The Nordic Expert Group for Criteria Documentation of Health Risks from Chemicals

# 138. Microbial volatile organic compounds (MVOCs)

Anne Korpi, Jill Järnberg and Anna-Liisa Pasanen

ARBETE OCH HÄLSA ISBN 978-91-7045-815-6 vetenskaplig skriftserie ISSN 0346-7821



#### Arbete och Hälsa

Arbete och Hälsa (Work and Health) is a scientific report series published by the National Institute for Working Life. The series presents research by the Institute's own researchers as well as by others, both within and outside of Sweden. The series publishes scientific original works, dissertations, criteria documents and literature surveys.

Arbete och Hälsa has a broad targetgroup and welcomes articles in different areas.

Summaries in Swedish and English as well as the complete original text are available at www.arbetslivsinstitutet.se/ as from 1997.

#### ARBETE OCH HÄLSA

Editor-in-chief: Staffan Marklund Co-editors: Marita Christmansson, Kjell Holmberg, Birgitta Meding, Bo Melin and Ewa Wigaeus Tornqvist

© National Institute for Working Life & authors 2007

National Institute for Working Life, S-113 91 Stockholm, Sweden

ISBN 978-91-7045-815-6 ISSN 0346-7821 http://www.arbetslivsinstitutet.se/ Printed at Elanders Gotab, Stockholm

# Preface

The main task of the Nordic Expert Group for Criteria Documentation of Health Risks from Chemicals (NEG) is to produce criteria documents to be used by the regulatory authorities as the scientific basis for setting occupational exposure limits for chemical substances.

For each document NEG appoints one or several authors. Evaluation is made of all relevant published, peer-reviewed original literature found. The document aims at establishing dose-response/dose-effect relationships and defining a critical effect. No numerical values for occupational exposure limits are proposed.

Whereas NEG adopts the document by consensus procedures, thereby granting the quality and conclusions, the authors are responsible for the factual content of the document.

The evaluation of the literature and the drafting of this document on *Microbial volatile organic compounds (MVOCs)* were made by Dr. Anne Korpi at the University of Kuopio, Finland, Dr. Jill Järnberg at the National Institute for Working Life, Sweden, and Prof. Anna-Liisa Pasanen at the Finnish Institute of Occupational Health, Finland. The draft document was discussed within the group and the final version was accepted by NEG on November 28, 2006. The following individuals participated in the elaboration of the document:

Gunnar Johanson	Institute of Environmental Medicine, Karolinska Institutet and National Institute for Working Life, Sweden (chairman)
Maria Albin	Department of Occupational and Environmental Medicine, University Hospital, Lund, Sweden (NEG expert)
Karin Sørig Hougaard	National Institute of Occupational Health, Denmark (NEG expert)
Kristina Kjærheim	Cancer Registry of Norway, Norway (NEG expert)
Vidir Kristjansson	Administration of Occupational Safety and Health, Iceland (former NEG expert)
Kai Savolainen	Finnish Institute of Occupational Health, Finland (NEG expert)
Vidar Skaug	National Institute of Occupational Health, Norway (NEG expert)
Jill Järnberg and Anna-Karin Alexandrie	National Institute for Working Life, Sweden (NEG secretariat)

Editorial work and technical editing were performed by the NEG secretariat. This work was financially supported by the Swedish National Institute for Working Life, the Norwegian Ministry of Labour and Social Inclusion and the Nordic Council of Ministers.

All criteria document produced by the Nordic Expert Group may be down-loaded from *www.nordicexpertgroup.org*.

Gunnar Johanson, Chairman of NEG

Contents

Preface	
Abbreviations and acronyms	
1. Introduction	1
2. Substance identification	2
3. Physical and chemical properties	15
4. Occurrence	18
5. Measurements and analysis of MVOCs	19
6. Exposure data	20
7. Toxicokinetics	23
8. Biological monitoring	24
9. Mechanism of toxicity	25
10. Effects in animals and <i>in vitro</i> studies	26
10.1 Irritation and sensitisation	26
10.2 Effects of single exposure	28
10.3 Effects of short-term and long-term exposure 10.4 Mutagenicity and genotoxicity	30 32
10.5 Carcinogenicity	33
10.6 Reproductive and developmental studies	41
11. Observations in man	42
11.1 Odour sensation, irritation and sensitisation	42
11.2 Effects of single and short-term exposure	45
11.3 Effects of long-term exposure 11.4 Genotoxic effects	47 48
11.5 Carcinogenic effects	48
11.6 Reproductive and developmental effects	48
12. Dose-effect and dose-response relationships	48
12.1 Animal data	48
12.2 Human data	49
12.3 Extrapolation of animal data on sensory irritation responses to human	
13. Previous evaluations by national and international bodies	55
14. Evaluation of human health risks	59
14.1 Assessment of health risks	59 50
14.2 Groups at extra risk 14.3 Scientific basis for an occupational exposure limit	59 59
15. Research needs	60
16. Summary	61
17. Summary in Swedish	63
18. References	65
19. Data bases used in search of literature	78

# Abbreviations and acronyms

ACGIH	American Conference of Governmental Industrial Hygienists
ASTM	American Society for Testing of Materials
DNPH	2,4-dinitrophenyl hydrazine
FID	flame ionisation detector
GC	gas chromatography
IARC	International Agency for Research on Cancer
$LC_{50}$	lethal concentration for 50 % of the animals at single exposure
LCLo	lowest observed lethal concentration
$LD_{50}$	lethal dose for 50 % of the exposed animals at single administration
LOEL	lowest observed effect level
MAK	Maximale Arbeitsplatz-Konzentration
MVOC	microbial volatile organic compound
MS	mass spectrometry
NOEL	no observed effect level
NTP	National Toxicology Program
OEL	occupational exposure limit
PMN	polymorphonuclear neutrophil
PP	pyrophosphate
RD <sub>50</sub>	concentration associated with a 50 $\%$ decrease in the respiratory rate
RIL	recommended indoor air level
SPME	solid-phase microextraction
STEL	short-term exposure limit
TLV®	threshold limit value
TWA	time weighted average
VOC	volatile organic compound

#### 1. Introduction

Microbial volatile organic compounds (MVOCs) are produced in the metabolism of microorganisms such as fungi and bacteria. They are formed during both the primary metabolism (from the synthesis of e.g. DNA and amino and fatty acids) and the secondary metabolism (from intermediates of the primary metabolism) as side-products, mainly in the metabolic oxidation of glucose from various intermediates (23). Thus, the production of MVOCs is greatly affected by microbial species, growth phase and conditions (nutrients, pH, humidity, temperature) (19, 125, 216). More than 200 compounds have been regarded as MVOCs in the literature. The compounds also have other environmental sources than microbial metabolism. Thus, compounds originating solely from microbial metabolism hardly exist.

The interest to utilise MVOCs as indicators of biocontamination was originally raised by the food-processing industry in the 1970s, when analysis of unpleasantly smelling MVOCs was suggested to be a practical and rapid tool to detect undesirable or spoilage processes caused by microorganisms during the storage or processing of foodstuffs (36, 38, 53, 54, 99, 100, 131, 147, 226). Later, MVOC analyses and profiles were applied to the taxonomy research to identify and separate microbial (mainly fungal) species or strains (71, 95, 103, 126, 127, 222, 235). MVOCs were analysed in indoor air environments for the first time in the 1990s (20, 149, 197, 210, 211, 224); with MVOC analysis, a possibility to detect hidden microbial growth behind interior surfaces without opening building structures was presented. It was assumed that, as gases, MVOCs may enter the indoor air (e.g. through water vapour barriers) more easily than spores (135, 197, 213). The concern about possible health risks related to MVOC exposure in indoor environments was also raised in the 1990s. As eye and upper respiratory tract irritation was frequently reported by occupants in buildings with moisture and mould damage, these symptoms were concluded to be associated with exposure to irritative substances of microbial origin (34). Interestingly enough, much less attention has been paid to MVOCs and their possible adverse health effects in work environments with productive microbial sources or high levels of contamination, where occurrence of at least some MVOCs is obviously more abundant than in indoor environments.

This document reviews the literature on compounds most frequently denoted MVOCs. From 96 typical MVOCs listed, 15 compounds were chosen for closer toxicological evaluation (Tables 2-3); for selection criteria see page 4. The data on the individual compounds presented in this document are utterly condensed, focusing on inhalation studies and the lowest administered doses, and are largely based on toxicological reviews and TOXNET® (a collection of toxicology and environmental health databases) data. Thus, high dose effects of individual compounds are not dealt with, as they are considered irrelevant in the context of MVOCs. However, some of the compounds denoted MVOCs are also industrial chemicals. As industrial exposure levels are generally much higher than those

encountered in the MVOC context also the lowest available doses are high when compared with levels of microbial origin. Most of the concern regarding MVOC exposure has been raised for home environments. In the present document, focus is on the non-industrial working population rather than the general public, although the majority of the available data originates from dwellings.

