0	20	41	28	0	0
Paper IV	0	0	35	0	14	0	0
Paper V	20	0	22	0	26	0	0

Table 1. Overview of the number of patients and control subjects from Papers I-V.

Mx, mitoxantrone; Rx, rituximab; Nz, natalizumab.

\*Immunosuppressive therapies.

\*\*Second-line immunomodulatory therapy.

**Table 2.** Demographics and clinical characteristics of the long-term follow-up cohort of patients (n = 25) and HC (n = 28) at baseline from Paper I.

Study group	AII MS	RRMS	SPMS	Healthy controls
Subjects, n	25	15	10	28
Gender, F/M	18/7	14/1	4/6	8/20
Mean age (range), years	41 (21-59)	40 (21-59)	43 (27-52)	33 (18-53)
Median EDSS (range)	3.0 (1.0-8.0)	2.75 (1.0-4.5)	5.5 (2.0-8.0)	NA

NA, not applicable.

**Table 3.** Demographics and clinical characteristics of the long-term follow-up cohort of patients (n = 23) and HC (n = 27), and the cross-sectional cohort of patients (n = 100), HC/SAS (n = 114), and SLE (n = 67) at baseline from Paper II.

Group	MS	SLE	Healthy controls
Subjects, n	100	67	114
Gender, F/M	33/67	10/57	46/68
Mean age (range), years	42 (17-65)	42 (17-75)	42 (18-82)
Median EDSS (range)	3.0 (0-8.0)	NA	NA

Some patients and HC were initially included in the cross-sectional study and thereafter included in the long-term follow-up cohort.

NA, not applicable.

**Table 4.** Demographics and clinical characteristics of the long-term follow-up cohort of patients (n = 26) and HC (n = 28), and patients of the treated cohort (natalizumab n = 41, mitoxantrone n = 20) at baseline from Paper III.

Group	Subjects, n	Gender F/M	Age, Median (IQR), years	Baseline EDSS Median (IQR)
Study populati	on at baseline			
RRMS	54	39/15	37 (32-46)	3.5 (2.0-4.5)
SPMS	33	17/16	53 (43-58)	6.0 (4.8-7.0)
Controls	28	7/21	40 (35-52)	NA
Subpopulation Natalizumab	of natalizuma	b and mitoxar 21/8	ntrone treated patie	ents 3.5 (2.0-6.0)
Mitoxantrone	9 SPMS	3/7	45 (38-50)	· · · · ·
	1 RRMS		10 (00 00)	6.0 (4.5-6.4)
Subpopulation	_	immunoprec	ipitation and mass	
Subpopulation RRMS	_	immunoprec 6/4		
	analysed with		ipitation and mass	spectrometry

IQR, interquartile range; NA, not applicable.

**Table 5.** Demographics and clinical characteristics of the treated cohort (mitoxantrone or rituximab, n = 35) and 14 HC of the long-term cohort from Paper IV.

Group	MS patients	HC
Subjects, n	35	14
Gender, F/M	15/20	5/9
Mean age, (range), years	48 (22-65)	42 (31-61)
EDSS, median (range)	6.0 (3.0-8.0)	NA

NA, not applicable.

**Table 6**. Demographics and clinical characteristics of the long-term follow-up cohort of patients (n = 20) and HC (n = 26), and patients of the treated cohort (mitoxantrone, n = 22) from Paper V.

Group 1	Subjects, n	Gender F/M	Age <sup>*</sup>	EDSS Baseline <sup>**</sup>
All MS	20	15/5	41.5 (21-59)	3.0 (1.0-8.0)
RRMS	9	9/0	33.7 (21-48)	3.0 (0-8.0)
SPMS	7	3/4	46.7 (41-52)	7.5 (2.0-8.0)
Transitory	4	3/1	50 (44-59)	3.25 (2.0-4.0)
Group 2	n	Gender F/M	Age*	EDSS Baseline**
All MS	22	11/11	45.2 (22-60)	6.0 (3.0-8.0)
RRMS	3	3/0	42.6 (37-50)	6.0 (4.5-6.5)
SPMS	15	7/8	46.1 (22-60)	6.0 (3.0-8.0)
PPMS	4	1/3	43.5 (34-52)	6.5 (6.0-7.5)
Group Controls	n	Gender F/M	Age*	EDSS Baseline**
HC	26	8/18	33.1 (18-60)	NA

\*Data are mean (range), years.

\*\*Data are median (range).

# 6.3 Clinical assessments, MRI, and serum and CSF sampling

Disease activity was measured by annual relapse rate (Papers I-V) or presence of  $Gd^+T1$ -weighted images (Paper IV). A relapse was defined as patient-reported or objectively observed events typical of an acute inflammatory demyelinating event in the CNS, current or historical, with duration of at least 24 hours, in the absence of fever or infection (63). Neurological disability was determined by EDSS and its seven functional systems (152), and disease severity or progression was scored according to progression index (158) or MSSS (156).

The MRI investigations (Paper IV) were performed according to the standard MS protocols of Sahlgrenska University Hospital, Örebro University Hospital, and Umeå University Hospital. The sequences used in the standard program are T1, T1 with standard-dose gadolinium contrast, T2, DWI, and Flair. MRI scans were performed with 1.5 or 3.0 Tesla machines, using 3-mm slice thickness, and took place no more than three months from clinical neurological examination and lumbar puncture. Serum samples were obtained by venepuncture and CSF samples were obtained by lumbar puncture; samples were handled and stored according to the recommendations of Bio MS (269). The clinical assessments and the serum and CSF sampling were performed once in the subjects of the cross-sectional cohorts, and twice in subjects of the long-term follow-up and the treated cohorts, with intervals of 8-10 years and 12-24 months, respectively.

## 6.4 Assays

All biochemical analyses in the studies of this thesis were run at the laboratory of neurochemistry, Sahlgrenska University Hospital, Mölndal, Sweden. They were performed by experienced and certified laboratory technicians who were blinded to the clinical and MRI data. To minimize variation, baseline and follow-up samples were analysed side by side on each assay plate, using one batch of reagents. All intra assay coefficients of variation were < 10%.

## 6.4.1 Polyclonal NFL assay (Paper I)

NFL was measured by an in-house-developed sandwich ELISA (205). Rabbit polyclonal anti-NFL IgG was used as the secondary antibody. Bound secondary antibody was detected using peroxidase-conjugated donkey anti-rabbit IgG. The standard curve ranged from 125 to 16 000 ng/L. The assay sensitivity was 125 ng/L.

## 6.4.2 Monoclonal NFL assay (Paper IV)

NFL levels in CSF were determined by an enzyme-linked ELISA developed by the UMAN Diagnostics. CSF samples were incubated in pre-coated anti-NFL plates. As secondary antibodies, we used two monoclonal antibodies that are highly specific for NFL. Streptavidin horse radish peroxidase was used for detection. The assay sensitivity was 31 ng/L (209).

## 6.4.3 Glial fibrillary acidic protein assay (Papers I and IV)

GFAP was measured using an in-house developed ELISA, as previously described (270). Rabbit polyclonal anti-GFAP IgG was used as the secondary antibody. Captured secondary antibody was detected using peroxidase-conjugated donkey anti-rabbit IgG. The procedure was performed at room temperature. The sensitivity of the GFAP assay was 32 ng/L. The standard curve ranged from 32 to 16 000 ng/L.

## 6.4.4 CXCL13 (Papers IV and V)

CXCL13 was measured by an ELISA Human CXCL13/BLC/BCA-1 Immunoassay (R&D Systems Inc., Abingdon, United Kingdom) according to the manufacturer's instructions. In brief, this assay employs a sandwich enzyme immunoassay technique. A monoclonal antibody specific for CXCL13 was precoated on a microplate. HRP-labelled CXCL13 monoclonal soluble detection antibody was used. The quantification limit was 7.8 pg/mL, and CSF samples below that level were designated as 3.9 pg/mL.

## 6.4.5 Albumin ratio (Paper V)

Quantitative determinations of albumin in serum and CSF were performed by immunonephelometry on a Beckman Image Immunochemistry system (Beckman Instruments, Beckman Coulter, Brea, CA, USA). Briefly, the nephelometer measured the increased light scattered as a result of particles created after addition of antialbumin antibodies to a solution. The albumin ratio was calculated as CSF albumin (mg/L)/serum albumin (g/L)  $\times$  1000.

## 6.4.6 IgG index, CSF-specific oligoclonal IgG bands (Paper V)

Quantitative determinations of IgG in serum and CSF were performed by immunonephelometry on a Beckman Image Immunochemistry system (Beckman Instruments, Beckman Coulter, Brea, CA, USA). The IgG index was calculated as CSF IgG (mg/L)/serum IgG (mg/L)/albumin ratio. OCB were visualized in CSF and serum by isoelectric focusing followed by silver staining, as previously described (271). Ocular quantification of CSF-specific OCB was performed independently by two experienced researchers. Patients were divided into five different groups according to the number of OCB: 0, 1, 2-4, 5-10, and >10.

## 6.4.7 BACE1 (β-Secretase ) activity (Paper II)

CSF BACE1 activity was measured using a highly sensitive and specific solutionbased assay, as previously described (272). Briefly, the assay consisted of an enzymatic reaction of CSF with a biotinylated 15-mer optimized BACE1 substrate (biotin-KTEEISEVNFEVEFR) and the use of pepstatin A to block non-specific protease activity, followed by cleavage product detection using a neoepitope-specific antibody. The extent of BACE1 activity was quantified using recombinant BACE1 standards. The sensitivity of the assay was below 1.0 pM of recombinant BACE1.

## 6.4.8 $\alpha$ -sAPP and $\beta$ -sAPP (Papers II and III)

The CSF  $\alpha$ -sAPP and  $\beta$ -sAPP were determined using the sAPP $\alpha$ /sAPP $\beta$  Multiplex Assay for Meso Scale Discovery (MSD) platform (MSD, Gaithersburg, MD, USA), as described by the manufacturer. Briefly, this assay employed antibodies specific for the two fragments, run at the same time in two different electrodes coated with antibodies for 6E10 (an epitope within amino acids 3-8 of the A $\beta$ sequence) to capture  $\alpha$ -sAPP, and a neoepitope-specific antibody on the C-terminal to capture  $\beta$ -sAPP. Both also had matching control electrodes. Both species were detected by the SULFO-TAG-labelled anti-APP antibody, p2-1. The sensitivity of this method was 120 pg/mL for  $\alpha$ -sAPP, and 52 pg/mL for  $\beta$ -sAPP.

## 6.4.9 A\u03c6X-38, A\u03c6X-40, and A\u03c6X-42 (Papers II and III)

CSF A $\beta$ X-38, A $\beta$ X-40, and A $\beta$ X-42 were determined using the MSD A $\beta$  Triplex kit (Human A $\beta$  peptide Ultra-Sensitive Kits), as described by the manufacturer. Briefly, this assay ran three samples at the same time on three attached electrodes. The N-terminus included 4G8 antibodies (an epitope within amino acids 18-22 of the A $\beta$  sequence) as capturing antibodies, while the C-terminus included SULFO-TAG-labelled specific antibodies for detecting the different A $\beta$  peptides. The method sensitivity was 6 pg/mL for A $\beta$ X-38, 2 pg/mL for A $\beta$ X-40, and 7 pg/mL for A $\beta$ X-42.

## 6.4.10 Aβ1-42 (Papers II and III)

CSF A $\beta$ 1-42 was determined using the INNO-BIA AlzBio3 kit (Innogenetics, Ghent, Belgium) on the xMAP Luminex platform, as described previously (273). Briefly, one monoclonal antibody (4D7A3) was attached to a capturing microsphere, and a second antibody (3D6) was attached to the opposite end. Both ends included antibodies for establishing the sequence 1-42. The flourochrome phycoerythrin was

coupled to streptavidin to measure the amount of A $\beta$ 1-42 on each microsphere. The method was flow cytometric, allowing microspheres with different antibodies to be run at the same time. Each microsphere was separated based on different signals after laser excitation. The sensitivity of the method was 25 ng/L.

## 6.4.11 Immunoprecipitation and mass spectrometry (Paper III)

This analysis used immunoprecipitation (IP) followed by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOFMS) (274, 275). Briefly, aliquots of the monoclonal antibodies 6E10 and 4G8 were separately added to magnetic beads coated with sheep anti-mouse IgG (Life Technologies, Carlsbad) and incubated. 6E10 + 4G8 antibody-bead solutions and Tween in phosphate-buffered saline (PBS) were added to CSF samples. The solution was then transferred to a magnetic particle processor (KingFisher Flex; Thermo Scientific, Waltham, MA, USA) for automatic washing and elution. The collected supernatant was dried by speed-vac and redissolved in formic acid (FA) in acetonitrile (ACN). The MALDI samples were prepared with the seed layer method, and analysed by MALDI-TOFMS (Bruker AutoFLEX (Bruker Daltonics). Each sample was analysed in duplicate. The mass/charge ratio (m/z) used to identify the A $\beta$ -peptide sequences represents the monoisotopic peak of the protonated molecule [M+ H<sup>+</sup>].

## 6.4.12 Liquid chromatography and tandem mass spectrometry (Paper III)

To confirm the identities of the A $\beta$  species, the immunoprecipitates (IP) were analysed by liquid chromatography (LC) combined with high-resolution tandem mass spectrometry (MS-MS) (276). Briefly, LC-MS/MS analysis was performed on a chromatographic system (GE Healthcare) using HotSep Kromasil C4 columns (G&T Septech) coupled to a mass spectrometer (Thermo Fisher Scientific). Collision-induced dissociation (CID) was used to obtain fragment ion data.

## 6.5 Statistics

Statistical inference was performed using SPSS software (several versions, updated over time), PASW Statistics 18 software (Paper IV), GraphPad Prism 5.00 (GraphPad Software, San Diego, CA, USA) (Paper III), and Excel 2007 (Microsoft Corporation, Redmond, WA, USA) (Paper III).

Normally-distributed data were analysed by parametric tests. Between-group comparisons were made using Student's *t*-test, ANOVA, and ANCOVA. Within-subject differences over time were analysed by paired *t*-test. Correlation analyses were performed using Pearson correlation, linear regression analysis, partial correlation analysis, and one-way ANOVA. Data with skewed distributions were analysed using non-parametric tests. Between-group differences were analysed by Mann-Whitney U test, Kruskal-Wallis test, and non-parametric ANCOVA. The Wilcoxon rankedsign test was used for within-subject differences over time. The correlations between variables were calculated using Spearman's rank correlation.

## 6.5.1 Multivariate analyses

In Paper III, multivariate analysis was used to differentiate two groups based on their biomarker profiles. Each marker formed an orthogonal projection, which combined to form an algorithm. This algorithm determines the direction (score vector) in the multivariate orthogonal space spanned by biomarkers that provides the best separation between the predefined groups. A stepwise variable reduction was performed until only the markers with positive contributions toward the separation were left, as judged by their relative contributions to the model. The calculations were implemented using the SIMCA P+ v. 12 software (Umetrics, Umea, Sweden).

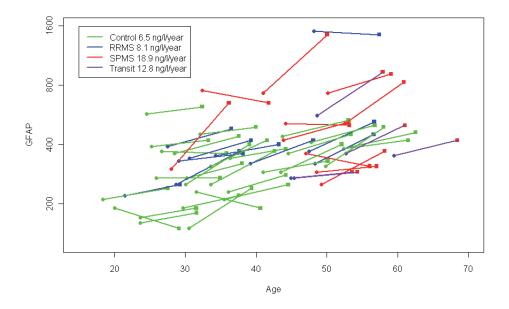
Corrections for multiple comparisons were done using Bonferroni-correction. The data were reported as the mean when presumed to be normally distributed, otherwise the median was given.  $P \le 0.05$  indicated statistical significance and  $P \le 0.01$  highly significance.

## 7. Results

## 7.1 Paper I

#### 7.1.1 Increased CSF GFAP levels in MS patients

With adjustment for age, increased CSF GFAP levels were recorded in patients compared to in HC (P < 0.05), whereas similar CSF NFL levels were observed in patients and HC. No significant differences were found between the RRMS and SPMS patients. GFAP and NFL levels were both correlated with age (both r = 0.50, P < 0.01) (*Figure 7*).



#### Figure 7

Individual GFAP levels at two occasions for patients with relapsing-remitting multiple sclerosis (RRMS, n = 10), secondary progressive multiple sclerosis (SPMS, n = 10), patients converting from RRMS to SPMS (Transit, n = 5), and healthy controls (n = 28). The first examination is indicated with circles and the second examination with squares. The mean annual GFAP increase is indicated for each group

#### 7.1.2 CSF GFAP correlated with progression

When the data were corrected for age, CSF GFAP levels increased between the assessments (P < 0.05). This increase was most prominent in the SPMS group. The mean annual GFAP increases were 6.5 ng/L for HC, 8.1 ng/L for RRMS patients,

12.8 ng/L for patients converting from RRMS to SPMS, and 18.9 ng/L for SPMS patients. However, this progressive trend was not accompanied by significant between-group differences.

## 7.1.3 GFAP correlated with clinical disability and was closely related to neurological disability

GFAP levels correlated with neurological disability as measured by EDSS (r = 0.51, P < 0.05), and with disease severity or progression determined by MSSS (r = 0.47, P < 0.05). Dividing disability into the seven FS of EDSS revealed significant correlations between GFAP and the FS scores of pyramidal (r = 0.51, P < 0.01), bowel and bladder (r = 0.51, P < 0.01), and brain stem (r = 0.53, P < 0.01). EDSS (P < 0.01) and progression index (P < 0.01) increased between assessments, mainly driven by increases in the scores of SPMS and patients changing course from RRMS to SPMS during long-term follow-up.

## 7.1.4 GFAP had predictive value

GFAP level at the first assessment had predictive value for neurological disability (EDSS) 8-10 years later (r = 0.45, P < 0.05). This was most pronounced in the SPMS group (r = 0.80, P < 0.01). However, GFAP level could not predict the increase of EDSS between the two assessments. The group with high GFAP levels (>350 ng/L) had a higher risk of achieving high EDSS (>5.5), with an OR of 4.5.

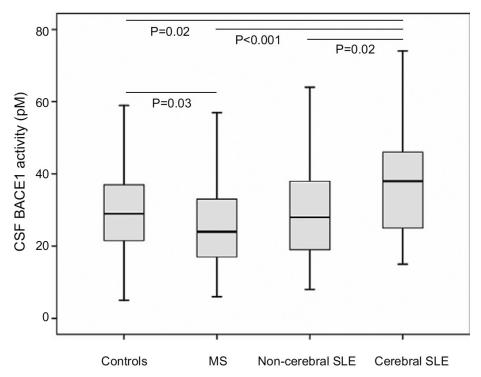
## 7.1.5 NFL was not elevated in clinically stable MS but some patients showed signs of subclinical disease activity

NFL levels were not increased in MS patients compared to in HC; they had no relationship with disability or progression, and no prognostic value for disability development. Two patients had increased NFL levels on only one occasion, and three patients showed high levels on multiple occasions, indicating acute axonal damage not associated with relapse.