### 2. Substance identification

MVOCs are formed during both the primary and the secondary metabolism as side-products, mainly in the metabolic oxidation of glucose, from various precursors, such as acetate, amino acids, fatty acids, and keto acids (23). The primary metabolism of microorganisms comprises the synthesis of DNA and amino and fatty acids, whereas the secondary metabolism consists of reactions following the primary metabolism. As the primary metabolism involves an interrelated series of enzyme-catalysed chemical reactions, it is basically the same for all living systems (113). Thus, for several compounds denoted MVOCs also other sources, such as vegetation and even mammalian breath, sweat, and skin emanations, have been identified (83, 203). The identified MVOCs are alcohols, ketones, terpenes, esters, lactones, hydrocarbons, aldehydes, sulphur and nitrogen compounds (93, 126, 221). The complex metabolic pathways for MVOC formation are depicted in Figure 1 and the precursors of some common MVOCs are presented in Table 1.

For convenience, it is often stated that MVOCs are side-products of the primary metabolism of microorganisms, and mycotoxins are end-products of the seconddary metabolism. However, since the division between primary and secondary metabolism is not absolute (21), it can only be stated that MVOCs are formed during both (23). As nutritional imbalances and disorders (e.g. a lack of primary carbon and nitrogen sources) lead to expression of the secondary metabolism, changes in the nutritional state may often promote or trigger the production of several MVOCs (23, 27, 113, 205). On the other hand, it has been suggested that secondary metabolites may be inhibitors of the primary metabolism (198), and volatile metabolites of certain bacteria may stimulate mycotoxin production (18). The production of certain fungal MVOCs has also been suggested to be associated with mycotoxin production. Evidence of such relationships has been reported between the production of sequiterpenes and aflatoxins, between monoterpenes, sesquiterpenes and trichothecenes, and between ketones and ochratoxins (57, 58, 95, 169, 223, 235).

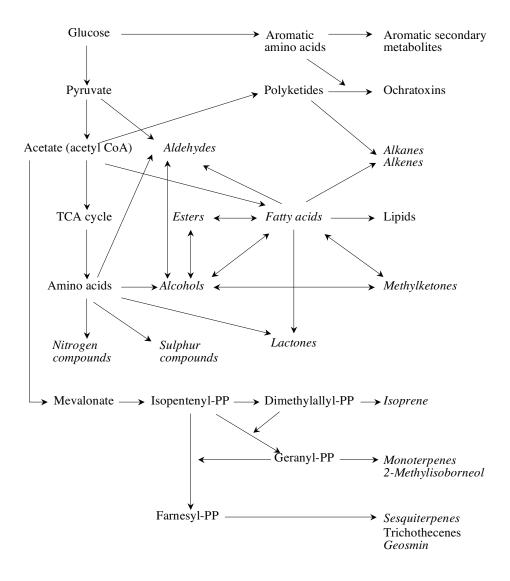
Chemical reactions in the environment may further convert the produced MVOCs to other compounds. For example, alcohols are easily oxidised to aldehydes and further to carboxylic acids (224), and ketones may react with hydroxyl radicals in the air to form aldehydes (15).

Chemical reactions may also produce compounds denoted MVOCs in the atmosphere; the reactions between ozone (and other oxidants) and unsaturated hydrocarbons (isoprenes/terpenes) have recently been investigated experimentally. The main products in these reactions are aldehydes, ketones, and organic acids,

but the intermediate products formed during the reactions have been suggested to be much more irritating than the corresponding original reactants and endproducts (208, 228, 229). For example, under humid conditions, the reaction between ozone and isoprene produces hydrogen peroxide, methacrolein, and methylvinyl ketone (179), all of which are known irritants. 3-Methylfuran is suspected to be another oxidation product of isoprene (83).

Finally, it must not be overlooked that the same compounds denoted MVOCs may also have other sources in the environment, such as building materials, human activities, traffic, foodstuffs, smoking, etc. (83, 183, 198).

So far, more than 200 individual compounds have been recognised as MVOCs in laboratory studies (93, 126, 221). The majority of the experimental studies has



**Figure 1.** Main metabolic pathways for the production of some MVOCs and mycotoxins (73, 113, 198, 205, 215). Volatile compounds are in *italics*. Abbreviations: Co A = coenzyme A, PP = pyrophosphate, TCA = tricarboxylic acid

Precursor	Volatile product(s)	Reference
Amino acids		
Alanine	Acetaldehyde	(81)
α-Amino acids	Alkyl methoxy pyrazine	(198)
Glycine	Formaldehyde	(81)
Leucine	3-Methyl-1-butanol	(23, 27, 191)
Methionine, cysteine	Dimethyl disulphide	(27, 198)
Valine	2-Methyl-1-propanol	(23, 27, 191)
Phenylalanine	Phenyl acetaldehyde, styrene	(23, 27, 81, 126, 191)
Organic acids		
Fatty acids	Alkenes and alkadienes, aldehydes methylketones with one carbon less than the original fatty acid (e.g. 2- butanone, 2-pentanone, 2-hexanone, 2-heptanone, 2-undecanone)	(81, 198, 221)
Medium-chain fatty acids	Acetates	(23)
$\gamma$ - or $\delta$ -Hydroxy acids, keto acids, long-chain fatty acids	4-Hexanolide, 6-pentyl-α-pyrone	(44, 68, 198)
Linoleic acid, linolenic acid	1-Octen-3-ol, 3-octanol, 3-octanone, hexanal, heptanal, nonanal	(27, 45, 126, 231)
Others		
Isopentenyl pyrophosphate	Terpenoid compounds: monoterpenes, sesquiterpenes and their alcohols, geosmin	(22, 27, 198, 205, 206)

Table 1. Some common MVOCs and their precursors in the microbial metabolism.

been carried out with pure cultures of selected, individual microbial species, often grown on agar, cereals and other foodstuffs, bedding materials (e.g. straw, peat, shavings) or building materials (e.g. wood, wall paper, gypsum, chip-, card- and plasterboards, insulation materials like glass and mineral wool), degradable house-hold waste, and house dust (30, 36, 37, 39, 69, 71, 72, 74, 75, 77, 95, 99, 100, 118, 123, 145, 146, 169, 182, 186, 187, 198, 200, 201, 211, 215, 220-224, 235, 236, 238). In a few studies, MVOCs produced by mixed cultures on building materials have been investigated (33, 49, 50, 64, 116, 214). In these studies, MVOCs were produced by species/strains of microbial genera common in the environment, such as *Absidia, Acremonium, Alternaria, Aspergillus, Botrytis, Candida, Chetomium, Cladosporium, Coniophora, Fusarium, Paecilomyces, Penicillium, Phialophora, Poria, Pseudomonas, Rhizopus, Saccharomyces, Serpula, Stachybotrys, Streptomyces, Trichoderma, Ulocladium and Wallemia.* 

The 15 MVOCs that were selected for closer examination in the present document are listed in Table 2 and represent compounds analysed and reported in laboratory or field studies. In these studies, the selection of compounds identified has often been limited to 10-15 due to study design and analytical restraints, and the whole range of MVOCs has not been monitored. For example, acetaldehyde, nonanal, 2-pentanone, limonene, and sesquiterpenes are among the most commonly identified microbial metabolites in laboratory experiments, still they have not been reported in field samples, probably due to non-microbial sources in the field, and analytical limitations (sesquiterpenes).

A more comprehensive list covering 96 frequently reported MVOCs including substance identification data are given in Table 3. For additional lists of compounds, the reader is referred to the publications by Jelen and Wasowics (93) and Larsen and Frisvad (126).

Thus, based on present knowledge, it is difficult to make a reliable list of relevant MVOCs. This is due to the fact that in the majority of experimental studies, control experiments are missing, as respective sterile materials and their qualitative and quantitative emissions have seldom been reported. Therefore, the concepts of VOC and MVOC overlap inasmuch as the origin of a compound reported as an MVOC may well be the emission of a substrate as well. This hampers the interpretation of the data in field settings. For example, Wilkins and Larsen (221) have suspected that toluene, xylenes, and ethyl benzene might not result from microbial metabolism, even though these compounds are often reported as MVOCs. Furthermore, an individual MVOC cannot be related to a certain microbial species, because the same MVOC may be produced by different microorganisms; e.g. bacterial and fungal species share the same MVOCs. This is natural due to the similarities in metabolism as well as growth conditions that are one of the key factors for the MVOC production in any microbial species. Finally, the methodology used for MVOC analyses varies between studies and affects the MVOC profiles reported in the literature considerably. Attempts have been made to apply principal component analysis in order to identify areas of microbial contamination relying on the VOC profiles of environmental samples (224).

Compound	-	ion factors 101.3 kPa)	Reference
	1 ppm =	$1 \text{ mg/m}^3 =$	_
2-Methyl-1-propanol	3.03 mg/m <sup>3</sup>	0.330 ppm	(197)
3-Methyl-1-butanol	3.61 mg/m <sup>3</sup>	0.277 ppm	(66, 135, 144, 148, 149, 192, 197)
3-Methyl-2-butanol	$3.61 \text{ mg/m}^3$	0.277 ppm	(197)
2-Pentanol	$3.61 \text{ mg/m}^3$	0.277 ppm	(66, 135, 144, 192, 197)
3-Octanol	$5.33 \text{ mg/m}^3$	0.188 ppm	(40, 66, 192, 197)
1-Octen-3-ol	5.24 mg/m <sup>3</sup>	0.191 ppm	(40, 66, 135, 144, 149, 164, 192, 197)
2-Octen-1-ol	$5.24 \text{ mg/m}^3$	0.191 ppm	(40, 66, 148, 149, 192, 197)
3-Methylfuran	$3.36 \text{ mg/m}^3$	0.298 ppm	(135, 144, 149, 164, 192, 197)
2-Hexanone	$4.10 \text{ mg/m}^3$	0.244 ppm	(66, 135, 197)
2-Heptanone	$4.67 \text{ mg/m}^3$	0.214 ppm	(66, 135, 148, 149, 192, 197)
3-Octanone	$5.24 \text{ mg/m}^3$	0.191 ppm	(66, 135, 144, 192, 197)
2-Methylisoborneol	6.88 mg/m <sup>3</sup>	0.145 ppm	(192, 197)
2-Isopropyl-3-methoxy-pyrazine	$6.22 \text{ mg/m}^3$	0.161 ppm	(192, 197)
Geosmin	$7.46 \text{ mg/m}^3$	0.134 ppm	(192, 197)
Dimethyl disulphide	$3.85 \text{ mg/m}^3$	0.260 ppm	(135, 144)

Table 2. Most often reported MVOCs in living environments, and conversion factors (163).

Common name	IUPAC-name	Common name IUPAC-name Synonyms (selected)	Chemical	Molecular	CAS-
or name used in			formula	weight	number
document					
Alcohols					
1-Butanol	Butan-1-ol	<i>n</i> -Butanol; <i>n</i> -butyl alcohol; propyl carbinol	$\rm C_4H_{10}O$	74.12	71-36-3
4-Decanol	Decan-4-ol		$C_{10}H_{22}O$	158.28	2051-31-2
Ethanol	Ethanol	Ethyl alcohol; ethyl hydroxide; methyl carbinol; spirit	$C_2H_6O$	46.07	64-17-5
2-Ethyl-1-hexanol	2-Ethylhex an-1-ol	2-Ethylhexanol	$C_8H_{18}O$	130.23	104-76-7
2-Heptanol	Heptan-2-ol	sec-Heptyl alcohol; 2-heptyl alcohol; isoheptyl alcohol; 2–	$\rm C_7 H_{16} O$	116.20	543-49-7
		hydroxyheptane; 1-methylhexanol; methyl pentyl carbinol; methyl <i>n</i> -amyl carbinol			
1-Hexanol	Hexan-1-ol	1-Hexyl alcohol; n-hexyl alcohol; n-hexanol; amyl carbinol	$C_{6}H_{14}O$	102.18	111-27-3
2-Methyl-1-propanol	2-Methylpropan-1-ol	1-Hydroxymethylpropane; 2-methylpropyl alcohol; isobutanol; isobutyl alcohol: isopropyl carbinol	$C_4H_{10}O$	74.12	78-83-1
2-Methyl-1-butanol	2-Methylbutan-1-ol	Sec-Butyl carbinol	$C_5H_{12}O$	88.15	137-32-6
3-Methyl-1-butanol	3-Methylbutan-1-ol	1-Hydroxy-3-methylbutane; 2-methyl-butanol-4; 3-methylbutanol; isoamyl alcohol; isobutyl carbinol; isopentanol; isopentyl alcohol	$C_5H_{12}O$	88.15	123-51-3
3-Methyl-2-butanol	3-Methylbutan-2-ol	2-Hydroxy-3-methylbutane; <i>sec</i> -isoamyl alcohol; methylisopropylcarbinol	$C_5H_{12}O$	88.15	598-75-4
1-Octanol	Octan-1-ol	1-Octyl alcohol; <i>n</i> -octanol; <i>n</i> -octyl alcohol; 1-hydroxyoctane; heptyl carbinol; caprylic alcohol	$C_8H_{18}O$	130.23	111-87-5
3-Octanol	Octan-3-ol	n-Octan-3-ol; 1-ethyl-1-hexanol; $n$ -amyl ethyl carbinol	$C_8H_{18}O$	130.23	589-98-0 and 20296-29-1
1-Octen-3-ol	Oct-1-en-3-ol	3-Octenol; octen-3-ol; vinyl hexanol; 3-hydroxy-1-octene; <i>n</i> -oct-1-en-3-ol; amyl vinyl carbinol; pentyl vinyl carbinol	$C_8H_{16}O$	128.21	3391-86-4
2-Octen-1-ol	Oct-2-en-1-ol	2-Octenol: 4-hutvl-2-huten-1-ol	C.H.O	128.21	22104-78-5

.

<b>Table 3.</b> Chemical in laboratory studies. T	dentification of the compo he 15 substances selected	<b>Table 3.</b> Chemical identification of the compounds frequently reported as MVOCs of fungi and bacteria common in the environment based on laboratory studies. The 15 substances selected for further investigation are highlighted (46, 47, 151).	the environ	ment based or	c
Common name	IUPAC-name	Synonyms (selected)	Chemical	Molecular	CAS-
or name used in document			formula	weight	number
1-Pentanol	Pentan-1-ol	1-Pentol; pentanol-1; <i>n</i> -pentan-1-ol; <i>n</i> -pentyl alcohol; <i>n</i> -pentanol; <i>n</i> -amyl alcohol; <i>n</i> -butyl carbinol	$C_5H_{12}O$	88.15	71-41-0
2-Pentanol	Pentan-2-ol	Pentanol-2; sec-amyl alcohol; methyl propyl carbinol; sec-pentyl alcohol	$C_5H_{12}O$	88.15	6032-29-7
1-Propanol	Propan-1-ol	<i>n</i> -Propyl alcohol; <i>n</i> -propanol; ethyl carbinol	$C_3H_8O$	60.10	71-23-8
Aldehydes					
Acetaldehyde	Acetaldehyde	Acetic aldehyde; acetylaldehyde; ethanal; ethyl aldehyde	$C_2H_4O$	44.05	75-07-0
Acrolein	Prop-2-enal	2-Propen-1-one; 2-propenal; prop-2-en-1-al; acraldehyde; acrylaldehyde; acrylic aldehyde; allyl aldehyde; ethylene aldehyde; propenal;	$C_3H_4O$	56.06	107-02-8
		propenaldehyde; propylene aldehyde; <i>trans</i> -acrolein			
Benzaldehyde	Benzaldehyde	Benzoic aldehyde; benzoyl hydride; phenylmethanal	$C_7H_6O$	106.12	100-52-7
Decanal	Decanal	Decyl aldehyde; decaldehyde; decanaldehyde; decylic aldehyde;	$\mathrm{C}_{10}\mathrm{H}_{20}\mathrm{O}$	156.27	112-31-2
		<i>n</i> -decylaldehyde; <i>n</i> -decanal; capric aldehyde			
Formaldehyde	Formaldehyde	Formic aldehyde; methanal; methaldehyde; methyl aldehyde; methylene oxide; oxomethane; oxomethylene; oxymethylene	$CH_2O$	30.03	50-00-0
Heptanal	Heptanal	n-Heptaldehyde; $n$ -heptanaldehyde; $n$ -heptyl aldehyde; $n$ -heptanal; enanthic aldehyde	$C_7H_{14}O$	114.19	111-71-7
Hexanal	Hexanal	1-Hexanal; hexaldehyde; hexoic aldehyde; <i>n</i> -hexanal; <i>n</i> -hexyl aldehyde; caproic aldehyde	$C_6H_{12}O$	100.16	66-25-1
Nonanal	Nonanal	1-Nonaldehyde; 1-nonanal; 1-nonyl aldehyde; <i>n</i> -nonyl aldehyde; nonanoic aldehyde; nonoic aldehyde	$C_9H_{18}O$	142.24	124-19-6
Octanal	Octanal	1-Octaldehyde; 1-octanal; 1-octylaldehyde; n-octaldehyde; n-octanal; n-octyl aldehyde; octanoic aldehyde; caprylic aldehyde	$C_8H_{16}O$	128.21	124-13-0
Phenylacetaldehyde	2-Phenylacetaldehyde	1-Oxo-2-phenyl ethane; alpha-tolualdehyde; alpha-toluic aldehyde; benzeneacetaldehyde; benzyl carboxaldehyde; phenyl acetic aldehyde; phenylethanal	$C_8H_8O$	120.15	122-78-1