## 7.2 Paper II

## 7.2.1 BACE1 distinguished MS from other inflammatory disease and controls

MS patients had lower CSF BACE1 activity than controls (P < 0.05) and patients with cerebral SLE (P < 0.01) (*Figure 8*). Patients with cerebral SLE had higher BACE1 activity than any other group (P < 0.05 for all comparisons). SPMS patients tended to have lower levels compared to other MS groups (P < 0.07). In MS, BACE1 activity correlated negatively with disease duration (r = -0.26, P = 0.01) and neurological disability as determined by EDSS (r = -0.25, P < 0.05).



CSF BACE1 activity in patients with MS, patients with SLE with and without cerebral involvement, and controls.

## 7.2.2 BACE1 activity correlated with APP metabolites that were altered in MS

In MS patients, BACE1 activity correlated weakly with levels of sAPP/A $\beta$  markers (A $\beta$ 42,  $\alpha$ -sAPP, and  $\beta$ -sAPP). Subgroup analysis revealed stronger correlations in SPMS patients (A $\beta$ 42: r = 0.40, P < 0.05;  $\alpha$ -sAPP: r = 0.66, P < 0.01;  $\beta$ -sAPP: r = 0.77, P < 0.01) than in RRMS patients (A $\beta$ 42: r = 0.38, P < 0.01;  $\alpha$ -sAPP: r = 0.40, P < 0.05). These correlations with A $\beta$ 42,  $\alpha$ -sAPP, and  $\beta$ -sAPP, respectively, were also significant in non-cerebral (r = 0.54, r = 0.66, r = 0.64, all P < 0.01) and cerebral (r = 0.75, r = 0.55, r = 0.54, all P < 0.01) SLE. In controls, only a weak correlation was found between BACE 1 and  $\beta$ -sAPP (r = 0.19, P < 0.05).

The CSF A $\beta$ 42,  $\alpha$ -sAPP, and  $\beta$ -sAPP levels were significantly reduced in patients with MS compared to in controls *(Table 7)*. However, the reductions in  $\alpha$ -sAPP and  $\beta$ -sAPP were not specific to MS but were also seen in cerebral and non-cerebral SLE *(Table 7)*. Compared with MS patients in remission, patients with on-going

or recent relapse (within one month) had lower levels of  $\alpha$ -sAPP [respectively, 476 ng/L (range, 265-1195 ng/L) and 711 ng/L (range, 186-1564 ng/L); P < 0.01] and  $\beta$ -sAPP [respectively, 239 ng/L (range, 146-559 ng/L) and 301 ng/L (range, 105-463 ng/L); P < 0.01].

 Table 7. Demographics and biomarker concentrations in MS, cerebral SLE, noncerebral SLE, and controls

	MS ( <i>n</i> = 100)	Cerebral SLE (n = 26)	Noncerebral SLE ( $n = 41$ )	Controls (n = 114)
Age, years	42 (17–65)	41 (19–75)	43 (17–75)	42 (18–82)
Gender, men/women	33/67	3/23	7/34	46/68*
α-sAPP, ng/mL	582 (186–1564)**	585 (142–1106)†	510 (234–1285)†	669 (238–1727)
β-sAPP, ng/mL	260 (105–559) <sup>‡</sup>	225 (110–395)†	210 (112–447)†	285 (93–615)
Aβ42, pg/mL	648 (220–1118) <sup>†</sup>	707 (345–1182)	632 (262–952)‡	722 (140–1132)

MS, multiple sclerosis; SLE, systemic lupus erythematosus.

Data are median (range).

\*P < 0.001 versus MS, cerebral and noncerebral SLE.

\*\*P < 0.001 versus controls.

 $^{\dagger}P < 0.01$  versus controls.  $^{\ddagger}P < 0.05$  versus controls.

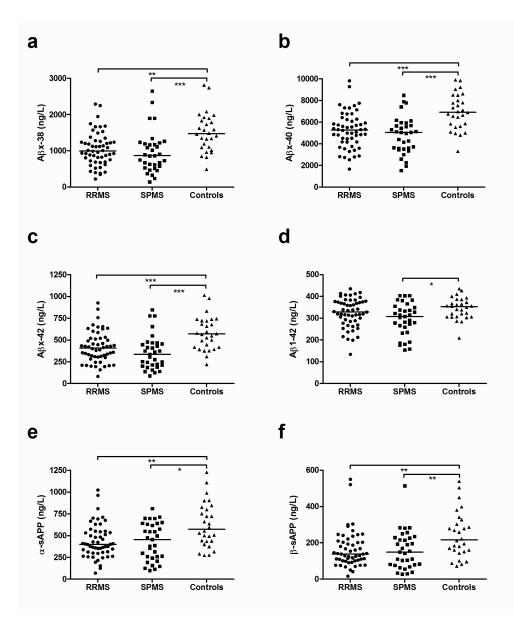
**7.2.3 BACE1 activity decreased towards progressive disease course** BACE1 activity decreased over time in MS patients (P < 0.05) who were assessed twice with an interval of 8-10 years. This decrease was most pronounced in RBMS

twice with an interval of 8-10 years. This decrease was most pronounced in RRMS patients, whereas SPMS patients had constantly low activity. None of the markers showed prognostic abilities or relation to clinical disability (EDSS).

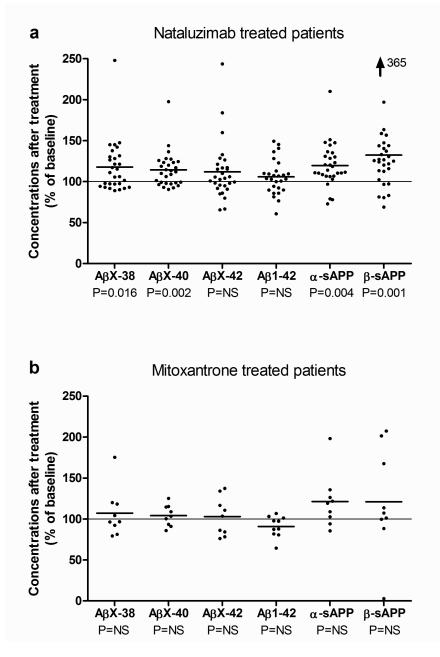
## 7.3 Paper III

## 7.3.1 The APP metabolites were decreased in MS patients

RRMS and SPMS patients had lower baseline CSF levels of A $\beta$ X-38, A $\beta$ X-40, A $\beta$ X-42,  $\alpha$ -sAPP, and  $\beta$ -sAPP than control subjects, while only SPMS patients showed lower A $\beta$ 1-42 level (*Figure 9*). No significant differences were found between RRMS and SPMS patients. Since the study groups differed in age and gender distribution, these factors were corrected for in the analysis.



Baseline levels of biomarkers, as measured by immunoassays. \* P<0.05, \*\* P<0.01 and \*\*\* P<0.001



Changes in biomarkers after treatment.

Patients treated with natalizumab (Panel (a)) and mitoxantrone (Panel (b)). The number of patients analyzed varied between biomarkers, due to missing samples (Panel (a), N = 27-29; Panel (b), N = 8-10). The graphs show that follow-up concentrations normalized in relation to baseline levels (100%).

## 7.3.2 Natalizumab normalized the APP metabolite levels

The APP metabolite levels were normalized in patients treated with natalizumab, with only two exceptions (A $\beta$  x-42 and A $\beta$  1-42), while no changes in APP metabolites were seen after treatment with mitoxantrone (*Figure 10*). However, patients treated with natalizumab were younger than those treated with mitoxantrone (*P* < 0.05). Within the natalizumab-treated group, older patients had a lower absolute change in  $\beta$ -sAPP compared to younger patients (r = 0.43, *P* < 0.05). The age differences seen between the groups may have influenced the  $\beta$ -sAPP trajectories in the two treatment groups; however, the overall dynamic biomarker response after natalizumab treatment did not seem to be confounded by age, and the correlation between age and  $\beta$ -sAPP was not significant after multiple comparisons.

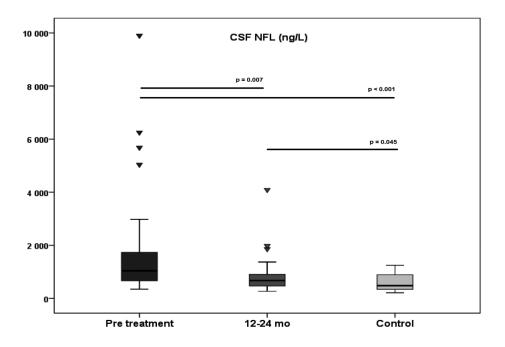
## 7.3.3 Progressive MS patients exhibited altered sAPP/Aβ metabolism

The CSF was analysed with IP-MS and mass spectrometry, and 18 different  $A\beta$  species were isolated. Only small univariate differences in the relative levels of individual peptides were observed between the disease courses. Multivariate analysis revealed distinctly different patterns of sAPP/A $\beta$  between SPMS and controls, with an intermediate pattern observed in RRMS.

## 7.4 Paper IV

## 7.4.1 NFL levels were reduced by immunosuppressive treatment in progressive MS patients

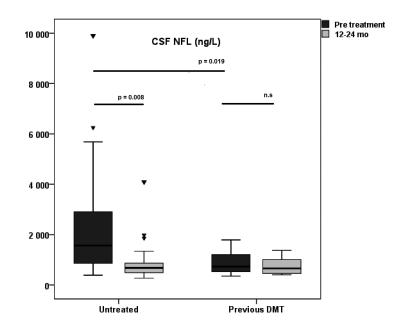
The mean NFL level in patients with progressive MS was reduced following 12-24 months of mitoxantrone (n = 30) or rituximab treatment (n = 5), from 1780 ng/L (SD, 2018) to 870 ng/L (SD, 693) (P < 0.01). However, these reduced NFL levels in patients did not reach those found in HC [577 ng/L (SD, 326); P < 0.05] (*Figure 11*).



Neurofilament light (NFL) in cerebrospinal fluid (CSF) before and following mitoxantrone (n=30) or rituximab (n=5) treatment. NFL levels in the CSF were compared to levels obtained in 14 healthy individuals. Box indicates interquartile range, bar indicates median, and whiskers indicating 95% CI.

## 7.4.2 First-line disease-modifying therapies seemed to influence the NFL levels in progressive MS

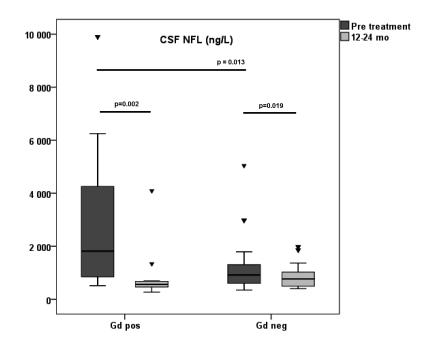
In previously untreated patients (n = 20), the pre-treatment mean NFL level (2462 ng/L; SD, 2452) was significantly higher than in patients with on-going first-line disease-modifying therapies (DMT) (874 ng/L; SD, 434; n = 15; P < 0.05) (*Figure 12*). Only the group of previously untreated patients showed significant NFL level reduction following 12-24 months of immunosuppressive treatment (P<0.01).



Neurofilament light (NFL) in cerebrospinal fluid (CSF) before and following 12-24 months of mitoxantrone or rituximab treatment in previously untreated patients (n=20) or patients with ongoing disease modifying therapies (DMT, n=15) at study baseline. Box indicates interquartile range, bar indicates median, and whiskers indicating 95% CI.

## 7.4.3 NFL levels seemed to influence Gadolinium enhancement on MRI in progressive MS

Progressive MS patients with Gd<sup>+</sup> lesions at study baseline (n = 12) presented with significantly higher pre-treatment NFL levels than patients without Gd<sup>+</sup> lesions on MRI (n = 23) [2925 ng/L (SD, 2884) vs. 1184 ng/L (SD, 1024); P < 0.05] (Figure 13). Only one patient with Gd<sup>+</sup> lesions had been previously treated. Although patients with Gd<sup>+</sup> lesions had the most marked NFL decrease [2925 ng/L (SD, 2884) vs. 889 (SD, 1041); P < 0.01], patients without signs of BBB damage on MRI also showed significantly reduced NFL levels following immunosuppressive treatment [1184 ng/L (SD, 1024) vs. 866 (SD, 449); P < 0.05].



Neurofilament light (NFL) in cerebrospinal fluid (CSF) before and following 12-24 months of mitoxantrone or rituximab treatment in patients with (n=12) or without (n=23) pretreatment gadolinium enhancing lesions on MRI. Box indicates interquartile range, bar indicates median, and whiskers indicating 95% CI.

## 7.4.4 CXCL13 levels in CSF of progressive MS were normalized by immunosuppressive treatment

CXCL13 levels increased to above the ELISA detection limit were only found in 11 of the 70 CSF samples obtained from progressive MS patients. The mean pretreatment CXCL13 level was significantly higher in MS patients (9.7 pg/mL) compared to in HC subjects (<7.8 pg/mL; P < 0.05). Moreover, the mean CXCL13 level at treatment follow-up was significantly reduced (4.4 pg/mL; P < 0.01). With one exception, all of the CSF samples with detectable CXCL13 levels also had high NFL levels (>1200 ng/L, i.e. 2 SD above the mean NFL level of HC).

## 7.4.5 GFAP levels were increased in progressive MS patients but unaffected by immunosuppressive treatment

GFAP levels were higher in progressive MS patients compared to HC [650 ng/L (SD, 240) vs. 490 ng/L (SD, 170); P < 0.05). Immunosuppressive treatment had no significant effect on GFAP levels at follow-up. However, mean GFAP level was

significantly higher in patients with pre-treatment Gd<sup>+</sup> compared to patients without signs of disease activity on MRI [803 ng/L (SD, 224) vs. 578 ng/L (SD, 216); P < 0.01].

## 7.4.6 NFL were correlated with CXCL 13, and GFAP in progressive MS

Pre-treatment CSF NFL levels were correlated significantly with CXCL13 (r = 0.527, P < 0.01) and GFAP levels (r = 0.471, P < 0.01). Following 12-24 months of immunosuppressive treatment, NFL levels were only correlated with GFAP levels (r = 0.440, P < 0.01). The pre- to post-treatment differences were significantly correlated between NFL and CXCL13 (r = 0.475, P < 0.01) and between NFL and GFAP (r = 0.391, P < 0.05).

## 7.5 Paper V

## 7.5.1 The OCB pattern changed over 8-10 years of follow-up

Of 18 patients examined twice with an interval of 8-10 years, 16 had  $\geq$ 2 OCB and two had one IgG band selectively produced in the CSF. Between the two examinations, the OCB pattern changed in 15 patients. Without any isolated predisposing factor, the number of bands increased in four patients and decreased in six, three patients both gained and lost bands, and two had unchanged OCB number but with differences in intensity.

## 7.5.2 Immunosuppressive treatment did not affect the OCB

Of the 22 patients examined before and after 1-2 years of mitoxantrone treatment, the OCB counts or patterns changed in four patients (all with SPMS). Two exhibited increased OCB and two demonstrated both new and lost bands. OCB counts were not correlated with clinical course, EDSS, or MSSS.

## 7.5.3 CXCL13 levels were decreased following immunosuppressive treatment of progressive MS

CXCL13 was analysed at the second assessment in the long-term follow-up group, and at both pre- and post-treatment assessments, performed 24 months apart. Patients of the long-term follow-up group had CXCL13 levels similar to those of controls. Among progressive patients, the pre-treatment CXCL13 levels were significantly higher than in HC [13.8 (SD, 1.22) vs. 4.6 (SD, 2.6); P < 0.01]. After 2 years of mitoxantrone treatment, the mean CXCL13 level was significantly reduced [4.3 (SD, 1.9); P < 0.01] to the levels of HC.

## 7.5.4 Correlations between OCB, IgG synthesis, CXCL13, and clinical parameters

In the long-term follow-up group, the IgG index was correlated with the numbers of OCB at both the first (r = 0.60, P < 0.01) and second assessments (r = 0.47, P < 0.01). No significant relationship was found between OCB and EDSS in SPMS. However, in RRMS patients (n = 9), EDSS correlated both with the OCB counts (r = 0.70, P < 0.05), and with the changed numbers of bands between the assessments (r = 0.68, P < 0.05).

In mitoxantrone-treated patients, the number of OCB correlated with the IgG index at both pre-treatment (r = 0.58, P < 0.01) and post-treatment (r = 0.45, P < 0.01) assessments.

CXCL13 level correlated with the IgG index in the long-term follow-up group (r = 0.51, P < 0.05), as well as in the mitoxantrone-treated group (r = 0.61, P < 0.01). CXCL13 levels also correlated with the number of OCB (r = 0.49, P < 0.05) in mitoxantrone-treated patients. We found weak correlations of CXCL13 levels with EDSS (r = 0.32, P < 0.05) and with MSSS (P = 0.32, P < 0.05) in both investigated groups.

## 8. Discussion

## 8.1 General considerations regarding biomarkers in MS

Recent years have seen the development of a growing number of therapeutic agents for MS, some of which have been introduced or are currently awaiting approval. There remains an obvious need for objective and reliable biomarkers to assess disease activity, rate of progression, and therapeutic efficacy. Investigations of biomarkers in MS may also reveal important information about pathophysiological processes. Currently available evidence indicates an indisputable role of inflammation in the demyelination and axonal damage in early RRMS, whereas immune activity is less clearly involved in neurodegeneration in PMS. A set of biomarkers reflecting different aspects of MS immunopathogenesis could be useful for further exploring this subject.

MRI measurements are currently the dominant biomarkers in MS. Visualization of focal inflammation can be used to confirm clinical or detect sub-clinical disease activity (277). MRI is less useful in PMS; hence, there is a great need for new biomarkers reflecting the pathological processes of this stage. Unlike MRI, biochemical biomarkers in CSF have the potential to specifically and directly mirror the immunopathological processes, indicating which inflammatory or degenerative pathway is activated. Additionally, our knowledge of the autoaggressive immune attack on the CNS in RRMS is strongly based on the animal model of experimental autoimmune encephalomyelitis (EAE), but no such reliable model exists for PMS. Therefore, investigations of CSF biomarkers may be of particular importance for resolving PMS pathophysiology. The present thesis discusses the value of biochemical CSF biomarkers as a complement to clinical and MRI investigations in PMS, particularly regarding their potential for monitoring disease activity, disability progression, disease severity, prognosis, and therapeutic efficacy.

# 8.2 CSF biochemical biomarkers related to PMS disease activity

A biochemical biomarker of MS disease activity should reflect the occurrence of relapse or the appearance of new lesions. Accumulating numbers of inflammatory biomarkers, e.g. cytokines or chemokines (234, 278-281), and CNS parenchymal biomarkers (206, 207, 209, 236, 282-284) have been associated with increased disease activity in MS. However, only few studies have shown an association between on-going inflammation and signs of CNS damage (206, 285).

## 8.2.1 Neurofilament light protein

Several studies have reported elevated CSF levels of neurofilament proteins during all stages of MS, with increased levels associated with relapse (207-209, 282, 286) and with Gd<sup>+</sup> lesions on MRI (206). In the present study, we detected increased NFL levels in PMS patients with signs of increased activity on MRI (Paper IV). Increased NFL was also related to CXCL13, suggesting increased immune activity in these patients. In RRMS, the CSF NFL level increases 3- to 10-fold during relapse (208, 209), then declines to low levels within 3 months (208, 286). In Paper IV, we found that PMS patients had increased NFL levels prior to immunosuppressive treatment, whereas the RRMS and SPMS patients from Paper I exhibited NFL levels similar to those observed in HC. These differences between studies may be partly due to the differences in patient selection. In Paper I, patients were included based on their low clinical disease activity, with no relapse documented within 3 months prior to study enrolment. In contrast, the PMS patients eligible for immunosuppressive treatment in Paper IV had obvious disability progression, with several of them showing subclinical disease activity on MRI. Furthermore, the NFL immunoassay used in Paper I had lower sensitivity compared to the NFL ELISA used in Paper IV. Therefore, low-grade NFL changes that were detected in Paper IV may have been below the detection limit of the assay used in Paper I.

#### 8.2.2 Glial fibrillary acidic protein

Previous studies have reported augmented CFS GFAP levels in RRMS, but these changes did not correlate with relapse rates (223). In Paper IV, we confirmed increased GFAP levels in PMS patients compared to in HC (208). Interestingly, we also found that patients with  $Gd^+$  lesions on MRI prior to immunosuppressive treatment had significantly higher GFAP levels compared to Gd-negative patients, indicating that GFAP was partially influenced by disease activity.

## 8.2.3 CXCL13

Previous studies have shown elevated CXCL13 in MS, and that CXCL13 level correlates with disease activity in RRMS (234, 236). In Paper IV, we found increased CXCL13 levels in PMS compared to in HC; however, only 11 of the 35 PMS patients had CXCL13 levels above the detection limit of the assay. The highest CXCL13 levels were found in patients without previous DMT and with Gd<sup>+</sup> lesions on MRI. The correlation between CXCL13 and disease activity reported for RRMS also seemed to be present in PMS. A clinically stable group of RRMS and SPMS patients without relapses occurring within 3 months prior to assessment (n = 20, Paper V) exhibited CXCL13 levels similar to those found in HC; again, only a minority (n = 3) of the 20 patients in this study had elevated CXCL13 levels. This finding is in

contrast with previous reports describing elevated CXCL13 levels in considerably higher proportions of patients (234, 236). This difference may be due to the lower proportion of immune active patients in our study populations, which comprised mainly progressive cases. The low frequency of CXCL13-positive CSF samples in our studies implies that the use of CXCL13 as a biomarker for PMS may be limited.

## 8.2.4 BACE1 activity and sAPP/Aβ metabolites

In the first study of these biomarkers, we showed significantly reduced CSF BACE1 activity (Paper II) and lower CSF sAPP and A $\beta$  metabolite levels (Papers II and III) in MS compared to in HC. The CSF concentrations of these biomarkers did not significantly differ between RRMS and SPMS. However, decreased levels of  $\alpha$ - and  $\beta$ -sAPP were found in CSF of RRMS patients during relapse, compared to in stages of remission (Paper II), suggesting that clinical disease activity in RRMS seemed to reduce CSF levels of  $\alpha$ - and  $\beta$ -sAPP.

# 8.3 Biochemical biomarkers in CSF reflecting disability development, progression, or severity of PMS

Ideally, biochemical biomarkers should predict and reflect disability progression and severity of MS. However, the search for suitable biomarkers is complicated by several unsolved questions regarding MS pathophysiology, as well as by the frequently weak reliability of clinical measurements of disability. Changes of MRI measurements are currently the dominant surrogate marker for MS progression. MRI can show correlations between T1 and T2 lesion volume (167), cerebral or cervical spinal cord atrophy (184, 185), and disability development in MS. However, few biochemical biomarkers in CSF have been convincingly associated with disability progression (209, 211, 223, 287) or with signs of degeneration on MRI (210).

In the present studies (Papers I-V), disability was evaluated by EDSS, which is the most widely used scale but has low inter- and intra-rater sensitivity (153, 154). To reduce EDSS variability in the present studies, EDSS scoring was performed by only a few experienced neurologists (154). Other limitations of EDSS include its bimodal character and the relative lack of linearity, which particularly affects the scoring of PMS. Since EDSS scores above 3.5 mostly rely on ambulation, patients may progress in disability without influencing the EDSS score. This makes EDSS unreliable, especially in studies of shorter duration. To some extent, this limitation was overcome by the long-term follow-up of patients in Papers I, II, and V.

Disease severity and disability progression rate were determined by MSSS and progression index, respectively. These methods are derived directly from the EDSS, and provided temporal and prognostic information about the patient.

## 8.3.1 Neurofilament light protein

We found no correlation between NFL and disability (EDSS) or progression (MSSS), and NFL showed no prognostic value (Papers I and IV). This was in contrast to the results of previous studies (209, 288), which obtained CSF for diagnostic reasons and found that NFL levels predicted disability assessed at 4 and 14 years later. However, the EDSS score from the diagnostic investigation was not reported, and hence the dependence of developed disease severity on early disability was not calculated. Nevertheless, these previous findings imply that the extent of early axonal damage predicts future disability. It should be noted that early disability is mainly dependent on the location of individual lesions, while disability during progression results from accumulated lesion volumes and the development of cerebral and spinal cord atrophy. Moreover, the NFL level in CSF is also dependent on the interval between relapse onset and lumbar puncture (208, 286). The patients from Paper I were subjected to long-term follow-up of 8-10 years; however, no predictive value of NFL was observed. As discussed in the previous section, this may have been due to the patient selection and the sensitivity of the NFL assay.

## 8.3.2 Glial fibrillary acidic protein

Our findings confirmed that patients with MS had increased GFAP levels compared to HC, and that the levels were particularly high in SPMS patients (208). Among MS patients, GFAP correlated to clinical disability (EDSS) and progression or severity (MSSS) (Paper I). Similar results were previously reported in patients with high EDSS (>6.5) (289). Here, we also found that GFAP had prognostic value for future disability development. The risk for high EDSS (>5.5) was increased (OR = 4.5) in patients with GFAP levels above 350 ng/L at the first assessment. However, this relationship was dependent on the EDSS found at the first assessment (Paper I).

In chronic neurological diseases, including PMS, sustained clinical disability is expected to be irreversible. High sustained EDSS can be achieved early in MS due to severe relapses, or can result from slow progression over several years. To compensate for this imbalance, disease duration is included in the calculations of progression index and MSSS, which should result in more accurate measurements of progression rate or disease severity. We found that GFAP also correlated with progression index and MSSS, and that the annual GFAP increase was twice as high in SPMS compared to RRMS.

In the long-term follow-up setting (Paper I), GFAP seemed to have both prognostic value and to be a biomarker for progression; however, this was not true in PMS patients subjected to immunosuppressive treatment during 12-24 months (Paper IV).

## 8.3.3 CXCL13

CXCL13 has been previously shown to exhibit a non-significant trend of correlating with clinical disability in RRMS (234). We found that CXCL13 weakly correlated with EDSS and MSSS in the long-term follow-up group and in patients about to start mitoxantrone treatment (Paper V). The prognostic value of CXCL13 was not measured because only a minority of the studied patients had detectable levels.

## 8.3.4 Intrathecal IgG synthesis

The clinical relevance of OCB and increased IgG index is unclear, apart from their diagnostic uses (290). In contrast to earlier studies (291), here we found that both the number of OCB and the change in OCB numbers were correlated with clinical disability (EDSS) in RRMS patients.

## 8.3.5 BACE1 activity and sAPP/Aβ metabolites

BACE1 activity and the sAPP/A $\beta$  metabolites were found to be decreased in MS, and were negatively correlated with disability (EDSS) and severity (MSSS) in SPMS but not in RRMS. In RRMS, BACE1 activity decreased after 8-10 years of follow-up.

# 8.4 CSF biochemical biomarkers for discriminating between MS clinical courses

At some point, each of the investigated CSF biochemical biomarkers showed significantly higher (NFL, GFAP, CXCL13, OCB, and IgG index) or lower (BACE1 activity and sAPP/A $\beta$  metabolites) levels in MS patients than in HC; however, none could differentiate between the clinical courses of MS. None of the studied biomarkers showed any distinct borders differentiating between RRMS and PMS, although some observations indicated a gradual change towards progressive course. GFAP levels tended to increase with disability progression, and the annual rate of GFAP increase seemed higher among SPMS patients than RRMS patients. Additionally, multivariate analysis revealed that the A $\beta$  metabolite patterns were altered in MS patients, and that SPMS patients could be distinguished from HC based on the pattern of APP degradation products. RRMS patients seemed to constitute an intermediate group, but further discriminations were not possible with so few patients. These studies represent promising attempts to identify biomarkers that detect patients who are entering MS progression.

## 8.5 Biomarkers for determining therapeutic efficacy in MS

In MS, therapeutic intervention with immunomodulatory or immunosuppressive therapies can change or inhibit immune activity and, if successful, reduce damage and degenerative processes of the CNS. MRI evidence of lesion formation, lesion load, and atrophy development are used as end-points in MS trials, and no biochemical biomarkers have been accepted as surrogate markers for evaluating therapeutic efficacy in MS. Investigating biochemical biomarkers during therapeutic intervention could increase our knowledge of the possible mechanism of action of the applied treatments, as well as of the pathophysiology of the disease. Such biomarkers may also be able to serve as objective measures in conjunction with clinical and MRI measures for monitoring therapy efficacy.

In the present thesis, we investigated the effects of natalizumab (Paper III), mitoxantrone (Papers III, IV, and V) and rituximab (Paper IV) on a number of biomarkers. We also observed the treatment effects of first-line disease modifying treatment (DMT) (interferon beta and glatiramer acetate) in PMS (Paper IV).

#### 8.5.1 Neurofilament light protein

In a previous study of relapsing MS (n = 83), 6-12 months of natalizumab treatment reduced NFL levels to those found in HC (264). Natalizumab treatment also affected NFL levels in 9 SPMS patients with prior superimposed relapses. In Paper IV, we showed that 12-24 months of mitoxantrone or rituximab treatment reduced CSF NFL to levels approaching those observed in age-matched HC subjects. This effect was not confined to patients with Gd+ activity on MRI, but was also found in patients with increased NFL levels in the absence of Gd+ lesions at treatment onset. Pre-treatment NFL levels from patients who were previously treated with DMT were significantly lower than those from patients who were untreated at study baseline, suggesting that first-line DMT might influence NFL levels in PMS.

## 8.5.2 Glial fibrillary acidic protein

In patients with PMS, immunosuppressive treatment with mitoxantrone or rituximab had no significant impact on GFAP levels. However, PMS patients with Gd+ activity on MRI had increased GFAP levels prior to immunosuppressive treatment. Their GFAP levels were reduced post-treatment, indicating a treatment effect only in active PMS.

## 8.5.3 CXCL13 and intrathecal IgG synthesis

PMS patients in Paper IV and SPMS patients in Paper V showed reduced CXCL13 levels after 12-24 months of immunosuppressive treatment; IgG production and

OCB were not affected (Paper V). To our knowledge, this result has not been previously shown.

We confirmed that the number and pattern of OCB were unchanged, even following potent immunosuppressive treatment (292, 293). Natalizumab treatment decreases IgG synthesis (294) and, to date, is the only treatment shown to reduce the proportion of OCB-positive patients (294, 295).

## 8.5.4 sAPP/Aβ metabolites

In RRMS patients, natalizumab treatment increased sAPP/A $\beta$  metabolites (except A $\beta$  1-42 and x-42) towards normal levels. On the other hand, after 12-24 months of mitoxantrone treatment, 9 SPMS patients and 1 RRMS patient exhibited no changes in A $\beta$  and sAPP metabolites.

## 8.6 Biochemical biomarkers for exploring the pathophysiology of progressive MS

## 8.6.1 Axonal degeneration

An early phenomenon in MS, axonal loss and degeneration is observed in all MS courses (112), and has been established as the major cause of cerebral and spinal cord atrophy and disability progression in MS (115, 296). The highest levels of neurofilament protein occur during clinical or MRI signs of activity (206, 285). However, high neurofilament levels have also been observed in a plethora of diagnoses and pathological processes, such as ischemic stroke (297); CNS infections (201) and other autoimmune CNS diseases (268); and neurodegenerative diseases, such as ALS (205) and atypical Parkinson disorders (PSP, CBD, MSA-C, and MSA-P) (203).

We observed significantly increased NFL levels in PMS, with the highest levels in those with Gd+ lesions on MRI (Paper IV). Elevated NFL levels were associated with increased CXCL13 levels. Previous reports have shown similar associations between inflammatory biomarkers (CXCL13, osteopontin and MMP9) and NFL levels (207). In a previous study, we observed normalized NFL levels following 6-12 months of natalizumab treatment of RRMS (264), and in Paper IV we showed that immunomodulation/suppression also reduced axonal damage in PMS. Overall, these findings indicate that the increased NFL release in CSF is largely mediated by the on-going CNS inflammation. However, it cannot be ruled out that other non-inflammatory degenerative mechanisms may also participate in PMS evolution.

### 8.6.2 Astrogliosis

Astrocyte-mediated gliosis is a physiological reaction that is secondary to all forms of CNS damage (298). In MS, GFAP is considered the morphological basis of astrogliosis, and is the major constituent of chronic MS lesions (219). GFAP is considered a valuable astrogliosis biomarker and is elevated in MS, with the highest levels occurring during SPMS (208). Our results imply that astrogliosis increases with MS progression, but this process seemed to be essentially unresponsive to immunosuppressive treatment (Paper IV).

### 8.6.3 The role of the B cell lineage in PMS (CXCL13, IgG index, and OCB)

In recent years, evidence has accumulated indicating the importance of B cells in MS pathogenesis. One important finding is the reduction of MS disease activity achieved by anti-CD20 therapy (299-301), which results in B cell depletion. However, despite the B cell eradication in both the periphery and the CNS (101), IgG production in the CNS was not affected (293). Additionally, ectopic B cell follicles have been found in the meninges of PMS patients (129, 130, 302), which are reportedly associated with a gradient of subpial cortical demyelination, neuronal loss, and cortical atrophy (130). These follicles are aggregates of B cells and plasma cells, and may explain the maintained CSF IgG production in OCB that is observed in PMS (1).

Increased intrathecal IgG synthesis, including presence of OCBs, is observed in >95% of MS patients and has diagnostic value (63, 228, 239, 290). In Paper V, we showed that the OCB patterns changed over the long-term follow-up, and were not related to disease course. This was in contrast to the widespread opinion that OCBs are stable over the course of MS, and represent an immunological "fingerprint" for each patient (290). This concept was based on a number of studies that have mostly been small, short in follow-up, and have used different methodologies for CSF OCB detection (247, 303-305). Our findings are consistent with the different OCB patterns found in different MS plaques from the same brain (306), and suggest a dynamic shift of clonally expanded cells of the B cell lineage over the course of MS. Another explanation could be the rearranged immunoglobulin genes with extensive mutations that have been observed in CSF B cells (307) and in the brains of MS patients (308). Such mutations may be antigen driven (309), and the exposure of new antigens following inflammation and degeneration might induce new mutations of immunoglobulin genes and possibly explain the altered OCB patterns.

We did not find that OCB or IgG production were influenced by immunosuppressive treatment (Paper V). This result was in line with reports from previous studies that OCB patterns were stable after potent immunomodulatory or immunosuppressive treatments, such as rituximab (293) and hematopoietic stem cell transplantation

(292). Only natalizumab treatment has been shown to induce OCB disappearance in MS (294, 295). Mitoxantrone did not alter IgG production, despite its powerful ability to depress both B and T cells. Mitoxantrone does not cross the BBB, which supports the suspicion that an autonomous and long-lived B cell/plasma cell population is compartmentalized in the CNS.

CXCL13 is a chemokine that can attract and maintain B cells within the CSF/CNS. Here we confirmed previous findings that the CXCL13 level in the CSF was associated with IgG synthesis (234). These results support a B cell dependence of IgG synthesis. Furthermore, the complexity of this system is illustrated by the fact that only CXCL13, and not IgG synthesis, was affected by mitoxantrone.

### 8.6.4 BACE1 activity and sAPP/Aβ metabolism

In Paper II, we illustrated for the first time that BACE1 activity and sAPP/A $\beta$  metabolites are involved in MS pathophysiology. The  $\beta$ -pathway has been considered to be of special interest in studies of MS, since animal studies demonstrated that BACE1 cleaves neuregulin 1 from its membrane-bound form to stimulate myelination by oligodendrocytes. Demyelination and defective remyelination is a dominant feature in MS (148). Here we found reduced levels of BACE1 in MS patients compared to in controls and in patients with cerebral SLE used as inflammatory controls (Paper II); this difference was most pronounced in patients with long-term SPMS. While there is no clear direct parallel between myelination in mouse studies and remyelination in humans, these findings suggest one possible explanation for the altered oligodendrocyte function observed in late MS (250, 310). These results cannot be considered a general phenomenon due to CNS inflammation, since cerebral SLE demonstrated significantly increased CSF BACE1 levels.

We observed reduced levels of sAPP/A $\beta$  in the CSF of MS patients (Papers II and III), which has also been recently confirmed by other groups (311, 312). Based on our findings and the available literature, we concluded that these reduced levels were a result of general neuroinflammation (258-261). Although it is unclear why these levels decrease during inflammation, possible explanations include decreased production, increased aggregation, or increased clearance. However, amyloid plaques are uncommon in MS (313). Neuronal A $\beta$  release is related to synaptic activity (314), and CSF A $\beta$  levels correlate to CSF levels of granins (315), which are released during regulated neuronal secretion. Since oligodendrocytes initiate myelination preferentially on active axons, the decreased A $\beta$  levels might represent parts of a vicious circle of axon inactivation (316) and disturbed remyelination. Our findings in Paper III are consistent with a reversible neuronal impairment early in the disease (reversible reductions of sAPP/A $\beta$  peptides in RRMS) and irreversibility during the progressive course (irreversible reductions of sAPP/A $\beta$  peptides in SPMS).

# 9. Concluding remarks and future considerations

The pathophysiology of MS is complex, prominently featuring numerous active mechanisms, such as demyelination, astrogliosis, and axonal degeneration. It is obvious that no biomarker alone can describe this. The aim of this thesis was to investigate if biomarkers in the cerebrospinal fluid (CSF) of MS patients could provide new insights into the pathophysiology of MS progression, and if biomarker levels could reflect disease activity, disability progression, or therapeutic efficacy.

We found elevated levels of NFL, illustrated ongoing axonal damage, in PMS patients in fast progression; most pronounced in patients with  $Gd^+$  lesions on MRI without first line DMT (Paper IV). This was in contrast to findings in clinically stabile patients at long-term follow-up, who had levels as HC (Paper I). We found thus that the axonal damage was determined by the disease activity regardless of disease course.

In long-term follow-up (Paper I) GFAP was confirmed as a biomarker reflecting disability and disease severity in PMS. GFAP also showed a prognostic value for future disability. These findings were in line with accumulating astrogliosis in PMS.

The increased levels of NFL and CXCL13 in PMS were associated with disease activity and the levels decreased after immune suppressive and immune modulatory treatment. These biomarkers are potential tools for monitoring therapeutic efficacy.

We found signs of B-cell activity in all stages of MS (Paper IV and V). CXCL13 was associated to disease activity and axonal damage and also to the IgG production in CNS and OCB. We illustrated, in opposite to the general view, changed number and pattern of OCB in CSF over time (Paper V). These findings illustrated an on-going and a dynamic B-cell activation in all courses of MS and a B-cell dependence of intrathecal IgG synthesis.