Common name	IUPAC-name	Common name IUPAC-name Synonyms (selected)	Chemical	Molecular	CAS-
or name used in			formula	weight	number
document					
Hydrocarbons					
Benzene	Benzene	Benzol; cyclohexatriene; phenyl hydride	$C_6H_6$	78.11	71-43-2
Ethylbenzene	Ethylbenzene	Ethylbenzol; phenylethane	$C_8H_{10}$	106.17	100-41-4
l-Heptene	Hept-1-ene	<i>n</i> -Heptene; <i>n</i> -hept-1-ene	$C_7H_{14}$	98.19	592-76-7
Toluene	Toluene	Methyl benzene; methylbenzol; phenyl methane; toluol	$\mathrm{C_7H_8}$	92.14	108-88-3
1-Methyl-4-	<i>p</i> -Cymene	1-Methyl-4-isopropylbenzene; 1-methyl-4-(methylethyl)-benzene;	$C_{10}H_{14}$	134.22	9-87-6
methylethyl benzene		<pre>para-cymene; 4-isopropyltoluene; p-methyl cumene; 4-methyl isopropylbenzene</pre>			
2-Methyl-1,3-	2-Methylbuta-1,3-diene	2-Methylbutadiene; beta-methylbivinyl; isopentadiene; isoprene	$C_5H_8$	68.12	78-79-5
butadiene					
1-Nonene	Non-1-ene		$C_9H_{18}$	126.24	124-11-8
1,3-Octadiene	Octa-1,3-diene		$C_8H_{14}$	110.20	1002-33-1
1-Octene	Oct-1-ene		$\mathrm{C_8H_{16}}$	112.21	111-66-0
Styrene	Styrene	Ethenylbenzene; phenylethene; phenylethylene; styrol; vinyl benzene; vinylbenzol	$C_8H_8$	104.15	100-42-5
Xylenes	o-, m-, p-Xylene	Dimethylbenzenes; xylols; methyltoluenes	$C_8H_{10}$	106.17	1330-20-7
Acids			1 xylene	1 xylene	
Acetic acid	Acetic acid	Ethylic acid; methanecarboxylic acid	$C_2H_4O_2$	60.05	64-19-7
Octanoic acid	Octanoic acid	1-Heptanecarboxylic acid; n-octanoic acid; n-octylic acid	$C_8H_{16}O_2$	144.21	124-07-2
Elhers					
Anisole	Anisole	Methoxybenzene; methyl phenyl ether; phenyl methyl ether	$C_7H_8O$	108.14	100-06-3
1,3-Dimethoxy- benzene	1,3-Dimethoxybenzene	<i>m</i> -Dimethoxybenzene; dimethylresorcinol	$C_8H_{10}O_2$	138.17	151-10-0
2,5-Dimethylfuran	2,5-Dimethylfuran		$C_6H_8O$	96.13	625-86-5
1-Methoxy-3- methvlbenzene	1-Methoxy-3- methvlbenzene	3-Methylanisole; <i>m</i> -methoxytoluene; <i>m</i> -methylanisole; 3-cresol methyl ether: 3-methoxytoluene; 3-methyl-1-methoxybenzene; <i>m</i> -cresol methyl	$C_8H_{10}O$	122.17	100-84-5

 $\infty$ 

Common name	IUPAC-name	Svnonvms (selected)	Chemical	Molecular	CAS-
or name used in			formula	weight	number
document		مفهمت معافر الماعة سامهما والمعاصر المعافران			
		enter; <i>m</i> -cresyt memyt enter; memyt <i>m</i> -totyt enter			
1-Methoxy-3-	1-Methoxy-3-	4-Methoxy-2-methylbutane; isopentyl methyl ether;	$C_6H_{14}O$	102.17	626-91-5
methylbutane	methylbutane	methyl isopentyl ether			
2-Methylfuran	2-Methylfuran	5-Methylfuran; alpha-methylfuran; methyl furan	$C_5H_6O$	82.10	534-22-5
3-Methylfuran	3-Methylfuran		$C_5H_6O$	82.10	930-27-8
2,3,5-Trimethylfuran	2,3,5-Trimethylfuran		$\mathrm{C_7H_{10}O}$	110.15	10504-04-8
Esters					
Ethyl acetate	Ethyl acetate	Acetoxyethane; ethyl acetic ester	$C_4H_8O_2$	88.11	141-78-6
Ethyl-2 methyl	Ethyl 2-methyl	2-Methylpropanoic acid ethyl ester; ethyl isobutyrate	$C_6H_{12}O_2$	116.16	97-62-1
propionate	propanoate				
Ethyl propionate	Ethyl propanoate	Ethyl <i>n</i> -propanoate; propanoic acid ethyl ester	$C_5H_{10}O_2$	102.13	105-37-3
Methyl acetate	Methyl acetate	Acetic acid methyl ester; methyl acetic ester; methyl ester acetic acid; methyl ethanoate	$C_3H_6O_2$	74.08	79-20-9
3-Methyl-1-butyl	3-Methylbutyl acetate	3-Methyl-1-butanol acetate; acetic acid 3-methyl butyl ester; isoamyl	$\mathbf{C}_{7}\mathbf{H}_{14}\mathbf{O}_{2}$	130.19	123-92-2
acetate		enanoate; isopentyl ester acetic acid; isopentyl acetate; isoamyl acetate			
Methyl-2- methylpropionate	Methyl 2- methylpropanoate	Methyl isobutyrate; 2-methylpropanoic acid methyl ester; methyl 2.2-dimethylacetate	$C_5H_{10}O_2$	102.13	547-63-7
Propyl acetate	Propyl acetate	1-Propyl acetate; <i>n</i> -propyl acetate; <i>n</i> -propyl ethanoate	$C_5H_{10}O_2$	102.13	109-60-4

ase	
ã,	
nt	
ē	
н	
Ę	
0	
·Ħ	
2	
5	
E .	
Ŧ	
Ц	
•	
n	
2	
Ξ	
Ц	
5	
ŏ	
B	
٠Ħ	
G	
5	<i></i>
ğ	-
р,	S
-	-
ă	~ '
a	5
ed as MVOCs of fungi and bacteria common in the enviro	ighlighted (46, 47, 151).
$\overline{o}$	ю.
Ξ	4
Ľ	Ċ
ц.	g
0	ъ
S	È
$\bigcirc$	ы
õ	Ξ
$\leq$	÷
$\leq$	.2
2	Ч
s MVOCs of fungi	Ð
ä	E
ŏ	E
ť	.¤
Ö	H
ġ.	ы
<u>e</u>	٠Ē
$\sim$	ŝ
<u>l</u>	ves
ntly	nves
ently	inves
uently	er inves
quently	ner inves
requently	ther inves
frequently	urther inves
ls frequently	further inves
ids frequently	r further inves
ands frequently reported as MVOCs of fungi and bacteria common in the environment ba	for further investigation are highlighted
ounds frequently	l for further inves
pounds frequently	ed for further inves
npounds frequently	ted for further inves
ompounds frequently	ected for further inves
compounds frequently	lected for further inves
e compounds frequently	selected for further inves
he compounds frequently	s selected for further inves
the compounds frequently	es selected for further investigation are highlighted (46, 47, 151).
of the compounds frequently	ces selected for further inves
of the compounds frequently	inces selected for further inves
on of the compounds frequently	tances selected for further inves
ion of the compounds frequently	ostances selected for further inves
ation of the compounds frequently	ubstances selected for further inves
cation of the compounds frequently	substances selected for further inves
fication of the compounds frequently	5 substances selected for further inves
tification of the compounds frequently	15 substances selected for further inves
intification of the compounds frequently	a 15 substances selected for further inves
lentification of the compounds frequently	he 15 substances selected for further inves
identification of the compounds frequently	The 15 substances selected for further inves
Il identification of the compounds frequently	. The 15 substances selected for further inves
cal identification of the compounds frequently	s. The 15 substances selected for further investives
ical identification of the compounds frequently	ies. The 15 substances selected for further inves
nical identification of the compou	idies. The 15 substances selected for further investigation of the selected for further investigation of the selected for the
nical identification of the compou	tudies. The 15 substances selected for further inves
nical identification of the compou	studies. The 15 substances selected for further inves
nical identification of the compou	v studies. The 15 substances selected for further inves
nical identification of the compou	ory studies. The 15 substances selected for further investigation of the selected for the s
nical identification of the compou	atory studies. The 15 substances selected for further investigation of the selected for the
nical identification of the compou	ratory studies. The 15 substances selected for further inves
nical identification of the compou	oratory studies. The 15 substances selected for further inves
<b>Fable 3.</b> Chemical identification of the compounds frequently	uboratory studies. The 15 substances selected for further inves

Common nameIUPAC-nameSynonyms (seleor name used inAcetoneSynonyms (seleKetonestKetoneSynonyms (seleocumentKetone2-Propanone; pketoneButan-2-oneEthyl methyl ke2-ButanoneButan-2-oneEthyl methyl ke2-HeptanoneRetoneSetocyclopenta2-HeptanoneHeptan-2-oneButyl acetone; n2-HeptanoneHeptan-2-oneButyl acetone; n3-Hydroxy-2-3-Hydroxybutan-2-onen=Butyl methyl hethyl hethyl hethyl butan-2-one3-Hydroxy-2-3-Methylbutan-2-onen=Butyl methyl hethyl hethyl hethyl hethyl butanone3-Methyl-2-butanone3-Methylbentan-2-oneSec-Butyl methyl sec-butyl hethyl hethyl butanone3-Methyl-2-butanone3-Methylbentan-2-oneSec-Butyl methyl seco-butyl seco-butyl secone; n3-Methyl-2-butanone3-Methylbentan-2-oneSec-Butyl nethyl secone; n3-Methyl-2-butanone3-Methylbentan-2-oneSec-Butyl nethyl secone; n3-Methyl-2-butanone3-Methylbentan-2-oneSec-Butyl secone; n3-Methyl-2-butanone3-Methylbentan-2-oneSec-Butyl secone; n3-Methyl-3-hexanone3-Methylbentan-2-oneSec-Butyl secone; n3-Methyl-3-hexanone3-Methylbentan-2-oneSec-Butyl secone; n3-Methyl-3-hexanone4-Methylbentan-2-oneSec-Butyl secone; n3-Methyl-3-hexanone1-Methylbentan-2-oneSec-Butyl secone; n3-OtanoneNonan-2-oneNonan-2-oneMethyl secone; n3-Detanone2-Detanone2-Detanone; n				
d in Acetone Acetone Butan-2-one Cyclopentanone e Heptan-2-one Heptan-2-one 3-Hydroxybutan-2-one butanone 3-Methylbutan-2-one butanone 3-Methylbutan-2-one e Nonan-2-one hexanone 4-Methylbutan-2-one e Pentan-2-one ctan-3-one e Pentan-3-one e Pentan-2-one butanone a d-Methylbutan-2-one calactone 5-Hexyloxolan-2-one	Synonyms (selected)	Chemical	Molecular	CAS-
AcetoneAcetoneButan-2-onenoneCyclopentanoneeHeptan-2-onebutanone3-Hydroxybutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone0-ctan-2-onebutanone0-ctan-3-oneePentan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebutanone1-Methylbutan-2-onebut		formula	weight	number
AcetoneAcetoneButan-2-oneoneCyclopentanoneeHeptan-2-onebutanone3-Hydroxybutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbentan-2-onebutanone3-Methylbentan-2-onebutanone3-Methylbentan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-3-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-onebutanone0-ctan-2-on				
AcetoneButan-2-oneboneButan-2-onecCyclopentanoneeHeptan-2-onebutanone3-Hydroxybutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone0-Ctan-3-onecOctan-3-oneePentan-2-onebutanone1-Methylbertan-3-onebutanone1-Methylbertan-3-onebutanone1-Methylbertan-3-onebutanone1-Methylbertan-3-onebutanone1-Methylbertan-3-onebutanone1-Methylbertan-3-onebutanone1-Methylbertan-2-onebutanone1-Methylbertan-3-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-Methan-2-onebutanone1-M				
Butan-2-oneIoneCyclopentanoneeHeptan-2-onebHeptan-2-onebHetan-2-oneb-3-Hydroxybutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onehexanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone4-Methylbutan-2-onecOctan-3-onecPentan-2-onecPentan-2-onecOctan-3-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPentan-2-onecPe	ropanone; propanone; dimethyl ketone; beta-ketopropane; me propane	$C_3H_6O$	58.08	67-64-1
noneCyclopentanoneeHeptan-2-oneeHexan-2-one2-3-Hydroxybutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone3-Methylbutan-2-onebutanone4-Methylbutan-2-onehexanone4-Methylbutan-2-onecNonan-2-onecOctan-3-oneePentano-2-oneePentan-2-oneePentan-2-oneeNonan-2-oneeS-Hethylbutan-2-oneePentan-2-oneeS-IndeePentan-2-oneePentan-2-oneePentan-2-oneeS-IndeeS-IndeePentan-2-oneeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-IndeeS-In	Ethyl methyl ketone; methyl acetone; methyl ethyl ketone; oxobutane	$C_4H_8O$	72.11	78-93-3
Heptan-2-one         Hexan-2-one         3-Hydroxybutan-2-one         3-Methylbutan-2-one         3-Methylbutan-2-one         3-Methylbutan-2-one         Nonan-2-one         Nonan-2-one         Octan-2-one         Pentan-2-one         Pentan-2-one         Dotan-2-one         Nonan-2-one         Nonan-2-one         Nonan-2-one         Nonan-2-one         Nonan-2-one         Nonan-2-one         Nonan-2-one         None         Nonan-2-one         None         S-Hexyloxolan-2-one	Ketocyclopentane	$C_5H_8O$	84.12	120-92-3
Hexan-2-one3-Hydroxybutan-2-one3-Methylbutan-2-one3-Methylpentan-2-one•4-Methylhexan-3-oneNonan-2-oneOctan-2-oneOctan-2-onePentan-2-onePentan-3-onePentan-2-oneVindecan-2-oneUndecan-2-one5-Hexyloxolan-2-one	yl acetone; methyl $n$ -amyl ketone; $n$ -amyl methyl ketone;	$C_7 H_{14} O$	114.19	110-43-0
Hexan-2-one3-Hydroxybutan-2-one3-Methylbutan-2-one3-Methylpentan-2-one•4-Methylhexan-3-oneNonan-2-oneOctan-2-oneOctan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePenta	methyl pentyl ketone			
3-Hydroxybutan-2-one3-Methylbutan-2-one3-Methylpentan-2-one4-Methylhexan-3-oneNonan-2-oneOctan-2-oneOctan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-onePentan-2-one<	<i>n</i> -Butyl methyl ketone; methyl <i>n</i> -butyl ketone; propylacetone	$C_6H_{12}O$	100.16	591-78-6
3-Methylbutan-2-one3-Methylbutan-2-one4-Methylhexan-3-oneNonan-2-oneNonan-2-oneOctan-2-oneOctan-3-onePentan-2-onePentan-2-onePentan-2-oneVindecan-2-one5-Hexyloxolan-2-one	2,3-Butanolone; 2-butanol-3-one; gamma-hydroxy-beta-oxobutane	$C_4H_8O_2$	88.11	513-86-0
3-Methylbutan-2-one3-Methylpentan-2-one4-Methylhexan-3-oneNonan-2-oneOctan-2-oneOctan-3-onePentan-2-onePentan-2-onePentan-2-onePentan-2-oneS-Hexyloxolan-2-one5-Hexyloxolan-2-one				
pentanone3-Methylpentan-2-onehexanone4-Methylhexan-3-onehexanone4-Methylhexan-3-onehexanoneNonan-2-oneOctan-2-oneOctan-3-onehexanonePentan-2-onehexanoneNonan-3-onehexanoneNonan-2-onehexanonePentan-2-onehexanoneNonan-3-onehexanoneS-Hexyloxolan-2-onehexanoneS-Hexyloxolan-2-one	Isopropyl methyl ketone; methyl isopropyl ketone	$C_5H_{10}O$	86.13	563-80-4
hexanone4-Methylhexan-3-oneeNonan-2-oneOctan-2-oneOctan-3-oneePentan-2-oneePentan-2-oneePentan-2-oneneUndecan-2-onealactone5-Hexyloxolan-2-one	Sec-Butyl methyl ketone; methyl 1-methylpropyl ketone;	$C_6H_{12}O$	100.16	565-61-7
hexanone4-Methylhexan-3-oneeNonan-2-oneOctan-2-oneOctan-3-oneePentan-3-oneePentan-2-oneePentan-2-oneneUndecan-2-onealactone5-Hexyloxolan-2-one	methyl sec-butyl ketone			
e Nonan-2-one Octan-2-one Octan-3-one Pentan-2-one Pentan-3-one ne Undecan-2-one alactone 5-Hexyloxolan-2-one	Ethyl isobutyl ketone	$\mathbf{C}_7\mathbf{H}_{14}\mathbf{O}$	114.19	17042-16-9
Octan-2-oneOctan-3-onePentan-2-onePentan-3-oneneUndecan-2-onealactone5-Hexyloxolan-2-one	Methyl heptyl ketone; n-heptyl methyl ketone	$\rm C_9H_{18}O$	142.24	821-55-6
Octan-3-oneePentan-2-oneePentan-3-oneneUndecan-2-onealactone5-Hexyloxolan-2-one	yl methyl ketone; methyl <i>n</i> -hexyl ketone; 2-oxooctane	$\rm C_8H_{16}O$	128.21	111-13-7
Pentan-2-one Pentan-3-one Undecan-2-one actone 5-Hexyloxolan-2-one	Ethyl amyl ketone; <i>n</i> -amyl ethyl ketone; ethyl pentyl ketone	$\rm C_8H_{16}O$	128.21	106-68-3
Pentan-3-one Undecan-2-one actone 5-Hexyloxolan-2-one	Ethyl acetone; methyl <i>n</i> -propyl ketone; propyl methyl ketone	$C_5H_{10}O$	86.13	107-87-9
Undecan-2-one actone 5-Hexyloxolan-2-one	Diethyl ketone; dimethyl acetone; ethyl ketone; methacetone	$C_5H_{10}O$	86.13	96-22-0
Decalactone 5-Hexyloxolan-2-one	Methyl nonyl ketone; undecanone	$C_{11}H_{22}O$	170.29	112-12-9
5-Hexyloxolan-2-one				
	2-Decalactone; decanoic acid, gamma-lactone; decanolactone;	$\mathbf{C}_{10}\mathbf{H}_{18}\mathbf{O}_2$	170.25	706-14-9
4-decanolid	4-decanolide; decanolide-1,4; 5-hexyldihydro-2(3H)-furanone;			
gamma- <i>n</i> -he	gamma- <i>n</i> -hexyl-gamma-butyrolactone			