We showed for the first time an alteration of the sAPP/A $\beta$  metabolism in MS (Paper II and III). This was illustrated both by the reduced activity of BACE1 and by the reduced levels of sAPP/A $\beta$  metabolites in CSF and are possible signs of impaired neuronal activity in the CNS. The reduction of the sAPP/A $\beta$  metabolism was reversible in the RRMS after immune modulating treatment but irreversible in SPMS after immune suppressive treatment. We concluded this as a potential sign of irreversibility of neuronal impairment late in the disease. In PMS the levels of degradation products was not only reduced but the pattern of metabolites was also changed compared to HC. This was visualized by mass spectrometry.

All data in the present thesis illustrated a relationship between CNS inflammation and neurodegeneration in all courses of MS; however, it cannot be ruled out that other non-inflammatory degenerative mechanisms may also participate in PMS evolution.

Investigated biomarkers in CSF provide important information about the pathophysiology in PMS, and may serve as complement to clinical and MRI measures for assessment of disease activity, progression, severity and therapeutic efficacy.

NFL has previously been investigated in a number of studies and its relevance as a disease activity marker was confirmed in these studies also in PMS. NFL could be evaluated for use as a biomarker for monitoring treatment effect and could be suggested as an endpoint in phase II clinical trials. By combining NFL, CXCL13, GFAP and evaluation of disease activity and MRI a more complete evaluation of treatment effects could be achieved. Further investigations are needed to evaluate the correlation between CSF biomarkers and degeneration parameters on MRI, as T1 lesion volume and atrophy. The sAPP/A $\beta$  metabolism are warranted to be further investigated for evaluation of the clinical usefulness of these markers and further investigate the mass spect profiles of the A $\beta$  metabolites in CSF on larger patient populations. Access to these biomarkers also in blood should be of clinical importance but demands further development of more sensitive analysing methods.

## Acknowledgements

**Associate Professor Jan Lycke,** my scientific supervisor and a clinical role model. For introducing me to the MS field clinically and scientifically. Your sharp reflections, conclusions and your brilliance in the art of writing are a huge inspiration to me. Your energy and patience were essential for the completion of this project. You have stood by me all the way with honesty and heartfelt sincerity. It is an honor to work with you.

**Professor Henrik Zetterberg,** my inspiring co-supervisor. For making me understand some of the mysteries of laboratory medicine and for your enthusiasm for research.

**Clas Malmeström,** my colleague, co-author, room mate and friend. You have been a source of endless help. I hope I will have you as a colleague for many years to come.

**Niklas Mattsson,** my co-author and friend. Your brain works at double the speed of any normal person. You travel two times around the world in the time it takes me to get to work. I hope for more opportunities to work with you.

**Professor Lars Rosengren**, my co-author and former head of the Department of Neurology. Thank you for sharing your scientific knowledge.

**Mikael Edsbagge**, Head of the Department of Neurology, for signing the right papers and giving me the conditions in which to practice this science.

Martin Gunnarsson, my co-author and friend. Thank you for your support and your wonderful sense of humor.

To all my co-authors especially **Anders Svenningsson**, **Kristin Augutis** and **Sara Haghighi** who through energy and knowledge have contributed a lot to these studies.

To all co-workers at the laboratory of neurochemistry especially **Professor Kaj Blennow** and **Shirley Fridlund** for support and knowhow in the lab.

Friends and colleagues at the department of Neurology in particular **Marie Sjögren**, **Albert Hietala, Daniel Jons, Peter Vaghfeldt** and **Björn Runmarker** for help and encouragement when so was needed.

**Snezana, Ulla, Birgitta, Ann, Anne, Ann-Christine, Ann-Helene, Belinda** and all other former and present co-workers at the MS team for resolving challenges on the way and creating a pleasant atmosphere. To **Sirpa** for all your help and for the emptiness you left behind.

**Professor Oluf Andersen,** for clinical and scientific inspiration and for knowing everything about neurology.

The professors at the Institute of Clinical Neuroscience and Rehabilitation Lars Rönnbäck, Kristina Malmgren and Carsten Wikkelsö for your enthusiasm and encouragement.

**Head of the institute, Professor Agneta Holmäng,** for creating excellent working conditions at the Institute of Neuroscience and Physiology.

Oscar and Markus for excellent help with computers that did not work.

My lovely parents, **Rolf** and **Ingela** for your endless support through life. I hope I have made you proud of me for being me.

My brothers **Magnus**, **Hans** and **Lars** for being just that and for making research something simple and accessible. We share blood and I love you.

My parents in-law **Erling** and **Liv** who, in spite of hard times, never hesitated to come to my support.

The **Dahlberg** and **Ekenstråle** families for understanding and supporting when the need arose.

**Torunn,** my wonderful wife, who has stood by and supported me through the years and helped me to my hold my course. I hope I will get the opportunity to show you my love and gratitude for the rest of my life.

**Oscar, Anna, Carl and Ingrid,** my amazing children who remind me of the meaning of life everyday and for making me see myself without needing a mirror. Thank you for being understanding of me being away without understanding what I am doing. I love you.

The brave and kind people participating in these studies we called **patients**. Without you none of this would have been possible. This work I dedicate to you.

The works in this thesis were supported by grants from the NHR Foundation, the Edith Jacobsson Foundation, Anna-Lisa and Bror Björnsson Foundation, the Swedish Association of Persons with Neurological Disabilities, the Swedish Research Council, the Göteborg Medical Society, the Västra Götaland Region, Alzheimerfonden, Stiftelsen för Gamla Tjänarinnor, Med-Coast, Swedish Brain Power, the Åke Wiberg Foundation, Soderberg Foundation, Alzheimer's Association, Swedish Brain Power, Swedish State Support for Clinical Research, Sahlgrenska Academy, Lundbeck Foundation, Stiftelsen Gamla Tjanarinnor, Uppsala Universitets Medicinska Fakultet, Swedish Brain Fund, Goteborgs lakaresallskap, Thureus stiftelse, Pfannenstills stiftelse, Demensfonden, the Research Foundation of the Multiple Sclerosis Society of Gothenburg and Biogen Idec

### References

- Frischer JM, Bramow S, Dal-Bianco A, Lucchinetti CF, Rauschka H, Schmidbauer M, et al. The relation between inflammation and neurodegeneration in multiple sclerosis brains. Brain. 2009 May;132(Pt 5):1175-89. PubMed PMID: 19339255. Pubmed Central PM-CID: 2677799. Epub 2009/04/03.
- Kutzelnigg A, Lucchinetti CF, Stadelmann C, Bruck W, Rauschka H, Bergmann M, et al. Cortical demyelination and diffuse white matter injury in multiple sclerosis. Brain. 2005 Nov;128(Pt 11):2705-12. PubMed PMID: 16230320. Epub 2005/10/19.
- Prineas JW, Kwon EE, Cho ES, Sharer LR, Barnett MH, Oleszak EL, et al. Immunopathology of secondary-progressive multiple sclerosis. Ann Neurol. 2001 Nov;50(5):646-57. PubMed PMID: 11706971. Epub 2001/11/15.
- 4. Semra YK, Seidi OA, Sharief MK. Heightened intrathecal release of axonal cytoskeletal proteins in multiple sclerosis is associated with progressive disease and clinical disability. J Neuroimmunol. 2002 Jan;122(1-2):132-9. PubMed PMID: 11777552. Epub 2002/01/05.
- 5. Prineas JW, Parratt JD. Oligodendrocytes and the early multiple sclerosis lesion. Ann Neurol. 2012 Jul;72(1):18-31. PubMed PMID: 22829266. Epub 2012/07/26.
- 6. Fawcett JW, Asher RA. The glial scar and central nervous system repair. Brain Res Bull. 1999 Aug;49(6):377-91. PubMed PMID: 10483914. Epub 1999/09/14.
- Dalton CM, Brex PA, Jenkins R, Fox NC, Miszkiel KA, Crum WR, et al. Progressive ventricular enlargement in patients with clinically isolated syndromes is associated with the early development of multiple sclerosis. J Neurol Neurosurg Psychiatry. 2002 Aug;73(2):141-7. PubMed PMID: 12122170. Pubmed Central PMCID: 1737988.
- Schumacker GA, Beebe G, Kibler RF, Kurland LT, Kurtzke JF, McDowell F, et al. Problems of Experimental Trials of Therapy in Multiple Sclerosis: Report by the Panel on the Evaluation of Experimental Trials of Therapy in Multiple Sclerosis. Ann N Y Acad Sci. 1965 Mar 31;122:552-68. PubMed PMID: 14313512. Epub 1965/03/31.
- Tumani H, Hartung HP, Hemmer B, Teunissen C, Deisenhammer F, Giovannoni G, et al. Cerebrospinal fluid biomarkers in multiple sclerosis. Neurobiol Dis. 2009 Aug;35(2):117-27. PubMed PMID: 19426803.
- Ahlgren C, Oden A, Lycke J. High nationwide prevalence of multiple sclerosis in Sweden. Mult Scler. 2011 Aug;17(8):901-8. PubMed PMID: 21459810. Epub 2011/04/05.
- Sundstrom P, Nystrom L, Forsgren L. Incidence (1988-97) and prevalence (1997) of multiple sclerosis in Vasterbotten County in northern Sweden. J Neurol Neurosurg Psychiatry. 2003 Jan;74(1):29-32. PubMed PMID: 12486262. Pubmed Central PMCID: 1738162.
- Svenningsson A, Runmarker B, Lycke J, Andersen O. Incidence of MS during two fifteen-year periods in the Gothenburg region of Sweden. Acta Neurol Scand. 1990 Sep;82(3):161-8. PubMed PMID: 2270743.
- Hernan MA, Olek MJ, Ascherio A. Geographic variation of MS incidence in two prospective studies of US women. Neurology. 1999 Nov 10;53(8):1711-8. PubMed PMID: 10563617. Epub 1999/11/24.
- 14. Simpson S, Jr., Blizzard L, Otahal P, Van der Mei I, Taylor B. Latitude is significantly associated with the prevalence of multiple sclerosis: a meta-analysis. J Neurol Neurosurg Psychiatry. 2011 Oct;82(10):1132-41. PubMed PMID: 21478203. Epub 2011/04/12.

- 15. Midgard R. Incidence and prevalence of multiple sclerosis in Norway. Acta Neurol Scand Suppl. 2012 (195):36-42. PubMed PMID: 23278655.
- Pugliatti M, Sotgiu S, Rosati G. The worldwide prevalence of multiple sclerosis. Clin Neurol Neurosurg. 2002 Jul;104(3):182-91. PubMed PMID: 12127652.
- Wallin MT, Culpepper WJ, Coffman P, Pulaski S, Maloni H, Mahan CM, et al. The Gulf War era multiple sclerosis cohort: age and incidence rates by race, sex and service. Brain. 2012 Jun;135(Pt 6):1778-85. PubMed PMID: 22628389.
- Orton SM, Herrera BM, Yee IM, Valdar W, Ramagopalan SV, Sadovnick AD, et al. Sex ratio of multiple sclerosis in Canada: a longitudinal study. Lancet Neurol. 2006 Nov;5(11):932-6. PubMed PMID: 17052660. Epub 2006/10/21.
- Trojano M, Lucchese G, Graziano G, Taylor BV, Simpson S, Jr., Lepore V, et al. Geographical Variations in Sex Ratio Trends over Time in Multiple Sclerosis. PLoS One. 2012;7(10):e48078. PubMed PMID: 23133550. Pubmed Central PMCID: 3485003. Epub 2012/11/08.
- Kotzamani D, Panou T, Mastorodemos V, Tzagournissakis M, Nikolakaki H, Spanaki C, et al. Rising incidence of multiple sclerosis in females associated with urbanization. Neurology. 2012 May 29;78(22):1728-35. PubMed PMID: 22592376. Epub 2012/05/18.
- Orton SM, Ramagopalan SV, Brocklebank D, Herrera BM, Dyment DA, Yee IM, et al. Effect of immigration on multiple sclerosis sex ratio in Canada: the Canadian Collaborative Study. J Neurol Neurosurg Psychiatry. 2010 Jan;81(1):31-6. PubMed PMID: 19710047. Epub 2009/08/28.
- Willer CJ, Dyment DA, Risch NJ, Sadovnick AD, Ebers GC. Twin concordance and sibling recurrence rates in multiple sclerosis. Proc Natl Acad Sci U S A. 2003 Oct 28;100(22):12877-82. PubMed PMID: 14569025. Pubmed Central PMCID: 240712. Epub 2003/10/22.
- Compston A, Coles A. Multiple sclerosis. Lancet. 2008 Oct 25;372(9648):1502-17. Pub-Med PMID: 18970977.
- Gourraud PA, Harbo HF, Hauser SL, Baranzini SE. The genetics of multiple sclerosis: an up-to-date review. Immunol Rev. 2012 Jul;248(1):87-103. PubMed PMID: 22725956. Epub 2012/06/26.
- International Multiple Sclerosis Genetics C, Wellcome Trust Case Control C, Sawcer S, Hellenthal G, Pirinen M, Spencer CC, et al. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. Nature. 2011 Aug 11;476(7359):214-9. PubMed PMID: 21833088. Pubmed Central PMCID: 3182531.
- Healy BC, Liguori M, Tran D, Chitnis T, Glanz B, Wolfish C, et al. HLA B\*44: protective effects in MS susceptibility and MRI outcome measures. Neurology. 2010 Aug 17;75(7):634-40. PubMed PMID: 20713950. Pubmed Central PMCID: 2931768.
- Yeo TW, De Jager PL, Gregory SG, Barcellos LF, Walton A, Goris A, et al. A second major histocompatibility complex susceptibility locus for multiple sclerosis. Ann Neurol. 2007 Mar;61(3):228-36. PubMed PMID: 17252545. Pubmed Central PMCID: 2737610.
- Sawcer S, Hellenthal G, Pirinen M, Spencer CC, Patsopoulos NA, Moutsianas L, et al. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. Nature. 2011 Aug 11;476(7359):214-9. PubMed PMID: 21833088. Pubmed Central PMCID: 3182531. Epub 2011/08/13.

- International Multiple Sclerosis Genetics C, Hafler DA, Compston A, Sawcer S, Lander ES, Daly MJ, et al. Risk alleles for multiple sclerosis identified by a genomewide study. N Engl J Med. 2007 Aug 30;357(9):851-62. PubMed PMID: 17660530.
- Cotsapas C, Voight BF, Rossin E, Lage K, Neale BM, Wallace C, et al. Pervasive sharing of genetic effects in autoimmune disease. PLoS Genet. 2011 Aug;7(8):e1002254. PubMed PMID: 21852963. Pubmed Central PMCID: 3154137.
- Baranzini SE. The genetics of autoimmune diseases: a networked perspective. Curr Opin Immunol. 2009 Dec;21(6):596-605. PubMed PMID: 19896815.
- 32. Alter M, Leibowitz U, Speer J. Risk of multiple sclerosis related to age at immigration to Israel. Arch Neurol. 1966 Sep;15(3):234-7. PubMed PMID: 5912003. Epub 1966/09/01.
- Gale CR, Martyn CN. Migrant studies in multiple sclerosis. Prog Neurobiol. 1995 Nov-Dec;47(4-5):425-48. PubMed PMID: 8966212. Epub 1995/11/01.
- 34. Joensen P. Multiple sclerosis: variation of incidence of onset over time in the Faroe Islands. Mult Scler. 2011 Feb;17(2):241-4. PubMed PMID: 20978036. Epub 2010/10/28.
- Rosati G, Aiello I, Granieri E, Pirastru MI, Becciu S, Demontis G, et al. Incidence of multiple sclerosis in Macomer, Sardinia, 1912-1981: onset of the disease after 1950. Neurology. 1986 Jan;36(1):14-9. PubMed PMID: 3510404. Epub 1986/01/01.
- Kurtzke JF, Hyllested K. Validity of the epidemics of multiple sclerosis in the Faroe Islands. Neuroepidemiology. 1988;7(4):190-227. PubMed PMID: 3264056.
- Cohen JI. Epstein-Barr virus infection. N Engl J Med. 2000 Aug 17;343(7):481-92. Pub-Med PMID: 10944566.
- Lunemann JD. Epstein-Barr virus in multiple sclerosis: a continuing conundrum. Neurology. 2012 Jan 3;78(1):11-2. PubMed PMID: 22156986.
- Ascherio A, Munger KL. Environmental risk factors for multiple sclerosis. Part I: the role of infection. Ann Neurol. 2007 Apr;61(4):288-99. PubMed PMID: 17444504. Epub 2007/04/21.
- Handel AE, Williamson AJ, Disanto G, Handunnetthi L, Giovannoni G, Ramagopalan SV. An updated meta-analysis of risk of multiple sclerosis following infectious mononucleosis. PLoS One. 2010;5(9). PubMed PMID: 20824132. Pubmed Central PMCID: 2931696. Epub 2010/09/09.
- 41. Serafini B, Rosicarelli B, Franciotta D, Magliozzi R, Reynolds R, Cinque P, et al. Dysregulated Epstein-Barr virus infection in the multiple sclerosis brain. J Exp Med. 2007 Nov 26;204(12):2899-912. PubMed PMID: 17984305. Pubmed Central PMCID: 2118531. Epub 2007/11/07.
- 42. Hernan MA, Olek MJ, Ascherio A. Cigarette smoking and incidence of multiple sclerosis. Am J Epidemiol. 2001 Jul 1;154(1):69-74. PubMed PMID: 11427406. Epub 2001/06/28.
- 43. Mikaeloff Y, Caridade G, Tardieu M, Suissa S. Parental smoking at home and the risk of childhood-onset multiple sclerosis in children. Brain. 2007 Oct;130(Pt 10):2589-95. PubMed PMID: 17827175. Epub 2007/09/11.
- Hedstrom AK, Baarnhielm M, Olsson T, Alfredsson L. Exposure to environmental tobacco smoke is associated with increased risk for multiple sclerosis. Mult Scler. 2011 Jul;17(7):788-93. PubMed PMID: 21372120. Epub 2011/03/05.
- 45. Di Pauli F, Reindl M, Ehling R, Schautzer F, Gneiss C, Lutterotti A, et al. Smoking is a risk factor for early conversion to clinically definite multiple sclerosis. Mult Scler. 2008 Sep;14(8):1026-30. PubMed PMID: 18632775. Epub 2008/07/18.