Table 3. Chemical identification of the compounds frequently reported as MVOCs of fungi and bacteria common in the environment based on

Common name	IUPAC-name	Synonyms (selected)	Chemical	Molecular	CAS-
or name used in			formula	weight	number
document					
Terpenoids					
Acoradiene	1,8-Dimethyl-4-prop-1-	(-)-alpha-Acoradiene;	$\mathrm{C}_{15}\mathrm{H}_{24}$	204.35	24048-44-0
	en-2-yl-spiro[4.5]dec-8-	1,8-dimethyl-4-isopropenylspiro[4.5]dec-7-ene;			
	ene	1,8-dimethyl-4-(1-methylethenyl)-spiro[4.5]dec-7-ene,(1R,4S,5S)-; sniro[4 5]dec-7-ene 1_isonronenyl-4 8-dimethyl_(1S,4R_5S)-(2)-			
<b>3-Bisabolene</b>	6-Methyl-2-(4-methyl-1-	(-)-beta-Bisabolene;	$C_{15}H_{24}$	204.35	495-61-4
	cyclohex-3-enyl)-hepta-	1,5-heptadiene, 6-methyl-2-(4-methyl-3-cyclohexen-1-yl)-, (S)-(-)-;			
	1,5-diene	cyclohexene, 1-methyl-4-(5-methyl-1-methylene-4-hexenyl)-, (S)-;			
		(S)-1-methyl-4-(5-methyl-1-methylene-4-hexenyl)-1-cyclohexene			
Cadinene	1,6-Dimethyl-4-propan-	b-Cadinene; [1S-(1alpha,4alpha,4aalpha,6alpha,8alphabeta)]-decahydro-	$\mathrm{C}_{15}\mathrm{H}_{26}$	206.37	29350-73-0
	2-yl-1,2,3,4,4a,5,6,8a-	1,6-dimethyl-4-(1-methylethyl)naphthalene			
	octahydronaphthalene				
Δ3-Carene	3,7,7-Trimethyl-	3-Carene; car-3-ene; delta-3-carene; S-3-carene; isodiprene;	$\mathrm{C_{10}H_{16}}$	136.24	13466-78-9
	bicyclo[4.1.0]hept-3-ene	3,7,7-trimethyl bicyclohep-3-ene;			
		3,7,7-trimethylbicyclo[4.1.0]-3-heptene;			
		3-norcarene, 3,7,7-trimethyl-;			
		4,7,7-trimethyl-3-norcarene			
Camphene	2,2-Dimethyl-3-	(+/-)-Camphene; 2,2-dimethyl-3-methylene-bicyclo[2.2.1]heptane;	$\mathrm{C}_{10}\mathrm{H}_{16}$	136.24	79-92-5
	methylidene-norbornane	2,2-dimethyl-3-methylene norbornane;			
		3,3-dimethyl-2-methylene-norcamphane;			
		3.3-dimethyl-2-methylene norcamphone			

**Table 3.** Chemical identification of the compounds frequently reported as MVOCs of fungi and bacteria common in the environment based on

Common name or name used in document	IUPAC-name	Common name IUPAC-name Synonyms (selected)	Chemical formula	Molecular weight	CAS- number
β-Caryophyllene	4,11,11-Trimethyl-8- methylidene-bicyclo [7.2.0]undec-4-ene	<ul> <li>(-)-beta-Caryophyllene; l-caryophyllene; (-)-<i>trans</i>-caryophyllene;</li> <li>(-)-E-caryophyllene;</li> <li>[1R-(1R*,4E,9S*)]-8-methylene-4,11,11-trimethylbicyclo[7.2.0]-4-undecane;</li> <li>[1R-(1R*,4E,9S*)]-4,11,11-trimethyl-8-methylene-bicyclo[7.2.0]-4-undecene;</li> <li>4,11,11-trimethyl-8-methylene-bicyclo[7.2.0]undec-4-ene;</li> <li>bicyclo[7.2.0]undec-4-ene, 4,11,11-trimethyl-8-methylene-,</li> <li>(E)-(1R,9S)-(-)-</li> </ul>	C <sub>15</sub> H <sub>24</sub>	204.35	87-44-5
β-Chamigrene	1,1,9-Trimethyl-5- methylidene- spiro[5.5]undec-9-ene	Spiro[5.5]undec-2-ene, 3,7,7-trimethyl-11-methylene, (6R)-; (-)-3,7,7-trimethyl-11-methylenespiro[5.5]undec-2-ene	$C_{15}H_{24}$	204.35	18431-82-8
α-Curcumene	2-Methyl-6-(4-methyl- phenyl)-hept-2-ene	a-Curcumene; 2-Heptene, 2-methyl-6- <i>p</i> -tolyl-; 1-(1,5-dimethyl-4-hexenyl)-4-methylbenzene; 1-methyl-4-(6-methylhept-5-en-2-yl)benzene	$C_{15}H_{22}$	202.34	644-30-4
β-Elemene	1-Ethenyl-1-methyl-2,4- diprop-1-en-2-yl- cyclohexane	Cyclohexane, 1-ethenyl-1-methyl-2,4-bis(1-methylethenyl)-, (1alpha,2beta,4beta)-; 2,4-diisopropenyl-1-methyl-1-vinylcyclohexane, stereoisomer	$C_{15}H_{24}$	204.35	33880-83-0
α-Famesene	(3E,6E)-3,7,11- Trimethyldodeca- 1,3,6,10-tetraene	<i>trans</i> -alpha-Farnesene; (3E, 6E)-alpha-farnesene; 2,6,10-trimethyl-2,6,9,11-dodecatetraene; 3,7,11-trimethyl-1,3,6,10-dodecatetraene	$C_{15}H_{24}$	204.35	502-61-4
β-Farnesene	(6E)-7,11-Dimethyl-3- methylidene-dodeca- 1,6,10-triene	<i>trans</i> -beta-Farnesene; E-beta-farnesene; 7,11-dimethyl-3-methylene-1,6,10-dodecatriene	$C_{15}H_{24}$	204.35	18794-84-8

<b>Table 3.</b> Chemical in laboratory studies. T	dentification of the comported in 15 substances selected in the 15 substances selected in the 15 substances selected in the se	<b>Table 3.</b> Chemical identification of the compounds frequently reported as MVOCs of fungi and bacteria common in the environment based on laboratory studies. The 15 substances selected for further investigation are highlighted (46, 47, 151).	the environ	ment based	on
Common name	IUPAC-name	Synonyms (selected)	Chemical	Molecular	CAS-
or name used in document			formula	weight	number
Geosmin	4,8a-Dimethyldecalin-	1- $\alpha$ ,10- $\beta$ -Dimethyl-9 $\alpha$ -decalol, 2,6-dimethyl bicyclo[4.4.0]decan-1-ol;	$C_{12}H_{22}O$	182.31	23333-91-7
	4a-ol	octahydro-4,8a-dimethyl-4a(2H)-naphthalenol; trons-1 10-dimethyl-trans-9-decalol			and 19700-21-1
α-Gurjunene	No IUPAC name	(-)-α-Gurjunene;	C <sub>15</sub> H <sub>24</sub>	204.35	489-40-7
		1H-cycloprop[e]azulene, 1a,2,3,4,4a,5,6,7b-octahydro-1,1,4,7-			
		tetramethyl-, (1aR, 4R, 4aR, 7bS)-;			
		1H-cycloprop[e]azulene, 1a.beta.,2,3,4,4a.alpha.,5,6,7b.betaoctahydro-			
		1,1,4.beta7-tetramethyl-			
Limonene	1-Methyl-4-prop-1-en-2-	DL-Limonene; eulimen; DL-p-mentha-1, 8-diene; acintene DP dipentene;	$C_{10}H_{16}$	136.24	138-86-3
	yl-cyclohexene	cajeputene; ciene; cinene; cyclil decene; nesol; terpodiene;			
		1-methyl-4-(1-methylethenyl)cyclohexene; 4-(1-methylethenyl)-1-			
		methyl-cyclohexene; 4-isopropenyl-1-methyl-1-cyclohexene;			
		4-isopropenyl-1-methyl-cyclohexene;			
		methyl-4-(1-methylethenyl)cyclohexene;			
		methyl-4-isopropenyl-1-cyclohexene;			
		methyl-4-isopropenylcyclohexene			
Longifolene	No IUPAC name	Kuromatsuene; junipene; d-longifolene; (+)-longifolene;	$\mathrm{C}_{15}\mathrm{H}_{24}$	204.35	475-20-7
		(+)-longofolene;			
		[1S-(1alpha,3abeta,4alpha,8abeta)]-decahydro-4,8,8-trimethyl-9-			
		methylene-1,4-methanoazulene;			
		decahydro-4,8,8-trimethyl-9-methylene-1,4-methanoazulene			
2-Methylisoborneol	1,2,7,7-Tetramethyl-	Bicyclo [2.2.1] heptan-2-ol; 1,2,7,7-tetramethyl-, (1R, 2R, 4R)-rel-;	$C_{11}H_{20}O$	168.28	2371-42-8
	norbornan-2-ol	bicyclo [2.2.1] heptan-2-ol, 1,2,7,7, tetramethyl-, exo-;			
		2-norbornanol, 1,2,7,7-tetramethyl-, exo-;			
		2-endo-methyl-2-exo-bornanol			
		exo-1,2,7,7-tetramethylbicyclo[2.2.1]heptan-2-ol			