- 46. Zivadinov R, Weinstock-Guttman B, Hashmi K, Abdelrahman N, Stosic M, Dwyer M, et al. Smoking is associated with increased lesion volumes and brain atrophy in multiple sclerosis. Neurology. 2009 Aug 18;73(7):504-10. PubMed PMID: 19687451. Pubmed Central PMCID: 2833095. Epub 2009/08/19.
- 47. Ascherio A, Munger KL, Simon KC. Vitamin D and multiple sclerosis. Lancet Neurol. 2010 Jun;9(6):599-612. PubMed PMID: 20494325. Epub 2010/05/25.
- Lucas RM, Ponsonby AL. Considering the potential benefits as well as adverse effects of sun exposure: can all the potential benefits be provided by oral vitamin D supplementation? Prog Biophys Mol Biol. 2006 Sep;92(1):140-9. PubMed PMID: 16616326.
- Becklund BR, Severson KS, Vang SV, DeLuca HF. UV radiation suppresses experimental autoimmune encephalomyelitis independent of vitamin D production. Proc Natl Acad Sci U S A. 2010 Apr 6;107(14):6418-23. PubMed PMID: 20308557. Pubmed Central PM-CID: 2851981.
- Munger KL, Levin LI, Hollis BW, Howard NS, Ascherio A. Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. JAMA. 2006 Dec 20;296(23):2832-8. PubMed PMID: 17179460. Epub 2006/12/21.
- Simpson S, Jr., Taylor B, Blizzard L, Ponsonby AL, Pittas F, Tremlett H, et al. Higher 25-hydroxyvitamin D is associated with lower relapse risk in multiple sclerosis. Ann Neurol. 2010 Aug;68(2):193-203. PubMed PMID: 20695012. Epub 2010/08/10.
- Burton JM, Kimball S, Vieth R, Bar-Or A, Dosch HM, Cheung R, et al. A phase I/II doseescalation trial of vitamin D3 and calcium in multiple sclerosis. Neurology. 2010 Jun 8;74(23):1852-9. PubMed PMID: 20427749. Pubmed Central PMCID: 2882221. Epub 2010/04/30.
- 53. Ramagopalan SV, Heger A, Berlanga AJ, Maugeri NJ, Lincoln MR, Burrell A, et al. A ChIP-seq defined genome-wide map of vitamin D receptor binding: associations with disease and evolution. Genome Res. 2010 Oct;20(10):1352-60. PubMed PMID: 20736230. Pubmed Central PMCID: 2945184.
- Ramagopalan SV, Maugeri NJ, Handunnetthi L, Lincoln MR, Orton SM, Dyment DA, et al. Expression of the multiple sclerosis-associated MHC class II Allele HLA-DRB1\*1501 is regulated by vitamin D. PLoS Genet. 2009 Feb;5(2):e1000369. PubMed PMID: 19197344. Pubmed Central PMCID: 2627899.
- Hedstrom AK, Sundqvist E, Baarnhielm M, Nordin N, Hillert J, Kockum I, et al. Smoking and two human leukocyte antigen genes interact to increase the risk for multiple sclerosis. Brain. 2011 Mar;134(Pt 3):653-64. PubMed PMID: 21303861.
- Runmarker B, Andersen O. Prognostic factors in a multiple sclerosis incidence cohort with twenty-five years of follow-up. Brain. 1993 Feb;116 (Pt 1):117-34. PubMed PMID: 8453453. Epub 1993/02/01.
- Miller DH, Chard DT, Ciccarelli O. Clinically isolated syndromes. Lancet Neurol. 2012 Feb;11(2):157-69. PubMed PMID: 22265211.
- Glad SB, Aarseth JH, Nyland H, Riise T, Myhr KM. Benign multiple sclerosis: a need for a consensus. Acta Neurol Scand Suppl. 2010 (190):44-50. PubMed PMID: 20586735.
- Correale J, Ysrraelit MC, Fiol MP. Benign multiple sclerosis: does it exist? Curr Neurol Neurosci Rep. 2012 Oct;12(5):601-9. PubMed PMID: 22777531.
- 60. Correale J, Peirano I, Romano L. Benign multiple sclerosis: a new definition of this entity is needed. Mult Scler. 2012 Feb;18(2):210-8. PubMed PMID: 21865415.

- Confavreux C, Vukusic S. Age at disability milestones in multiple sclerosis. Brain. 2006 Mar;129(Pt 3):595-605. PubMed PMID: 16415309. Epub 2006/01/18.
- Poser CM, Paty DW, Scheinberg L, McDonald WI, Davis FA, Ebers GC, et al. New diagnostic criteria for multiple sclerosis: guidelines for research protocols. Ann Neurol. 1983 Mar;13(3):227-31. PubMed PMID: 6847134. Epub 1983/03/01.
- Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, Filippi M, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. Ann Neurol. 2011 Feb;69(2):292-302. PubMed PMID: 21387374. Pubmed Central PMCID: 3084507. Epub 2011/03/10.
- Lublin FD, Reingold SC. Defining the clinical course of multiple sclerosis: results of an international survey. National Multiple Sclerosis Society (USA) Advisory Committee on Clinical Trials of New Agents in Multiple Sclerosis. Neurology. 1996 Apr;46(4):907-11. PubMed PMID: 8780061. Epub 1996/04/01.
- Stark GR, Kerr IM, Williams BR, Silverman RH, Schreiber RD. How cells respond to interferons. Annu Rev Biochem. 1998;67:227-64. PubMed PMID: 9759489. Epub 1998/10/06.
- 66. Prat A, Biernacki K, Antel JP. Th1 and Th2 lymphocyte migration across the human BBB is specifically regulated by interferon beta and copolymer-1. J Autoimmun. 2005 Mar;24(2):119-24. PubMed PMID: 15829404. Epub 2005/04/15.
- Kieseier BC. The mechanism of action of interferon-beta in relapsing multiple sclerosis. CNS Drugs. 2011 Jun 1;25(6):491-502. PubMed PMID: 21649449. Epub 2011/06/09.
- Mirandola SR, Hallal DE, Farias AS, Oliveira EC, Brandao CO, Ruocco HH, et al. Interferon-beta modifies the peripheral blood cell cytokine secretion in patients with multiple sclerosis. Int Immunopharmacol. 2009 Jul;9(7-8):824-30. PubMed PMID: 19289181. Epub 2009/03/18.
- Graber JJ, McGraw CA, Kimbrough D, Dhib-Jalbut S. Overlapping and distinct mechanisms of action of multiple sclerosis therapies. Clin Neurol Neurosurg. 2010 Sep;112(7):583-91. PubMed PMID: 20627553. Epub 2010/07/16.
- Pittock SJ. Interferon beta in multiple sclerosis: how much BENEFIT? Lancet. 2007 Aug 4;370(9585):363-4. PubMed PMID: 17678997. Epub 2007/08/07.
- Li DK, Zhao GJ, Paty DW. Randomized controlled trial of interferon-beta-1a in secondary progressive MS: MRI results. Neurology. 2001 Jun 12;56(11):1505-13. PubMed PMID: 11402107. Epub 2001/06/13.
- Cohen JA, Cutter GR, Fischer JS, Goodman AD, Heidenreich FR, Kooijmans MF, et al. Benefit of interferon beta-1a on MSFC progression in secondary progressive MS. Neurology. 2002 Sep 10;59(5):679-87. PubMed PMID: 12221157. Epub 2002/09/11.
- 73. Interferon beta-1b is effective in relapsing-remitting multiple sclerosis. I. Clinical results of a multicenter, randomized, double-blind, placebo-controlled trial. The IFNB Multiple Sclerosis Study Group. Neurology. 1993 Apr;43(4):655-61. PubMed PMID: 8469318.
- 74. Panitch H, Miller A, Paty D, Weinshenker B, North American Study Group on Interferon beta-1b in Secondary Progressive MS. Interferon beta-1b in secondary progressive MS: results from a 3-year controlled study. Neurology. 2004 Nov 23;63(10):1788-95. PubMed PMID: 15557491.
- Giovannoni G, Southam E, Waubant E. Systematic review of disease-modifying therapies to assess unmet needs in multiple sclerosis: tolerability and adherence. Mult Scler. 2012 Jul;18(7):932-46. PubMed PMID: 22249762. Epub 2012/01/18.

- Hegen H, Schleiser M, Gneiss C, Di Pauli F, Ehling R, Kuenz B, et al. Persistency of neutralizing antibodies depends on titer and interferon-beta preparation. Mult Scler. 2012 May;18(5):610-5. PubMed PMID: 22013146.
- 77. Killestein J, Polman CH. Determinants of interferon beta efficacy in patients with multiple sclerosis. Nat Rev Neurol. 2011 Apr;7(4):221-8. PubMed PMID: 21364522. Epub 2011/03/03.
- Carter NJ, Keating GM. Glatiramer acetate: a review of its use in relapsing-remitting multiple sclerosis and in delaying the onset of clinically definite multiple sclerosis. Drugs. 2010 Aug 20;70(12):1545-77. PubMed PMID: 20687620. Epub 2010/08/07.
- Blanchette F, Neuhaus O. Glatiramer acetate: evidence for a dual mechanism of action. J Neurol. 2008 Mar;255 Suppl 1:26-36. PubMed PMID: 18317674. Epub 2008/04/09.
- Hestvik AL, Skorstad G, Price DA, Vartdal F, Holmoy T. Multiple sclerosis: glatiramer acetate induces anti-inflammatory T cells in the cerebrospinal fluid. Mult Scler. 2008 Jul;14(6):749-58. PubMed PMID: 18611988. Epub 2008/07/10.
- Brenner T, Arnon R, Sela M, Abramsky O, Meiner Z, Riven-Kreitman R, et al. Humoral and cellular immune responses to Copolymer 1 in multiple sclerosis patients treated with Copaxone. J Neuroimmunol. 2001 Apr 2;115(1-2):152-60. PubMed PMID: 11282165. Epub 2001/04/03.
- 82. Teitelbaum D, Brenner T, Abramsky O, Aharoni R, Sela M, Arnon R. Antibodies to glatiramer acetate do not interfere with its biological functions and therapeutic efficacy. Mult Scler. 2003 Dec;9(6):592-9. PubMed PMID: 14664472. Epub 2003/12/11.
- Mikol DD, Barkhof F, Chang P, Coyle PK, Jeffery DR, Schwid SR, et al. Comparison of subcutaneous interferon beta-1a with glatiramer acetate in patients with relapsing multiple sclerosis (the REbif vs Glatiramer Acetate in Relapsing MS Disease [REGARD] study): a multicentre, randomised, parallel, open-label trial. Lancet Neurol. 2008 Oct;7(10):903-14. PubMed PMID: 18789766. Epub 2008/09/16.
- Cadavid D, Wolansky LJ, Skurnick J, Lincoln J, Cheriyan J, Szczepanowski K, et al. Efficacy of treatment of MS with IFNbeta-1b or glatiramer acetate by monthly brain MRI in the BECOME study. Neurology. 2009 Jun 9;72(23):1976-83. PubMed PMID: 19279320. Epub 2009/03/13.
- O'Connor P, Filippi M, Arnason B, Comi G, Cook S, Goodin D, et al. 250 microg or 500 microg interferon beta-1b versus 20 mg glatiramer acetate in relapsing-remitting multiple sclerosis: a prospective, randomised, multicentre study. Lancet Neurol. 2009 Oct;8(10):889-97. PubMed PMID: 19729344. Epub 2009/09/05.
- Bornstein MB, Miller A, Slagle S, Weitzman M, Drexler E, Keilson M, et al. A placebocontrolled, double-blind, randomized, two-center, pilot trial of Cop 1 in chronic progressive multiple sclerosis. Neurology. 1991 Apr;41(4):533-9. PubMed PMID: 2011253.
- Wolinsky JS, Narayana PA, O'Connor P, Coyle PK, Ford C, Johnson K, et al. Glatiramer acetate in primary progressive multiple sclerosis: results of a multinational, multicenter, double-blind, placebo-controlled trial. Ann Neurol. 2007 Jan;61(1):14-24. PubMed PMID: 17262850.
- Johnson KP, Brooks BR, Cohen JA, Ford CC, Goldstein J, Lisak RP, et al. Copolymer 1 reduces relapse rate and improves disability in relapsing-remitting multiple sclerosis: results of a phase III multicenter, double-blind placebo-controlled trial. The Copolymer 1 Multiple Sclerosis Study Group. Neurology. 1995 Jul;45(7):1268-76. PubMed PMID: 7617181. Epub 1995/07/01.

- Polman CH, O'Connor PW, Havrdova E, Hutchinson M, Kappos L, Miller DH, et al. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. N Engl J Med. 2006 Mar 2;354(9):899-910. PubMed PMID: 16510744. Epub 2006/03/03.
- Vermersch P, Kappos L, Gold R, Foley JF, Olsson T, Cadavid D, et al. Clinical outcomes of natalizumab-associated progressive multifocal leukoencephalopathy. Neurology. 2011 May 17;76(20):1697-704. PubMed PMID: 21576685. Epub 2011/05/18.
- 91. Chataway J, Miller DH. Natalizumab therapy for multiple sclerosis. Neurotherapeutics. 2013 Jan;10(1):19-28. PubMed PMID: 23307290. Pubmed Central PMCID: 3557363. Epub 2013/01/12.
- Scott LJ, Figgitt DP. Mitoxantrone: a review of its use in multiple sclerosis. CNS Drugs. 2004;18(6):379-96. PubMed PMID: 15089110. Epub 2004/04/20.
- 93. Edan G, Miller D, Clanet M, Confavreux C, Lyon-Caen O, Lubetzki C, et al. Therapeutic effect of mitoxantrone combined with methylprednisolone in multiple sclerosis: a randomised multicentre study of active disease using MRI and clinical criteria. J Neurol Neurosurg Psychiatry. 1997 Feb;62(2):112-8. PubMed PMID: 9048709. Pubmed Central PMCID: 486720. Epub 1997/02/01.
- 94. Hartung HP, Gonsette R, Konig N, Kwiecinski H, Guseo A, Morrissey SP, et al. Mitoxantrone in progressive multiple sclerosis: a placebo-controlled, double-blind, randomised, multicentre trial. Lancet. 2002 Dec 21-28;360(9350):2018-25. PubMed PMID: 12504397. Epub 2002/12/31.
- 95. Marriott JJ, Miyasaki JM, Gronseth G, O'Connor PW, Therapeutics, Technology Assessment Subcommittee of the American Academy of N. Evidence Report: The efficacy and safety of mitoxantrone (Novantrone) in the treatment of multiple sclerosis: Report of the Therapeutics and Technology Assessment Subcommittee of the American Academy of Neurology. Neurology. 2010 May 4;74(18):1463-70. PubMed PMID: 20439849. Pubmed Central PMCID: 2871006. Epub 2010/05/05.
- 96. MS-association. Ts. Criteria for MS-treatment hwms, 2012). aa.
- Clynes RA, Towers TL, Presta LG, Ravetch JV. Inhibitory Fc receptors modulate in vivo cytotoxicity against tumor targets. Nat Med. 2000 Apr;6(4):443-6. PubMed PMID: 10742152. Epub 2000/03/31.
- Browning JL. B cells move to centre stage: novel opportunities for autoimmune disease treatment. Nat Rev Drug Discov. 2006 Jul;5(7):564-76. PubMed PMID: 16816838. Epub 2006/07/04.
- Dalakas MC. B cells as therapeutic targets in autoimmune neurological disorders. Nat Clin Pract Neurol. 2008 Oct;4(10):557-67. PubMed PMID: 18813230. Epub 2008/09/25.
- Shlomchik MJ, Craft JE, Mamula MJ. From T to B and back again: positive feedback in systemic autoimmune disease. Nat Rev Immunol. 2001 Nov;1(2):147-53. PubMed PMID: 11905822. Epub 2002/03/22.
- Petereit HF, Rubbert-Roth A. Rituximab levels in cerebrospinal fluid of patients with neurological autoimmune disorders. Mult Scler. 2009 Feb;15(2):189-92. PubMed PMID: 18971221. Epub 2008/10/31.
- Kitsos DK, Tsiodras S, Stamboulis E, Voumvourakis KI. Rituximab and multiple sclerosis. Clin Neuropharmacol. 2012 Mar-Apr;35(2):90-6. PubMed PMID: 22421587. Epub 2012/03/17.

- Naismith RT, Piccio L, Lyons JA, Lauber J, Tutlam NT, Parks BJ, et al. Rituximab add-on therapy for breakthrough relapsing multiple sclerosis: a 52-week phase II trial. Neurology. 2010 Jun 8;74(23):1860-7. PubMed PMID: 20530322. Pubmed Central PMCID: 2882224.
- 104. Hawker K, O'Connor P, Freedman MS, Calabresi PA, Antel J, Simon J, et al. Rituximab in patients with primary progressive multiple sclerosis: results of a randomized doubleblind placebo-controlled multicenter trial. Ann Neurol. 2009 Oct;66(4):460-71. PubMed PMID: 19847908. Epub 2009/10/23.
- 105. Lassmann H, Bruck W, Lucchinetti CF. The immunopathology of multiple sclerosis: an overview. Brain Pathol. 2007 Apr;17(2):210-8. PubMed PMID: 17388952. Epub 2007/03/29.
- Barnett MH, Prineas JW. Relapsing and remitting multiple sclerosis: pathology of the newly forming lesion. Ann Neurol. 2004 Apr;55(4):458-68. PubMed PMID: 15048884.
- 107. Prineas JW, Wright RG. Macrophages, lymphocytes, and plasma cells in the perivascular compartment in chronic multiple sclerosis. Lab Invest. 1978 Apr;38(4):409-21. PubMed PMID: 205724.
- Cannella B, Raine CS. The adhesion molecule and cytokine profile of multiple sclerosis lesions. Ann Neurol. 1995 Apr;37(4):424-35. PubMed PMID: 7536402. Epub 1995/04/01.
- 109. Huang D, Han Y, Rani MR, Glabinski A, Trebst C, Sorensen T, et al. Chemokines and chemokine receptors in inflammation of the nervous system: manifold roles and exquisite regulation. Immunol Rev. 2000 Oct;177:52-67. PubMed PMID: 11138785. Epub 2001/01/04.
- 110. Hochmeister S, Grundtner R, Bauer J, Engelhardt B, Lyck R, Gordon G, et al. Dysferlin is a new marker for leaky brain blood vessels in multiple sclerosis. J Neuropathol Exp Neurol. 2006 Sep;65(9):855-65. PubMed PMID: 16957579. Epub 2006/09/08.
- 111. Kwon EE, Prineas JW. Blood-brain barrier abnormalities in longstanding multiple sclerosis lesions. An immunohistochemical study. J Neuropathol Exp Neurol. 1994 Nov;53(6):625-36. PubMed PMID: 7964903. Epub 1994/11/01.
- Trapp BD, Peterson J, Ransohoff RM, Rudick R, Mork S, Bo L. Axonal transection in the lesions of multiple sclerosis. N Engl J Med. 1998 Jan 29;338(5):278-85. PubMed PMID: 9445407. Epub 1998/01/29.
- 113. Ferguson B, Matyszak MK, Esiri MM, Perry VH. Axonal damage in acute multiple sclerosis lesions. Brain. 1997 Mar;120 ( Pt 3):393-9. PubMed PMID: 9126051. Epub 1997/03/01.
- 114. Allen IV, McQuaid S, Mirakhur M, Nevin G. Pathological abnormalities in the normalappearing white matter in multiple sclerosis. Neurol Sci. 2001 Apr;22(2):141-4. PubMed PMID: 11603615. Epub 2001/10/18.
- Evangelou N, DeLuca GC, Owens T, Esiri MM. Pathological study of spinal cord atrophy in multiple sclerosis suggests limited role of local lesions. Brain. 2005 Jan;128(Pt 1):29-34. PubMed PMID: 15548559.
- Love S. Demyelinating diseases. J Clin Pathol. 2006 Nov;59(11):1151-9. PubMed PMID: 17071802. Pubmed Central PMCID: 1860500.
- Sofroniew MV, Vinters HV. Astrocytes: biology and pathology. Acta Neuropathol. 2010 Jan;119(1):7-35. PubMed PMID: 20012068. Pubmed Central PMCID: 2799634.

- 118. Voskuhl RR, Peterson RS, Song B, Ao Y, Morales LB, Tiwari-Woodruff S, et al. Reactive astrocytes form scar-like perivascular barriers to leukocytes during adaptive immune inflammation of the CNS. J Neurosci. 2009 Sep 16;29(37):11511-22. PubMed PMID: 19759299. Pubmed Central PMCID: 2768309. Epub 2009/09/18.
- 119. Silver J, Miller JH. Regeneration beyond the glial scar. Nat Rev Neurosci. 2004 Feb;5(2):146-56. PubMed PMID: 14735117.
- Lucchinetti C, Bruck W, Parisi J, Scheithauer B, Rodriguez M, Lassmann H. Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. Ann Neurol. 2000 Jun;47(6):707-17. PubMed PMID: 10852536. Epub 2000/06/14.
- 121. Leussink VI, Lehmann HC, Meyer zu Horste G, Hartung HP, Stuve O, Kieseier BC. Rituximab induces clinical stabilization in a patient with fulminant multiple sclerosis not responding to natalizumab. Evidence for disease heterogeneity. J Neurol. 2008 Sep;255(9):1436-8. PubMed PMID: 18685916.
- 122. Gold R, Linington C. Devic's disease: bridging the gap between laboratory and clinic. Brain. 2002 Jul;125(Pt 7):1425-7. PubMed PMID: 12076994.
- Peterson JW, Bo L, Mork S, Chang A, Trapp BD. Transected neurites, apoptotic neurons, and reduced inflammation in cortical multiple sclerosis lesions. Ann Neurol. 2001 Sep;50(3):389-400. PubMed PMID: 11558796. Epub 2001/09/18.
- Calabrese M, Gallo P. Magnetic resonance evidence of cortical onset of multiple sclerosis. Mult Scler. 2009 Aug;15(8):933-41. PubMed PMID: 19667021. Epub 2009/08/12.
- Lucchinetti CF, Popescu BF, Bunyan RF, Moll NM, Roemer SF, Lassmann H, et al. Inflammatory cortical demyelination in early multiple sclerosis. N Engl J Med. 2011 Dec 8;365(23):2188-97. PubMed PMID: 22150037. Pubmed Central PMCID: 3282172. Epub 2011/12/14.
- 126. Howell OW, Reeves CA, Nicholas R, Carassiti D, Radotra B, Gentleman SM, et al. Meningeal inflammation is widespread and linked to cortical pathology in multiple sclerosis. Brain. 2011 Sep;134(Pt 9):2755-71. PubMed PMID: 21840891. Epub 2011/08/16.
- Popescu BF, Lucchinetti CF. Meningeal and cortical grey matter pathology in multiple sclerosis. BMC Neurol. 2012;12:11. PubMed PMID: 22397318. Pubmed Central PM-CID: 3315403. Epub 2012/03/09.
- Wegner C, Esiri MM, Chance SA, Palace J, Matthews PM. Neocortical neuronal, synaptic, and glial loss in multiple sclerosis. Neurology. 2006 Sep 26;67(6):960-7. PubMed PMID: 17000961. Epub 2006/09/27.
- 129. Serafini B, Rosicarelli B, Magliozzi R, Stigliano E, Aloisi F. Detection of ectopic Bcell follicles with germinal centers in the meninges of patients with secondary progressive multiple sclerosis. Brain Pathol. 2004 Apr;14(2):164-74. PubMed PMID: 15193029. Epub 2004/06/15.
- Magliozzi R, Howell O, Vora A, Serafini B, Nicholas R, Puopolo M, et al. Meningeal Bcell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical pathology. Brain. 2007 Apr;130(Pt 4):1089-104. PubMed PMID: 17438020. Epub 2007/04/18.
- 131. Lassmann H, van Horssen J, Mahad D. Progressive multiple sclerosis: pathology and pathogenesis. Nat Rev Neurol. 2012 Nov 5;8(11):647-56. PubMed PMID: 23007702.

- 132. Bjartmar C, Wujek JR, Trapp BD. Axonal loss in the pathology of MS: consequences for understanding the progressive phase of the disease. J Neurol Sci. 2003 Feb 15;206(2):165-71. PubMed PMID: 12559505.
- 133. Frohman EM, Filippi M, Stuve O, Waxman SG, Corboy J, Phillips JT, et al. Characterizing the mechanisms of progression in multiple sclerosis: evidence and new hypotheses for future directions. Arch Neurol. 2005 Sep;62(9):1345-56. PubMed PMID: 16157741.
- 134. De Groot CJ, Bergers E, Kamphorst W, Ravid R, Polman CH, Barkhof F, et al. Postmortem MRI-guided sampling of multiple sclerosis brain lesions: increased yield of active demyelinating and (p)reactive lesions. Brain. 2001 Aug;124(Pt 8):1635-45. PubMed PMID: 11459754. Epub 2001/07/19.
- Czeh M, Gressens P, Kaindl AM. The yin and yang of microglia. Dev Neurosci. 2011;33(3-4):199-209. PubMed PMID: 21757877. Epub 2011/07/16.
- 136. Kornek B, Storch MK, Bauer J, Djamshidian A, Weissert R, Wallstroem E, et al. Distribution of a calcium channel subunit in dystrophic axons in multiple sclerosis and experimental autoimmune encephalomyelitis. Brain. 2001 Jun;124(Pt 6):1114-24. PubMed PMID: 11353727.
- 137. Ouardouz M, Coderre E, Basak A, Chen A, Zamponi GW, Hameed S, et al. Glutamate receptors on myelinated spinal cord axons: I. GluR6 kainate receptors. Ann Neurol. 2009 Feb;65(2):151-9. PubMed PMID: 19224535. Pubmed Central PMCID: 2902553.
- 138. Craner MJ, Newcombe J, Black JA, Hartle C, Cuzner ML, Waxman SG. Molecular changes in neurons in multiple sclerosis: altered axonal expression of Nav1.2 and Nav1.6 sodium channels and Na+/Ca2+ exchanger. Proc Natl Acad Sci U S A. 2004 May 25;101(21):8168-73. PubMed PMID: 15148385. Pubmed Central PMCID: 419575.
- Black JA, Liu S, Hains BC, Saab CY, Waxman SG. Long-term protection of central axons with phenytoin in monophasic and chronic-relapsing EAE. Brain. 2006 Dec;129(Pt 12):3196-208. PubMed PMID: 16931536.
- Trapp BD, Stys PK. Virtual hypoxia and chronic necrosis of demyelinated axons in multiple sclerosis. Lancet Neurol. 2009 Mar;8(3):280-91. PubMed PMID: 19233038.
- 141. Witte ME, Geurts JJ, de Vries HE, van der Valk P, van Horssen J. Mitochondrial dysfunction: a potential link between neuroinflammation and neurodegeneration? Mitochondrion. 2010 Aug;10(5):411-8. PubMed PMID: 20573557.
- Zambonin JL, Zhao C, Ohno N, Campbell GR, Engeham S, Ziabreva I, et al. Increased mitochondrial content in remyelinated axons: implications for multiple sclerosis. Brain. 2011 Jul;134(Pt 7):1901-13. PubMed PMID: 21705418. Pubmed Central PMCID: 3122369.
- 143. Witte ME, Bo L, Rodenburg RJ, Belien JA, Musters R, Hazes T, et al. Enhanced number and activity of mitochondria in multiple sclerosis lesions. J Pathol. 2009 Oct;219(2):193-204. PubMed PMID: 19591199.
- 144. Mahad DJ, Ziabreva I, Campbell G, Lax N, White K, Hanson PS, et al. Mitochondrial changes within axons in multiple sclerosis. Brain. 2009 May;132(Pt 5):1161-74. PubMed PMID: 19293237. Pubmed Central PMCID: 3605917.
- Mahad D, Lassmann H, Turnbull D. Review: Mitochondria and disease progression in multiple sclerosis. Neuropathol Appl Neurobiol. 2008 Dec;34(6):577-89. PubMed PMID: 19076696. Pubmed Central PMCID: 2981078.

- 146. Bolanos JP, Almeida A, Stewart V, Peuchen S, Land JM, Clark JB, et al. Nitric oxidemediated mitochondrial damage in the brain: mechanisms and implications for neurodegenerative diseases. J Neurochem. 1997 Jun;68(6):2227-40. PubMed PMID: 9166714.
- Smith KJ, Kapoor R, Felts PA. Demyelination: the role of reactive oxygen and nitrogen species. Brain Pathol. 1999 Jan;9(1):69-92. PubMed PMID: 9989453.
- Chang A, Tourtellotte WW, Rudick R, Trapp BD. Premyelinating oligodendrocytes in chronic lesions of multiple sclerosis. N Engl J Med. 2002 Jan 17;346(3):165-73. PubMed PMID: 11796850.
- 149. Veto S, Acs P, Bauer J, Lassmann H, Berente Z, Setalo G, Jr., et al. Inhibiting poly(ADPribose) polymerase: a potential therapy against oligodendrocyte death. Brain. 2010 Mar;133(Pt 3):822-34. PubMed PMID: 20157013. Pubmed Central PMCID: 2964508.
- Lopes KO, Sparks DL, Streit WJ. Microglial dystrophy in the aged and Alzheimer's disease brain is associated with ferritin immunoreactivity. Glia. 2008 Aug 1;56(10):1048-60. PubMed PMID: 18442088.
- 151. Zhang X, Haaf M, Todorich B, Grosstephan E, Schieremberg H, Surguladze N, et al. Cytokine toxicity to oligodendrocyte precursors is mediated by iron. Glia. 2005 Nov 15;52(3):199-208. PubMed PMID: 15968631.
- 152. Kurtzke JF. Rating neurologic impairment in multiple sclerosis: an expanded disability status scale (EDSS). Neurology. 1983 Nov;33(11):1444-52. PubMed PMID: 6685237. Epub 1983/11/01.
- 153. Amato MP, Fratiglioni L, Groppi C, Siracusa G, Amaducci L. Interrater reliability in assessing functional systems and disability on the Kurtzke scale in multiple sclerosis. Arch Neurol. 1988 Jul;45(7):746-8. PubMed PMID: 3390030. Epub 1988/07/01.
- 154. Goodkin DE, Cookfair D, Wende K, Bourdette D, Pullicino P, Scherokman B, et al. Interand intrarater scoring agreement using grades 1.0 to 3.5 of the Kurtzke Expanded Disability Status Scale (EDSS). Multiple Sclerosis Collaborative Research Group. Neurology. 1992 Apr;42(4):859-63. PubMed PMID: 1565242. Epub 1992/04/01.
- 155. Noseworthy JH, Vandervoort MK, Wong CJ, Ebers GC. Interrater variability with the Expanded Disability Status Scale (EDSS) and Functional Systems (FS) in a multiple sclerosis clinical trial. The Canadian Cooperation MS Study Group. Neurology. 1990 Jun;40(6):971-5. PubMed PMID: 2189084. Epub 1990/06/01.
- 156. Roxburgh RH, Seaman SR, Masterman T, Hensiek AE, Sawcer SJ, Vukusic S, et al. Multiple Sclerosis Severity Score: using disability and disease duration to rate disease severity. Neurology. 2005 Apr 12;64(7):1144-51. PubMed PMID: 15824338. Epub 2005/04/13.
- 157. Poser S, Ritter G, Bauer HJ, Grosse-Wilde H, Kuwert EK, Raun NE. HLA-antigens and the prognosis of multiple sclerosis. J Neurol. 1981;225(3):219-21. PubMed PMID: 6167687. Epub 1981/01/01.
- Trojano M, Liguori M, Bosco Zimatore G, Bugarini R, Avolio C, Paolicelli D, et al. Agerelated disability in multiple sclerosis. Ann Neurol. 2002 Apr;51(4):475-80. PubMed PMID: 11921053. Epub 2002/03/29.
- 159. Fischer JS, Rudick RA, Cutter GR, Reingold SC. The Multiple Sclerosis Functional Composite Measure (MSFC): an integrated approach to MS clinical outcome assessment. National MS Society Clinical Outcomes Assessment Task Force. Mult Scler. 1999 Aug;5(4):244-50. PubMed PMID: 10467383. Epub 1999/09/01.

- Cohen JA, Fischer JS, Bolibrush DM, Jak AJ, Kniker JE, Mertz LA, et al. Intrarater and interrater reliability of the MS functional composite outcome measure. Neurology. 2000 Feb 22;54(4):802-6. PubMed PMID: 10690966. Epub 2000/02/26.
- Bosma LV, Kragt JJ, Brieva L, Khaleeli Z, Montalban X, Polman CH, et al. The search for responsive clinical endpoints in primary progressive multiple sclerosis. Mult Scler. 2009 Jun;15(6):715-20. PubMed PMID: 19383646. Epub 2009/04/23.
- 162. Bosma L, Kragt J, Polman C, Uitdehaag B. Walking speed, rather than Expanded Disability Status Scale, relates to long-term patient-reported impact in progressive MS. Mult Scler. 2012 Aug 20. PubMed PMID: 22907939. Epub 2012/08/22.
- Kragt JJ, Thompson AJ, Montalban X, Tintore M, Rio J, Polman CH, et al. Responsiveness and predictive value of EDSS and MSFC in primary progressive MS. Neurology. 2008 Mar 25;70(13 Pt 2):1084-91. PubMed PMID: 18184917. Epub 2008/01/11.
- Bosma LV, Kragt JJ, Knol DL, Polman CH, Uitdehaag BM. Clinical scales in progressive MS: predicting long-term disability. Mult Scler. 2012 Mar;18(3):345-50. PubMed PMID: 21868487. Epub 2011/08/27.
- Polman CH, Rudick RA. The multiple sclerosis functional composite: a clinically meaningful measure of disability. Neurology. 2010 Apr 27;74 Suppl 3:S8-15. PubMed PMID: 20421572. Epub 2010/05/07.
- 166. van Walderveen MA, Kamphorst W, Scheltens P, van Waesberghe JH, Ravid R, Valk J, et al. Histopathologic correlate of hypointense lesions on T1-weighted spin-echo MRI in multiple sclerosis. Neurology. 1998 May;50(5):1282-8. PubMed PMID: 9595975. Epub 1998/05/22.
- 167. Rovaris M, Comi G, Rocca MA, Cercignani M, Colombo B, Santuccio G, et al. Relevance of hypointense lesions on fast fluid-attenuated inversion recovery MR images as a marker of disease severity in cases of multiple sclerosis. AJNR Am J Neuroradiol. 1999 May;20(5):813-20. PubMed PMID: 10369351. Epub 1999/06/16.
- Bruck W, Bitsch A, Kolenda H, Bruck Y, Stiefel M, Lassmann H. Inflammatory central nervous system demyelination: correlation of magnetic resonance imaging findings with lesion pathology. Ann Neurol. 1997 Nov;42(5):783-93. PubMed PMID: 9392578. Epub 1997/12/10.
- Meier DS, Weiner HL, Guttmann CR. MR imaging intensity modeling of damage and repair in multiple sclerosis: relationship of short-term lesion recovery to progression and disability. AJNR Am J Neuroradiol. 2007 Nov-Dec;28(10):1956-63. PubMed PMID: 17998417. Epub 2007/11/14.
- 170. Bitar R, Leung G, Perng R, Tadros S, Moody AR, Sarrazin J, et al. MR pulse sequences: what every radiologist wants to know but is afraid to ask. Radiographics. 2006 Mar-Apr;26(2):513-37. PubMed PMID: 16549614. Epub 2006/03/22.
- 171. Minagar A, Gonzalez-Toledo E, Pinkston J, Jaffe SL. Neuroimaging in multiple sclerosis. Int Rev Neurobiol. 2005;67:165-201. PubMed PMID: 16291023. Epub 2005/11/18.
- 172. Filippi M, Yousry T, Baratti C, Horsfield MA, Mammi S, Becker C, et al. Quantitative assessment of MRI lesion load in multiple sclerosis. A comparison of conventional spinecho with fast fluid-attenuated inversion recovery. Brain. 1996 Aug;119 (Pt 4):1349-55. PubMed PMID: 8813296. Epub 1996/08/01.
- Zivadinov R, Cox JL. Neuroimaging in multiple sclerosis. Int Rev Neurobiol. 2007;79:449-74. PubMed PMID: 17531854. Epub 2007/05/29.

- 174. Bilello M, Suri N, Krejza J, Woo JH, Bagley LJ, Mamourian AC, et al. An approach to comparing accuracies of two FLAIR MR sequences in the detection of multiple sclerosis lesions in the brain in the absence of gold standard. Acad Radiol. 2010 Jun;17(6):686-95. PubMed PMID: 20457413. Epub 2010/05/12.
- 175. Brex PA, O'Riordan JI, Miszkiel KA, Moseley IF, Thompson AJ, Plant GT, et al. Multisequence MRI in clinically isolated syndromes and the early development of MS. Neurology. 1999 Oct 12;53(6):1184-90. PubMed PMID: 10522870. Epub 1999/10/16.
- 176. Brex PA, Ciccarelli O, O'Riordan JI, Sailer M, Thompson AJ, Miller DH. A longitudinal study of abnormalities on MRI and disability from multiple sclerosis. N Engl J Med. 2002 Jan 17;346(3):158-64. PubMed PMID: 11796849. Epub 2002/01/18.
- Miller DH, Barkhof F, Nauta JJ. Gadolinium enhancement increases the sensitivity of MRI in detecting disease activity in multiple sclerosis. Brain. 1993 Oct;116 (Pt 5):1077-94. PubMed PMID: 8221048.
- 178. Nauta JJ, Thompson AJ, Barkhof F, Miller DH. Magnetic resonance imaging in monitoring the treatment of multiple sclerosis patients: statistical power of parallel-groups and crossover designs. J Neurol Sci. 1994 Mar;122(1):6-14. PubMed PMID: 8195804.
- 179. Simon JH. Brain atrophy in multiple sclerosis: what we know and would like to know. Mult Scler. 2006 Dec;12(6):679-87. PubMed PMID: 17262994. Epub 2007/02/01.
- Miller DH, Barkhof F, Frank JA, Parker GJ, Thompson AJ. Measurement of atrophy in multiple sclerosis: pathological basis, methodological aspects and clinical relevance. Brain. 2002 Aug;125(Pt 8):1676-95. PubMed PMID: 12135961. Epub 2002/07/24.
- Enzinger C, Fazekas F, Matthews PM, Ropele S, Schmidt H, Smith S, et al. Risk factors for progression of brain atrophy in aging: six-year follow-up of normal subjects. Neurology. 2005 May 24;64(10):1704-11. PubMed PMID: 15911795.
- Scahill RI, Frost C, Jenkins R, Whitwell JL, Rossor MN, Fox NC. A longitudinal study of brain volume changes in normal aging using serial registered magnetic resonance imaging. Arch Neurol. 2003 Jul;60(7):989-94. PubMed PMID: 12873856.
- 183. De Stefano N, Giorgio A, Battaglini M, Rovaris M, Sormani MP, Barkhof F, et al. Assessing brain atrophy rates in a large population of untreated multiple sclerosis subtypes. Neurology. 2010 Jun 8;74(23):1868-76. PubMed PMID: 20530323. Epub 2010/06/10.
- Fisniku LK, Chard DT, Jackson JS, Anderson VM, Altmann DR, Miszkiel KA, et al. Gray matter atrophy is related to long-term disability in multiple sclerosis. Ann Neurol. 2008 Sep;64(3):247-54. PubMed PMID: 18570297. Epub 2008/06/24.
- Lycklama G, Thompson A, Filippi M, Miller D, Polman C, Fazekas F, et al. Spinalcord MRI in multiple sclerosis. Lancet Neurol. 2003 Sep;2(9):555-62. PubMed PMID: 12941578. Epub 2003/08/28.
- Barkhof F, Calabresi PA, Miller DH, Reingold SC. Imaging outcomes for neuroprotection and repair in multiple sclerosis trials. Nat Rev Neurol. 2009 May;5(5):256-66. PubMed PMID: 19488083. Epub 2009/06/03.
- 187. Rieckmann P, Altenhofen B, Riegel A, Baudewig J, Felgenhauer K. Soluble adhesion molecules (sVCAM-1 and sICAM-1) in cerebrospinal fluid and serum correlate with MRI activity in multiple sclerosis. Ann Neurol. 1997 Mar;41(3):326-33. PubMed PMID: 9066353.
- Felgenhauer K. Evaluation of molecular size by gel electrophoretic techniques. Hoppe Seylers Z Physiol Chem. 1974 Oct;355(10):1281-90. PubMed PMID: 4461630.