Common name	<b>IUPAC-name</b>	Synonyms (selected)	Chemical	Molecular	CAS-
or name used in document		· · · · · · · · · · · · · · · · · · ·	formula	weight	number
β-Phellandrene	3-Methylidene-6-propan-	1(7)-2- <i>p</i> -Menthadiene;	$\mathrm{C}_{10}\mathrm{H}_{16}$	136.24	555-10-2
	2-yl-cyclohexene	3-methylene-6-(1-methylethyl)- cyclohexene			
α-Pinene	4,7,7-Trimethyl-	2-Pinene; pin-2(3)-ene; acintene A; cyclic dexadiene;	$\mathrm{C}_{10}\mathrm{H}_{16}$	136.24	80-56-8
	bicyclo[3.1.1]hept-3-ene	2,6,6-trimethylbicyclo[3.1.1]hept-2-ene			
β-Pinene	7,7-dimethyl-4-	2(10)-Pinene; nopinene; pseudopinene; terebenthene;	$\mathrm{C}_{10}\mathrm{H}_{16}$	136.24	127-91-3
	methylidene-	bicyclo[3.1.1]heptane, 6,6-dimethyl-2-methylene-			
	bicyclo[3.1.1]heptane				
Thujopsene	No IUPAC name	(-)-Thujopsene; sesquichamene; widdrene;	$\mathrm{C}_{15}\mathrm{H}_{24}$	204.35	470-40-6
		cyclopropa[d]naphthalene, 1,1a,4,4a,5,6,7,8-octahydro-2,4a,8,8-			
		tetramethyl-, (1aS, 4aS, 8aS)-			
Trichodiene	1,4-Dimethyl-4-(1-	Cyclohexene, 1,4-dimethyl-4-[(1S)-1-methyl-2-methylenecyclopentyl]-;	$\mathrm{C}_{15}\mathrm{H}_{24}$	204.35	28624-60-4
	methyl-2-methylidene-	cyclohexene, 1,4-dimethyl-4-(1-metyl-2-methylenecyclopentyl)-,			
	cyclopenty])-	$[S-(R^*, R^*)]$ -			
	cyclohexene				
Sulphur and nitrogen compounds	compounds				
Dimethyl disulphide	Methyldisulfanyl-	Methyl disulphide; 2,3-dithiabutane; (methyldithio)methane	$C_2H_6S_2$	94.19	624-92-0
	methane				
Dimethyltrisulphide	Methylsulfanyldisulfanyl	Methyl trisulphide; trisulphide, dimethyl; 2,3,4-trithiapentane	$C_2H_6S_3$	126.27	3658-80-8
2-Isopropyl-3-	2-Methoxy-3-propan-2-	2-Isopropyl-3,5 (or 6)-methoxypyrazine; 2-methoxy-3,5(6)-isopropyl	$C_8H_1N_0$	152.20	25773-40-4
methoxy-pyrazine	yl-pyrazine	pyrazine; 2-methoxy-3-isopropyl-pyrazine;	1		
2_Methovy nyrazine	2 Mathownmarina	Mathematica		110 10	2110 70 0

# 3. Physical and chemical properties

Some physical and chemical properties of MVOCs (from Table 3) are listed in Table 4. An indication of the volatility of a compound can be deduced from the number of carbon atoms, the molecular weight, boiling point, and the vapour pressure (198). The interpretation of vapour pressures of the MVOCs presented in Table 4 is as follows: 0.00001-0.001 kPa = moderately volatile, 0.001-0.1 kPA = volatile and >0.1 kPa = very volatile compound (159).

Compound	Boiling point (°C) <sup>a</sup> at 101.3 kPa	Vapour pressure (kPa) at 25 °C	Octanol:wat coefficien Experimental	
Alcohols				
1-Butanol	117.7	0.587-0.73 °	0.88	
4-Decanol	210.5	0.0049		3.71
Ethanol	78.5	5.9 °	-0.32	
2-Ethyl-1-hexanol	183.5	0.007 ° 0.048 °		2.73
2-Heptanol	159.2	0.164	2.31	
1-Hexanol	157.1-157.5	0.124	2.03	
2-Methyl-1-propanol	108	1.33	0.65 0.83	
2-Methyl-1-butanol	128	0.416	1.29	
3-Methyl-1-butanol	130.5	0.316	1.16	
3-Methyl-2-butanol	111.5	1.22	1.28	
1-Octanol	194.5-195	0.0106	3.00	
3-Octanol	169.0	0.068		2.73
1-Octen-3-ol	180	0.071		2.60
2-Octen-1-ol	$195.8 \pm 8.0$	0.014		2.59
1-Pentanol	137.8	0.218 °	1.42 1.48 1.51	
2-Pentanol	119.0-119.3	0.815	1.19	
1-Propanol	97.2-97.8	1.9 - 2 °	0.25	
1			0.34	
Aldehydes				
Acetaldehyde	20.2	100 <sup>c</sup>	-0.34	
Acrolein	52.5	28.5 °		-0.01
Benzaldehyde	178-179	0.133 <sup>d</sup>		1.48
Decanal	208	0.028		3.76
Formaldehyde	-19- (-)19.5	519 462	0.35	
Heptanal	152.8	0.469		
Hexanal	131	1.507		1.78
Nonanal	191	0.049		3.27
Octanal	163.4	0.28		2.78
Phenylacetaldehyde	200	0.049		

**Table 4.** Some physical and chemical properties of compounds reported as MVOCs. Data is partly based on calculated/predicted coefficients (41, 46, 47, 88, 89, 91, 151, 196). Substances selected for closer examination are highlighted.

Compound	Boiling point (°C) <sup>a</sup> at 101.3 kPa	Vapour pressure (kPa) at 25°C	Octanol:wat coefficien Experimental	
Hydrocarbons				
Benzene	80	12.7	1.18-1.9 2.13 2.15	
Ethylbenzene	136.2	1.28	3.15	
1-Heptene	94	7.62	0.110	3.99
Toluene	110.6	2.93 °	2.11-2.80	
1-Methyl-4-	177.10	0.20		4.10
methylethyl benzene				
2-Methyl-1,3-butadiene	34.067	73.33		2.42
1-Nonene	146.9	0.720		5.15
1,3-Octadiene	130-131	1.79		
1-Octene	121.2	2.32		4.57
Styrene	145.2	0.81	2.95	
Xylenes	129-150 <sup>b</sup>	0.8-0.867 °	3.12-3.20	
Acids				
Acetic acid	117.9	1.52 °	-0.31	
Octanoic acid	237; 239.7	0.133 °	0.63	
Ethers	201,2001	0.122	0.02	
Anisole	155.5	0.472		2.11
1,3-Dimethoxybenzene	217.5	0.026		2.21
2,5-Dimethylfuran	$93.1 \pm 9.0$	7.61		2.21
1-Methoxy-	177	0.24		2.66
3-methylbenzene	177	0.24		2.00
1-Methoxy-	90	11.06		1.96
3-methylbutane	50	11.00		1.90
2-Methylfuran	65	23.48		1.85
3-Methylfuran	65-66	21.46		1.91
2,3,5-Trimethylfuran	121-122	1.92		1.71
Esters	121-122	1.72		
Ethyl acetate	76.5-77.5	9.73 °	0.66 0.73	
Ethyl-2-methyl propionate	110.1	2.88		1.77
Ethyl propionate	99.1	5.0	1.21	
Methyl acetate	56.9	23.1 °	0.18	
		21.7 °		
3-Methyl-1-butyl acetate	142.5	0.75	2.26	
Methyl-2-methyl- propionate	93-95	6.72		1.28
Propyl acetate	101.6	4.67	1.39 1.60	
Ketones				
Acetone	56.2	24-24.7 °	-0.24	
2-Butanone	79.6	10.33 °	0.26	
0.1	120 (	1.50	0.29	0.04
Cyclopentanone	130.6	1.52	2.02	0.24
2-Heptanone	150.6; 151.5	0.213 0.28	2.03	