- 189. Wen SR, Liu GJ, Feng RN, Gong FC, Zhong H, Duan SR, et al. Increased levels of IL-23 and osteopontin in serum and cerebrospinal fluid of multiple sclerosis patients. J Neuroimmunol. 2012 Mar;244(1-2):94-6. PubMed PMID: 22329905.
- Vogt MH, Floris S, Killestein J, Knol DL, Smits M, Barkhof F, et al. Osteopontin levels and increased disease activity in relapsing-remitting multiple sclerosis patients. J Neuroimmunol. 2004 Oct;155(1-2):155-60. PubMed PMID: 15342207.
- 191. Malmestrom C, Gillett A, Jernas M, Khademi M, Axelsson M, Kockum I, et al. Serum levels of LIGHT in MS. Mult Scler. 2012 Oct 4. PubMed PMID: 23037546.
- 192. Avolio C, Ruggieri M, Giuliani F, Liuzzi GM, Leante R, Riccio P, et al. Serum MMP-2 and MMP-9 are elevated in different multiple sclerosis subtypes. J Neuroimmunol. 2003 Mar;136(1-2):46-53. PubMed PMID: 12620642.
- Eikelenboom MJ, Uitdehaag BM, Petzold A. Blood and CSF Biomarker Dynamics in Multiple Sclerosis: Implications for Data Interpretation. Mult Scler Int. 2011;2011:823176. PubMed PMID: 22096644. Pubmed Central PMCID: 3195856.
- 194. Lennon VA, Kryzer TJ, Pittock SJ, Verkman AS, Hinson SR. IgG marker of optic-spinal multiple sclerosis binds to the aquaporin-4 water channel. J Exp Med. 2005 Aug 15;202(4):473-7. PubMed PMID: 16087714. Pubmed Central PMCID: 2212860.
- Racke MK. Multiple sclerosis: The potassium channel KIR4.1-a potential autoantigen in MS. Nat Rev Neurol. 2012 Sep 18. PubMed PMID: 22986435.
- Lee MK, Cleveland DW. Neuronal intermediate filaments. Annu Rev Neurosci. 1996;19:187-217. PubMed PMID: 8833441. Epub 1996/01/01.
- 197. Petzold A. Neurofilament phosphoforms: surrogate markers for axonal injury, degeneration and loss. J Neurol Sci. 2005 Jun 15;233(1-2):183-98. PubMed PMID: 15896809. Epub 2005/05/18.
- 198. Fuchs E, Cleveland DW. A structural scaffolding of intermediate filaments in health and disease. Science. 1998 Jan 23;279(5350):514-9. PubMed PMID: 9438837. Epub 1998/02/07.
- Teunissen CE, Khalil M. Neurofilaments as biomarkers in multiple sclerosis. Mult Scler. 2012 May;18(5):552-6. PubMed PMID: 22492131. Epub 2012/04/12.
- 200. Kim S, Chang R, Teunissen C, Gebremichael Y, Petzold A. Neurofilament stoichiometry simulations during neurodegeneration suggest a remarkable self-sufficient and stable in vivo protein structure. J Neurol Sci. 2011 Aug 15;307(1-2):132-8. PubMed PMID: 21601889. Epub 2011/05/24.
- 201. Studahl M, Rosengren L, Gunther G, Hagberg L. Difference in pathogenesis between herpes simplex virus type 1 encephalitis and tick-borne encephalitis demonstrated by means of cerebrospinal fluid markers of glial and neuronal destruction. J Neurol. 2000 Aug;247(8):636-42. PubMed PMID: 11041333. Epub 2000/10/21.
- Nylen K, Karlsson JE, Blomstrand C, Tarkowski A, Trysberg E, Rosengren LE. Cerebrospinal fluid neurofilament and glial fibrillary acidic protein in patients with cerebral vasculitis. J Neurosci Res. 2002 Mar 15;67(6):844-51. PubMed PMID: 11891800. Epub 2002/03/14.
- Constantinescu R, Rosengren L, Johnels B, Zetterberg H, Holmberg B. Consecutive analyses of cerebrospinal fluid axonal and glial markers in Parkinson's disease and atypical Parkinsonian disorders. Parkinsonism Relat Disord. 2010 Feb;16(2):142-5. PubMed PMID: 19647470. Epub 2009/08/04.

- Tortelli R, Ruggieri M, Cortese R, D'Errico E, Capozzo R, Leo A, et al. Elevated cerebrospinal fluid neurofilament light levels in patients with amyotrophic lateral sclerosis: a possible marker of disease severity and progression. Eur J Neurol. 2012 Dec;19(12):1561-7. PubMed PMID: 22680408. Epub 2012/06/12.
- Rosengren LE, Karlsson JE, Karlsson JO, Persson LI, Wikkelso C. Patients with amyotrophic lateral sclerosis and other neurodegenerative diseases have increased levels of neurofilament protein in CSF. J Neurochem. 1996 Nov;67(5):2013-8. PubMed PMID: 8863508. Epub 1996/11/01.
- 206. Teunissen CE, Iacobaeus E, Khademi M, Brundin L, Norgren N, Koel-Simmelink MJ, et al. Combination of CSF N-acetylaspartate and neurofilaments in multiple sclerosis. Neurology. 2009 Apr 14;72(15):1322-9. PubMed PMID: 19365053. Epub 2009/04/15.
- 207. Romme Christensen J, Bornsen L, Khademi M, Olsson T, Jensen PE, Sorensen PS, et al. CSF inflammation and axonal damage are increased and correlate in progressive multiple sclerosis. Mult Scler. 2012 Nov 24. PubMed PMID: 23178691.
- Malmestrom C, Haghighi S, Rosengren L, Andersen O, Lycke J. Neurofilament light protein and glial fibrillary acidic protein as biological markers in MS. Neurology. 2003 Dec 23;61(12):1720-5. PubMed PMID: 14694036. Epub 2003/12/25.
- Norgren N, Sundstrom P, Svenningsson A, Rosengren L, Stigbrand T, Gunnarsson M. Neurofilament and glial fibrillary acidic protein in multiple sclerosis. Neurology. 2004 Nov 9;63(9):1586-90. PubMed PMID: 15534240. Epub 2004/11/10.
- Khalil M, Enzinger C, Langkammer C, Ropele S, Mader A, Trentini A, et al. CSF neurofilament and N-acetylaspartate related brain changes in clinically isolated syndrome. Mult Scler. 2012 Aug 23. PubMed PMID: 22917689.
- 211. Petzold A, Eikelenboom MI, Keir G, Polman CH, Uitdehaag BM, Thompson EJ, et al. The new global multiple sclerosis severity score (MSSS) correlates with axonal but not glial biomarkers. Mult Scler. 2006 Jun;12(3):325-8. PubMed PMID: 16764346.
- Petzold A, Mondria T, Kuhle J, Rocca MA, Cornelissen J, te Boekhorst P, et al. Evidence for acute neurotoxicity after chemotherapy. Ann Neurol. 2010 Dec;68(6):806-15. Pub-Med PMID: 21194151. Epub 2011/01/05.
- Petzold A, Keir G, Green AJ, Giovannoni G, Thompson EJ. A specific ELISA for measuring neurofilament heavy chain phosphoforms. J Immunol Methods. 2003 Jul;278(1-2):179-90. PubMed PMID: 12957406. Epub 2003/09/06.
- Karlsson JE, Rosengren LE, Haglid KG. Polyclonal antisera to the individual neurofilament triplet proteins: a characterization using ELISA and immunoblotting. J Neurochem. 1989 Sep;53(3):759-65. PubMed PMID: 2760619. Epub 1989/09/01.
- Cullen DK, Simon CM, LaPlaca MC. Strain rate-dependent induction of reactive astrogliosis and cell death in three-dimensional neuronal-astrocytic co-cultures. Brain Res. 2007 Jul 16;1158:103-15. PubMed PMID: 17555726. Pubmed Central PMCID: 3179863. Epub 2007/06/09.
- 216. Kepes JJ, Perentes E. Glial fibrillary acidic protein in chondrocytes of elastic cartilage in the human epiglottis: an immunohistochemical study with polyvalent and monoclonal antibodies. Anat Rec. 1988 Mar;220(3):296-9. PubMed PMID: 3364756. Epub 1988/03/01.
- 217. Omary MB, Lugea A, Lowe AW, Pandol SJ. The pancreatic stellate cell: a star on the rise in pancreatic diseases. J Clin Invest. 2007 Jan;117(1):50-9. PubMed PMID: 17200706. Pubmed Central PMCID: 1716214.

- Reeves SA, Helman LJ, Allison A, Israel MA. Molecular cloning and primary structure of human glial fibrillary acidic protein. Proc Natl Acad Sci U S A. 1989 Jul;86(13):5178-82. PubMed PMID: 2740350. Pubmed Central PMCID: 297581. Epub 1989/07/01.
- Eng LF, Vanderhaeghen JJ, Bignami A, Gerstl B. An acidic protein isolated from fibrous astrocytes. Brain Res. 1971 May 7;28(2):351-4. PubMed PMID: 5113526. Epub 1971/05/07.
- 220. Eng LF, Ghirnikar RS, Lee YL. Glial fibrillary acidic protein: GFAP-thirty-one years (1969-2000). Neurochem Res. 2000 Oct;25(9-10):1439-51. PubMed PMID: 11059815. Epub 2000/11/04.
- Wallin A, Blennow K, Rosengren LE. Glial fibrillary acidic protein in the cerebrospinal fluid of patients with dementia. Dementia. 1996 Sep-Oct;7(5):267-72. PubMed PMID: 8872418. Epub 1996/09/01.
- 222. Tullberg M, Rosengren L, Blomsterwall E, Karlsson JE, Wikkelso C. CSF neurofilament and glial fibrillary acidic protein in normal pressure hydrocephalus. Neurology. 1998 Apr;50(4):1122-7. PubMed PMID: 9566405. Epub 1998/05/05.
- Rosengren LE, Lycke J, Andersen O. Glial fibrillary acidic protein in CSF of multiple sclerosis patients: relation to neurological deficit. J Neurol Sci. 1995 Nov;133(1-2):61-5. PubMed PMID: 8583233. Epub 1995/11/01.
- 224. Nylen K, Ost M, Csajbok LZ, Nilsson I, Blennow K, Nellgard B, et al. Increased serum-GFAP in patients with severe traumatic brain injury is related to outcome. J Neurol Sci. 2006 Jan 15;240(1-2):85-91. PubMed PMID: 16266720. Epub 2005/11/04.
- Nylen K, Csajbok LZ, Ost M, Rashid A, Blennow K, Nellgard B, et al. Serum glial fibrillary acidic protein is related to focal brain injury and outcome after aneurysmal subarachnoid hemorrhage. Stroke. 2007 May;38(5):1489-94. PubMed PMID: 17395862. Epub 2007/03/31.
- Tibbling G, Link H, Ohman S. Principles of albumin and IgG analyses in neurological disorders. I. Establishment of reference values. Scand J Clin Lab Invest. 1977 Sep;37(5):385-90. PubMed PMID: 337459.
- Reiber H. Cerebrospinal fluid--physiology, analysis and interpretation of protein patterns for diagnosis of neurological diseases. Mult Scler. 1998 Jun;4(3):99-107. PubMed PMID: 9762655. Epub 1998/10/08.
- 228. Andersson M, Alvarez-Cermeno J, Bernardi G, Cogato I, Fredman P, Frederiksen J, et al. Cerebrospinal fluid in the diagnosis of multiple sclerosis: a consensus report. J Neurol Neurosurg Psychiatry. 1994 Aug;57(8):897-902. PubMed PMID: 8057110. Pubmed Central PMCID: 1073070. Epub 1994/08/01.
- 229. Tourtellotte W. On cerebrospinal fluid immunoglobulin-G (IgG) quotients in multiple sclerosis and other diseases. A review and a new formula to estimate the amount of IgG synthesized per day by the central nervous system. J Neurol Sci. 1970 Mar;10(3):279-304. PubMed PMID: 4909729. Epub 1970/03/01.
- 230. Legler DF, Loetscher M, Roos RS, Clark-Lewis I, Baggiolini M, Moser B. B cell-attracting chemokine 1, a human CXC chemokine expressed in lymphoid tissues, selectively attracts B lymphocytes via BLR1/CXCR5. J Exp Med. 1998 Feb 16;187(4):655-60. Pub-Med PMID: 9463416. Pubmed Central PMCID: 2212150. Epub 1998/03/28.
- 231. Ansel KM, Harris RB, Cyster JG. CXCL13 is required for B1 cell homing, natural antibody production, and body cavity immunity. Immunity. 2002 Jan;16(1):67-76. PubMed PMID: 11825566. Epub 2002/02/05.

- 232. de Leval L, Rickman DS, Thielen C, Reynies A, Huang YL, Delsol G, et al. The gene expression profile of nodal peripheral T-cell lymphoma demonstrates a molecular link between angioimmunoblastic T-cell lymphoma (AITL) and follicular helper T (TFH) cells. Blood. 2007 Jun 1;109(11):4952-63. PubMed PMID: 17284527. Epub 2007/02/08.
- Ljostad U, Mygland A. CSF B--lymphocyte chemoattractant (CXCL13) in the early diagnosis of acute Lyme neuroborreliosis. J Neurol. 2008 May;255(5):732-7. PubMed PMID: 18344056.
- Khademi M, Kockum I, Andersson ML, Iacobaeus E, Brundin L, Sellebjerg F, et al. Cerebrospinal fluid CXCL13 in multiple sclerosis: a suggestive prognostic marker for the disease course. Mult Scler. 2011 Mar;17(3):335-43. PubMed PMID: 21135023. Epub 2010/12/08.
- 235. Brettschneider J, Czerwoniak A, Senel M, Fang L, Kassubek J, Pinkhardt E, et al. The chemokine CXCL13 is a prognostic marker in clinically isolated syndrome (CIS). PLoS One. 2010;5(8):e11986. PubMed PMID: 20700489. Pubmed Central PMCID: 2916843.
- 236. Sellebjerg F, Bornsen L, Khademi M, Krakauer M, Olsson T, Frederiksen JL, et al. Increased cerebrospinal fluid concentrations of the chemokine CXCL13 in active MS. Neurology. 2009 Dec 8;73(23):2003-10. PubMed PMID: 19996075. Epub 2009/12/10.
- 237. Link H. Immunoglobulin G and low molecular weight proteins in human cerebrospinal fluid. Chemical and immunological characterisation with special reference to multiple sclerosis. Acta Neurol Scand. 1967;43:Suppl 28:1-136. PubMed PMID: 4167871. Epub 1967/01/01.
- Olsson T, Kostulas V, Link H. Improved detection of oligoclonal IgG in cerebrospinal fluid by isoelectric focusing in agarose, double-antibody peroxidase labeling, and avidinbiotin amplification. Clin Chem. 1984 Jul;30(7):1246-9. PubMed PMID: 6375900. Epub 1984/07/01.
- Kostulas VK, Link H, Lefvert AK. Oligoclonal IgG bands in cerebrospinal fluid. Principles for demonstration and interpretation based on findings in 1114 neurological patients. Arch Neurol. 1987 Oct;44(10):1041-4. PubMed PMID: 3632376. Epub 1987/10/01.
- 240. Cepok S, Rosche B, Grummel V, Vogel F, Zhou D, Sayn J, et al. Short-lived plasma blasts are the main B cell effector subset during the course of multiple sclerosis. Brain. 2005 Jul;128(Pt 7):1667-76. PubMed PMID: 15800022. Epub 2005/04/01.
- 241. Winges KM, Gilden DH, Bennett JL, Yu X, Ritchie AM, Owens GP. Analysis of multiple sclerosis cerebrospinal fluid reveals a continuum of clonally related antibody-secreting cells that are predominantly plasma blasts. J Neuroimmunol. 2007 Dec;192(1-2):226-34. PubMed PMID: 17997491. Epub 2007/11/13.
- Kabat EA, Moore DH, Landow H. An Electrophoretic Study of the Protein Components in Cerebrospinal Fluid and Their Relationship to the Serum Proteins. J Clin Invest. 1942 Sep;21(5):571-7. PubMed PMID: 16694947. Pubmed Central PMCID: 435175. Epub 1942/09/01.
- 243. Reiber H, Ungefehr S, Jacobi C. The intrathecal, polyspecific and oligoclonal immune response in multiple sclerosis. Mult Scler. 1998 Jun;4(3):111-7. PubMed PMID: 9762657. Epub 1998/10/08.
- 244. McLean BN, Luxton RW, Thompson EJ. A study of immunoglobulin G in the cerebrospinal fluid of 1007 patients with suspected neurological disease using isoelectric focusing and the Log IgG-Index. A comparison and diagnostic applications. Brain. 1990 Oct;113 ( Pt 5):1269-89. PubMed PMID: 2245296. Epub 1990/10/01.

- 245. Tourtellotte WW, Murthy K, Brandes D, Sajben N, Ma B, Comiso P, et al. Schemes to eradicate the multiple sclerosis central nervous system immune reaction. Neurology. 1976 Jun;26(6 PT 2):59-61. PubMed PMID: 179031. Epub 1976/06/01.
- 246. Lourenco P, Shirani A, Saeedi J, Oger J, Schreiber WE, Tremlett H. Oligoclonal bands and cerebrospinal fluid markers in multiple sclerosis: associations with disease course and progression. Mult Scler. Sep 7. PubMed PMID: 22961214. Epub 2012/09/11.
- 247. Rudick RA, Cookfair DL, Simonian NA, Ransohoff RM, Richert JR, Jacobs LD, et al. Cerebrospinal fluid abnormalities in a phase III trial of Avonex (IFNbeta-1a) for relapsing multiple sclerosis. The Multiple Sclerosis Collaborative Research Group. J Neuroimmunol. 1999 Jan 1;93(1-2):8-14. PubMed PMID: 10378864. Epub 1999/06/23.
- Blennow K, Hampel H, Weiner M, Zetterberg H. Cerebrospinal fluid and plasma biomarkers in Alzheimer disease. Nat Rev Neurol. 2010 Mar;6(3):131-44. PubMed PMID: 20157306. Epub 2010/02/17.
- Andreasson U, Portelius E, Andersson ME, Blennow K, Zetterberg H. Aspects of betaamyloid as a biomarker for Alzheimer's disease. Biomark Med. 2007 Jun;1(1):59-78. PubMed PMID: 20477461. Epub 2007/06/01.
- Hu X, Hicks CW, He W, Wong P, Macklin WB, Trapp BD, et al. Bace1 modulates myelination in the central and peripheral nervous system. Nat Neurosci. 2006 Dec;9(12):1520-5. PubMed PMID: 17099708. Epub 2006/11/14.
- 251. Willem M, Garratt AN, Novak B, Citron M, Kaufmann S, Rittger A, et al. Control of peripheral nerve myelination by the beta-secretase BACE1. Science. 2006 Oct 27;314(5799):664-6. PubMed PMID: 16990514. Epub 2006/09/23.
- 252. Taveggia C, Thaker P, Petrylak A, Caporaso GL, Toews A, Falls DL, et al. Type III neuregulin-1 promotes oligodendrocyte myelination. Glia. 2008 Feb;56(3):284-93. PubMed PMID: 18080294. Epub 2007/12/15.
- 253. Zhong Z, Ewers M, Teipel S, Burger K, Wallin A, Blennow K, et al. Levels of beta-secretase (BACE1) in cerebrospinal fluid as a predictor of risk in mild cognitive impairment. Arch Gen Psychiatry. 2007 Jun;64(6):718-26. PubMed PMID: 17548753.
- 254. Zetterberg H, Andreasson U, Hansson O, Wu G, Sankaranarayanan S, Andersson ME, et al. Elevated cerebrospinal fluid BACE1 activity in incipient Alzheimer disease. Arch Neurol. 2008 Aug;65(8):1102-7. PubMed PMID: 18695061.
- 255. Guglielmotto M, Aragno M, Autelli R, Giliberto L, Novo E, Colombatto S, et al. The upregulation of BACE1 mediated by hypoxia and ischemic injury: role of oxidative stress and HIF1alpha. J Neurochem. 2009 Feb;108(4):1045-56. PubMed PMID: 19196431.
- 256. Walker KR, Kang EL, Whalen MJ, Shen Y, Tesco G. Depletion of GGA1 and GGA3 mediates postinjury elevation of BACE1. J Neurosci. 2012 Jul 25;32(30):10423-37. Pub-Med PMID: 22836275. Pubmed Central PMCID: 3490187.
- 257. Hampel H, Frank R, Broich K, Teipel SJ, Katz RG, Hardy J, et al. Biomarkers for Alzheimer's disease: academic, industry and regulatory perspectives. Nat Rev Drug Discov. 2010 Jul;9(7):560-74. PubMed PMID: 20592748.
- Mattsson N, Bremell D, Anckarsater R, Blennow K, Anckarsater H, Zetterberg H, et al. Neuroinflammation in Lyme neuroborreliosis affects amyloid metabolism. BMC Neurol. 2010;10:51. PubMed PMID: 20569437. Pubmed Central PMCID: 2902447. Epub 2010/06/24.

- 259. Gisslen M, Krut J, Andreasson U, Blennow K, Cinque P, Brew BJ, et al. Amyloid and tau cerebrospinal fluid biomarkers in HIV infection. BMC Neurol. 2009;9:63. PubMed PMID: 20028512. Pubmed Central PMCID: 2807422. Epub 2009/12/24.
- Sjogren M, Gisslen M, Vanmechelen E, Blennow K. Low cerebrospinal fluid beta-amyloid 42 in patients with acute bacterial meningitis and normalization after treatment. Neurosci Lett. 2001 Nov 13;314(1-2):33-6. PubMed PMID: 11698140. Epub 2001/11/08.
- 261. Trysberg E, Hoglund K, Svenungsson E, Blennow K, Tarkowski A. Decreased levels of soluble amyloid beta-protein precursor and beta-amyloid protein in cerebrospinal fluid of patients with systemic lupus erythematosus. Arthritis Res Ther. 2004;6(2):R129-36. PubMed PMID: 15059276. Pubmed Central PMCID: 400431. Epub 2004/04/03.
- McDonald WI, Compston A, Edan G, Goodkin D, Hartung HP, Lublin FD, et al. Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the diagnosis of multiple sclerosis. Ann Neurol. 2001 Jul;50(1):121-7. PubMed PMID: 11456302. Epub 2001/07/18.
- 263. Polman CH, Reingold SC, Edan G, Filippi M, Hartung HP, Kappos L, et al. Diagnostic criteria for multiple sclerosis: 2005 revisions to the "McDonald Criteria". Ann Neurol. 2005 Dec;58(6):840-6. PubMed PMID: 16283615.
- 264. Gunnarsson M, Malmestrom C, Axelsson M, Sundstrom P, Dahle C, Vrethem M, et al. Axonal damage in relapsing multiple sclerosis is markedly reduced by natalizumab. Ann Neurol. 2011 Jan;69(1):83-9. PubMed PMID: 21280078. Epub 2011/02/01.
- Anckarsater R, Vasic N, Jideus L, Kristiansson M, Zetterberg H, Blennow K, et al. Cerebrospinal fluid protein reactions during non-neurological surgery. Acta Neurol Scand. 2007 Apr;115(4):254-9. PubMed PMID: 17376123.
- 266. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. Arthritis Rheum. 1997 Sep;40(9):1725. PubMed PMID: 9324032.
- Breitbach SA, Alexander RW, Daltroy LH, Liang MH, Boll TJ, Karlson EW, et al. Determinants of cognitive performance in systemic lupus erythematosus. J Clin Exp Neuropsychol. 1998 Apr;20(2):157-66. PubMed PMID: 9777469.
- Trysberg E, Nylen K, Rosengren LE, Tarkowski A. Neuronal and astrocytic damage in systemic lupus erythematosus patients with central nervous system involvement. Arthritis Rheum. 2003 Oct;48(10):2881-7. PubMed PMID: 14558094.
- 269. Teunissen CE, Petzold A, Bennett JL, Berven FS, Brundin L, Comabella M, et al. A consensus protocol for the standardization of cerebrospinal fluid collection and biobanking. Neurology. 2009 Dec 1;73(22):1914-22. PubMed PMID: 19949037. Pubmed Central PMCID: 2839806. Epub 2009/12/02.
- Rosengren LE, Wikkelso C, Hagberg L. A sensitive ELISA for glial fibrillary acidic protein: application in CSF of adults. J Neurosci Methods. 1994 Mar;51(2):197-204. PubMed PMID: 8051950.
- 271. Blennow K, Wallin A, Fredman P, Gottfries CG, Karlsson I, Svennerholm L. Intrathecal synthesis of immunoglobulins in patients with Alzheimer's disease. Eur Neuropsychopharmacol. 1990 Nov;1(1):79-81. PubMed PMID: 2136219.
- 272. Wu G, Sankaranarayanan S, Tugusheva K, Kahana J, Seabrook G, Shi XP, et al. Decrease in age-adjusted cerebrospinal fluid beta-secretase activity in Alzheimer's subjects. Clin Biochem. 2008 Aug;41(12):986-96. PubMed PMID: 18489907. Epub 2008/05/21.

- 273. Olsson A, Vanderstichele H, Andreasen N, De Meyer G, Wallin A, Holmberg B, et al. Simultaneous measurement of beta-amyloid(1-42), total tau, and phosphorylated tau (Thr181) in cerebrospinal fluid by the xMAP technology. Clin Chem. 2005 Feb;51(2):336-45. PubMed PMID: 15563479. Epub 2004/11/26.
- 274. Portelius E, Price E, Brinkmalm G, Stiteler M, Olsson M, Persson R, et al. A novel pathway for amyloid precursor protein processing. Neurobiol Aging. 2011 Jun;32(6):1090-8. PubMed PMID: 19604603. Epub 2009/07/17.
- 275. Portelius E, Westman-Brinkmalm A, Zetterberg H, Blennow K. Determination of beta-amyloid peptide signatures in cerebrospinal fluid using immunoprecipitation-mass spectrometry. J Proteome Res. 2006 Apr;5(4):1010-6. PubMed PMID: 16602710. Epub 2006/04/11.
- Portelius E, Tran AJ, Andreasson U, Persson R, Brinkmalm G, Zetterberg H, et al. Characterization of amyloid beta peptides in cerebrospinal fluid by an automated immunoprecipitation procedure followed by mass spectrometry. J Proteome Res. 2007 Nov;6(11):4433-9. PubMed PMID: 17927230. Epub 2007/10/12.
- Rocca MA, Messina R, Filippi M. Multiple sclerosis imaging: recent advances. J Neurol. Dec 21. PubMed PMID: 23263475. Epub 2012/12/25.
- 278. Franciotta D, Martino G, Zardini E, Furlan R, Bergamaschi R, Andreoni L, et al. Serum and CSF levels of MCP-1 and IP-10 in multiple sclerosis patients with acute and stable disease and undergoing immunomodulatory therapies. J Neuroimmunol. 2001 Apr 2;115(1-2):192-8. PubMed PMID: 11282170.
- Malmestrom C, Andersson BA, Haghighi S, Lycke J. IL-6 and CCL2 levels in CSF are associated with the clinical course of MS: implications for their possible immunopathogenic roles. J Neuroimmunol. 2006 Jun;175(1-2):176-82. PubMed PMID: 16626811. Epub 2006/04/22.
- Sindern E, Niederkinkhaus Y, Henschel M, Ossege LM, Patzold T, Malin JP. Differential release of beta-chemokines in serum and CSF of patients with relapsing-remitting multiple sclerosis. Acta Neurol Scand. 2001 Aug;104(2):88-91. PubMed PMID: 11493224.
- Uccelli A, Pedemonte E, Narciso E, Mancardi G. Biological markers of the inflammatory phase of multiple sclerosis. Neurol Sci. 2003 Dec;24 Suppl 5:S271-4. PubMed PMID: 14652787.
- Kuhle J, Leppert D, Petzold A, Regeniter A, Schindler C, Mehling M, et al. Neurofilament heavy chain in CSF correlates with relapses and disability in multiple sclerosis. Neurology. 2011 Apr 5;76(14):1206-13. PubMed PMID: 21346223.
- Sellebjerg F, Christiansen M, Nielsen PM, Frederiksen JL. Cerebrospinal fluid measures of disease activity in patients with multiple sclerosis. Mult Scler. 1998 Dec;4(6):475-9. PubMed PMID: 9987755.
- 284. Lamers KJ, de Reus HP, Jongen PJ. Myelin basic protein in CSF as indicator of disease activity in multiple sclerosis. Mult Scler. 1998 Jun;4(3):124-6. PubMed PMID: 9762659.
- 285. Lim ET, Sellebjerg F, Jensen CV, Altmann DR, Grant D, Keir G, et al. Acute axonal damage predicts clinical outcome in patients with multiple sclerosis. Mult Scler. 2005 Oct;11(5):532-6. PubMed PMID: 16193890.
- Lycke JN, Karlsson JE, Andersen O, Rosengren LE. Neurofilament protein in cerebrospinal fluid: a potential marker of activity in multiple sclerosis. J Neurol Neurosurg Psychiatry. 1998 Mar;64(3):402-4. PubMed PMID: 9527161. Pubmed Central PMCID: 2170011. Epub 1998/04/04.

- Petzold A, Eikelenboom MJ, Keir G, Grant D, Lazeron RH, Polman CH, et al. Axonal damage accumulates in the progressive phase of multiple sclerosis: three year follow up study. J Neurol Neurosurg Psychiatry. 2005 Feb;76(2):206-11. PubMed PMID: 15654034. Pubmed Central PMCID: 1739484.
- Salzer J, Svenningsson A, Sundstrom P. Neurofilament light as a prognostic marker in multiple sclerosis. Mult Scler. 2010 Mar;16(3):287-92. PubMed PMID: 20086018. Epub 2010/01/21.
- Petzold A, Eikelenboom MJ, Gveric D, Keir G, Chapman M, Lazeron RH, et al. Markers for different glial cell responses in multiple sclerosis: clinical and pathological correlations. Brain. 2002 Jul;125(Pt 7):1462-73. PubMed PMID: 12076997.
- Link H, Huang YM. Oligoclonal bands in multiple sclerosis cerebrospinal fluid: an update on methodology and clinical usefulness. J Neuroimmunol. 2006 Nov;180(1-2):17-28. PubMed PMID: 16945427.
- Koch M, Heersema D, Mostert J, Teelken A, De Keyser J. Cerebrospinal fluid oligoclonal bands and progression of disability in multiple sclerosis. Eur J Neurol. 2007 Jul;14(7):797-800. PubMed PMID: 17594338.
- Saiz A, Carreras E, Berenguer J, Yague J, Martinez C, Marin P, et al. MRI and CSF oligoclonal bands after autologous hematopoietic stem cell transplantation in MS. Neurology. 2001 Apr 24;56(8):1084-9. PubMed PMID: 11320183.
- Piccio L, Naismith RT, Trinkaus K, Klein RS, Parks BJ, Lyons JA, et al. Changes in Band T-lymphocyte and chemokine levels with rituximab treatment in multiple sclerosis. Arch Neurol. 2010 Jun;67(6):707-14. PubMed PMID: 20558389. Pubmed Central PM-CID: 2918395.
- 294. Harrer A, Tumani H, Niendorf S, Lauda F, Geis C, Weishaupt A, et al. Cerebrospinal fluid parameters of B cell-related activity in patients with active disease during natalizumab therapy. Mult Scler. 2012 Oct 23. PubMed PMID: 23093485. Epub 2012/10/25.
- 295. von Glehn F, Farias AS, de Oliveira AC, Damasceno A, Longhini AL, Oliveira EC, et al. Disappearance of cerebrospinal fluid oligoclonal bands after natalizumab treatment of multiple sclerosis patients. Mult Scler. 2012 Jul;18(7):1038-41. PubMed PMID: 22041091.
- 296. Losseff NA, Wang L, Lai HM, Yoo DS, Gawne-Cain ML, McDonald WI, et al. Progressive cerebral atrophy in multiple sclerosis. A serial MRI study. Brain. 1996 Dec;119 (Pt 6):2009-19. PubMed PMID: 9010005. Epub 1996/12/01.
- 297. Petzold A, Michel P, Stock M, Schluep M. Glial and axonal body fluid biomarkers are related to infarct volume, severity, and outcome. J Stroke Cerebrovasc Dis. 2008 Jul-Aug;17(4):196-203. PubMed PMID: 18589339.
- Pekny M, Nilsson M. Astrocyte activation and reactive gliosis. Glia. 2005 Jun;50(4):427-34. PubMed PMID: 15846805.
- 299. Bar-Or A, Calabresi PA, Arnold D, Markowitz C, Shafer S, Kasper LH, et al. Rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial. Ann Neurol. 2008 Mar;63(3):395-400. PubMed PMID: 18383069.
- 300. Hauser SL, Waubant E, Arnold DL, Vollmer T, Antel J, Fox RJ, et al. B-cell depletion with rituximab in relapsing-remitting multiple sclerosis. N Engl J Med. 2008 Feb 14;358(7):676-88. PubMed PMID: 18272891. Epub 2008/02/15.

- Kappos L, Li D, Calabresi PA, O'Connor P, Bar-Or A, Barkhof F, et al. Ocrelizumab in relapsing-remitting multiple sclerosis: a phase 2, randomised, placebo-controlled, multicentre trial. Lancet. 2011 Nov 19;378(9805):1779-87. PubMed PMID: 22047971.
- 302. Prineas JW. Multiple sclerosis: presence of lymphatic capillaries and lymphoid tissue in the brain and spinal cord. Science. 1979 Mar 16;203(4385):1123-5. PubMed PMID: 424741.
- 303. Vandvik B. Oligoclonal IgG and free light chains in the cerebrospinal fluid of patients with multiple sclerosis and infectious diseases of the central nervous system. Scand J Immunol. 1977;6(9):913-22. PubMed PMID: 410092.
- Olsson JE, Link H. Immunoglobulin abnormalities in multiple sclerosis. Relation to clinical parameters: exacerbations and remissions. Arch Neurol. 1973 Jun;28(6):392-9. Pub-Med PMID: 4121785.
- 305. Delmotte P, Gonsette R. Biochemical findings in multiple sclerosis IV. Isoelectric focusing of the CSF gamma globulins in multiple sclerosis (262 cases) and other neurological diseases (272 cases). J Neurol. 1977 Apr 28;215(1):27-37. PubMed PMID: 67197.
- Mattson DH, Roos RP, Arnason BG. Isoelectric focusing of IgG eluted from multiple sclerosis and subacute sclerosing panencephalitis brains. Nature. 1980 Sep 25;287(5780):335-7. PubMed PMID: 7421992.
- 307. Cameron EM, Spencer S, Lazarini J, Harp CT, Ward ES, Burgoon M, et al. Potential of a unique antibody gene signature to predict conversion to clinically definite multiple sclerosis. J Neuroimmunol. 2009 Aug 18;213(1-2):123-30. PubMed PMID: 19631394. Pubmed Central PMCID: 2785005.
- 308. Ligocki AJ, Lovato L, Xiang D, Guidry P, Scheuermann RH, Willis SN, et al. A unique antibody gene signature is prevalent in the central nervous system of patients with multiple sclerosis. J Neuroimmunol. 2010 Sep 14;226(1-2):192-3. PubMed PMID: 20655601. Pubmed Central PMCID: 2937103.
- 309. Meinl E, Krumbholz M, Hohlfeld R. B lineage cells in the inflammatory central nervous system environment: migration, maintenance, local antibody production, and therapeutic modulation. Ann Neurol. 2006 Jun;59(6):880-92. PubMed PMID: 16718690.
- Franklin RJ, Ffrench-Constant C. Remyelination in the CNS: from biology to therapy. Nat Rev Neurosci. 2008 Nov;9(11):839-55. PubMed PMID: 18931697.
- 311. Mori F, Rossi S, Sancesario G, Codeca C, Mataluni G, Monteleone F, et al. Cognitive and cortical plasticity deficits correlate with altered amyloid-beta CSF levels in multiple sclerosis. Neuropsychopharmacology. 2011 Feb;36(3):559-68. PubMed PMID: 20944553. Pubmed Central PMCID: 3055691.
- 312. Mai W, Hu X, Lu Z, Peng F, Wang Y. Cerebrospinal fluid levels of soluble amyloid precursor protein and beta-amyloid 42 in patients with multiple sclerosis, neuromyelitis optica and clinically isolated syndrome. J Int Med Res. 2011;39(6):2402-13. PubMed PMID: 22289560.
- Dal Bianco A, Bradl M, Frischer J, Kutzelnigg A, Jellinger K, Lassmann H. Multiple sclerosis and Alzheimer's disease. Ann Neurol. 2008 Feb;63(2):174-83. PubMed PMID: 17924575.
- 314. Cirrito JR, Kang JE, Lee J, Stewart FR, Verges DK, Silverio LM, et al. Endocytosis is required for synaptic activity-dependent release of amyloid-beta in vivo. Neuron. 2008 Apr 10;58(1):42-51. PubMed PMID: 18400162. Pubmed Central PMCID: 2390913.

- 315. Mattsson N, Johansson P, Hansson O, Wallin A, Johansson JO, Andreasson U, et al. Converging pathways of chromogranin and amyloid metabolism in the brain. J Alzheimers Dis. 2010;20(4):1039-49. PubMed PMID: 20413871.
- 316. Wake H, Lee PR, Fields RD. Control of local protein synthesis and initial events in myelination by action potentials. Science. 2011 Sep 16;333(6049):1647-51. PubMed PMID: 21817014. Pubmed Central PMCID: 3482340.