Compound	Boiling point (°C) <sup>a</sup> at 101.3 kPa	Vapour pressure (kPa) at 25°C	Octanol:wat coefficien Experimental	
2-Hexanone	126-128	1.47 ° 0.36 °	1.38	
3-Hydroxy-2-butanone	148	0.256		-0.36
3-Methyl-2-butanone	93	8.6 °		0.84
3-Methyl-2-pentanone	117.3-117.5	2.42		1.16
4-Methyl-3-hexanone	135-138	1.075		1.66
2-Nonanone	194	0.086		3.14
2-Octanone	173	0.23		2.37
3-Octanone	157-162	0.267		2.22
2-Pentanone	102	3.59 °	0.91	
3-Pentanone	102	4.7	0.99	
2-Undecanone	231.5	0.013		4.09
Lactones				
γ-Decalactone	$266.7 \pm 8.0$	0.00113		
Terpenoids				
Acoradiene	$273.1 \pm 15.0$	0.0013		6.99
β-Bisabolene	$275.4 \pm 15.0$	0.0011		7.12
Cadinene	120 <sup>f</sup>	5.33 <sup>g</sup>		6.19
$\Delta$ 3-Carene	167-170	0.248		4.61
Camphene	158.5	0.333		4.22
β-Caryophyllene	$268.4 \pm 10.0$	0.0017		6.30
β-Chamigrene	$273.2 \pm 15.0$	0.0013		7.02
α-Curcumene	$276.3 \pm 15.0$	0.0011		6.29
β-Elemene	$252.1 \pm 15.0$	0.0042		7.04
α-Farnesene	$279.6 \pm 15.0$	0.00090		7.10
β-Farnesene	$272.5 \pm 15.0$	0.0013		7.17
Geosmin	$252.4 \pm 8.0$	0.00041		3.57
α-Gurjunene	$263.9 \pm 7.0$	0.0022		6.18
Limonene	170	0.280 °		4.57
Longifolene	250-255	0.0042		5.48
2-Methylisoborneol	$208.7\pm8.0$	0.0065		3.31
β-Phellandrene	171.5	0.210		4.70
α-Pinene	155	0.47		4.83
β-Pinene	166.0	0.32		4.35
Thujopsene	$256.5\pm7.0$	0.0033		6.12
Trichodiene	$256.7 \pm 15.0$	0.0033		
Sulphur and nitrogen c	ompounds			
Dimethyl disulphide	109.8	3.83	1.77	
Dimethyl trisulphide	$183.1 \pm 23.0$	0.142		1.87
2-Isopropyl-3- methoxy-pyrazine	$210.8 \pm 30.0$	0.036		2.37
2-Methoxy pyrazine	$153.6 \pm 0.0$	0.56		0.73

Table 4. Cont.

 $^{a}\pm$  indicates that the value was obtained with an extrapolation model providing a range

<sup>b</sup> variable depending on isomer composition

 $^{\circ}$  92.3 °C, extremely low at room temperature  $^{f}$  9 mm Hg (1.2 kPa)  $^{g}$  180 °C

<sup>° 20 °</sup>C <sup>d</sup> 26 °C

## 4. Occurrence

As MVOCs are a result of microbial metabolism, factors that control microbial growth also influence MVOC production including: 1) microbial species and strains (this is the basis for approaches for species identification based on MVOC profile); 2) substrates and nutrients (e.g. lack of certain nutrients leads to terpene emissions, and presence of certain amino acids in the substrate results in sulphur and nitrogen compounds); 3) moisture conditions (water activity, relative humidity, which affects growth and thereby MVOC production); 4) ergosterol content of the growth substrate; 5) ambient VOCs in the air or in the growth substrate, and 6) temperature (27-30, 33, 36-39, 49, 50, 61, 69, 75, 87, 95, 116-118, 123, 127, 128, 169, 175, 182-184, 195, 200-202, 217, 222, 223, 225, 235). On the other hand, sporulation intensity (the concentration of culturable spores) has not been shown to affect the MVOC production (37, 38, 222). Contradictory results on the influences of some factors (e.g. production of metabolic CO<sub>2</sub>, oxygen concentration and growth phase / age of the colony) on MVOC profiles have also been reported. One explanation for this may be defects in sampling techniques and differences in study design (117). Though MVOCs mainly originate from fresh and metabolically active microbial contamination, certain MVOCs have been suggested to reflect emissions from an aged, perhaps even previous, microbial contamination because of their high absorption affinity to building materials (214). MVOC levels indoors are a balance between production rates, absorption to and desorption from building materials and furniture, and ventilation.

In minor scale, MVOCs were initially analysed in order to detect undesirable or spoilage processes during the storage or processing of foodstuffs (36, 38, 53, 54, 131, 147, 226). More recently, MVOC analysis was applied to recognise indoor odour sources and hidden microbial growth behind interior surfaces without opening building structures (20, 149, 197, 210, 211, 224), because it was assumed that, being gases, MVOCs may enter indoor air through water vapour barriers more easily than spores (197, 209).

As already mentioned, MVOCs possess an unspecific nature in field settings, because the same compounds may also originate from other sources, like building materials, human metabolism, cleaning, traffic, foodstuffs, smoking, vegetation, etc. (83, 171, 183, 198). In reactions between oxidants like ozone and unsaturated hydrocarbons (isoprene/terpenes), hydrogen peroxide, methacrolein, and methyl-vinyl ketone (179), all of which are known irritants, may be formed (208, 228, 229). 3-Methylfuran is suspected to be another oxidation product of isoprene (83).

Thus, it is not possible to conclude whether a compound derives from microbial metabolism or from the emission of substrates or environmental pollutants. This hampers the specificity and interpretation of MVOC analyses and limits the use of MVOCs for identifying contaminated areas in a building. For example, terpenes are commonly emitted from wood products, but are also reported as MVOCs in laboratory studies. Likewise, 2-ethyl-1-hexanol, which has been denoted an MVOC, is a degradation product of phthalates in polyvinyl chloride (PVC)

floorings under humid and alkaline conditions (76). Korpi et al reported that moist building materials, also when not microbiologically contaminated, emitted compounds that, when deriving from microbial metabolism, would be called MVOCs (116). Kuske et al suggested that MVOCs evaporating from humid materials could actually be used to indicate moisture problems anticipating fungal growth (121). In favourable conditions the microbial germination and growth can occur in one day (167). Still, the concentrations of MVOCs cannot unequivocally be related to mould grade. According to Laussmann *et al*, the sum of eight MVOCs failed to discriminate rooms according to their mould status (130). Schleibinger et al concluded similarly that MVOCs could not be used as predictors for mould damage in indoor environments (184). In a study of 40 dwellings with and 44 dwellings without mould damage, the occurrence of 2-methylfuran and 3-methylfuran correlated with smoking rather than with mould infestation. Even though 2methyl-1-butanol and 1-octen-3-ol were weakly correlated with fungal state, the sensitivity and specificity of these compounds were concluded to be too low to make them useful indicators (185).

# 5. Measurements and analysis of MVOCs

As there are no standards, consensus, or even recommendations regarding sampling and analysis of MVOCs, the methodology presented in the literature varies greatly and comparative data on different methods are scantily available. MVOCs can be collected from ambient air with active or passive sorbent sampling. Several sorbents or their combinations, like activated charcoal (e.g. Anasorb 747), graphitised carbon blacks (e.g. Carbotrap C, Carbopack B), silica gels (e.g. Porasil C), and polymers (e.g. Tenax TA or GR, Anasorb 727, Chromosorb 102, XAD-4) have been used for both sampling techniques in indoor environments (19, 50, 63, 198, 199). In addition, carbonyl compounds have been collected separately with 2,4-dinitrophenylhydrazine (DNPH)-silica Sep-Pak cartridges in some cases (116, 186). Tenax TA has been widely used because of favourable properties regarding recovery, breakthrough, and precision during sampling/analysis (199). On the other hand, activated charcoal enables longer sampling periods and collection of very volatile MVOCs (197).

Nowadays, MVOCs from environmental samples are mainly analysed with high-resolution gas chromatography (GC) and mass spectrometry (MS) and identified according to their mass spectra. Another applicable detector is the flame ionisation detector (FID) (19, 50, 116, 198). The sample preparation depends on the sorbent used; e.g. for Tenax polymers, the sample is led from the adsorbent to the GC column by a thermal desorption cold trap injector (198), whereas for charcoal sorbents (such as Anasorb) desorption with solvent (e.g. methylene chloride) is required before leading the sample into the GC (197). Carbonyl compounds collected in DNPH-silica Sep-Pak cartridges are analysed by high-performance liquid chromatography after extraction with acetonitrile (50, 116, 186).

Recently, a solid-phase microextraction (SPME)/GC-MS technique has been applied to analyse MVOCs qualitatively from microbial cultures or contaminated building materials (69, 207). In SPME, a short fused silica fibre coated with a polymeric organic material (e.g. polyacrylate and polydimethylsiloxane) is used as a secondary phase. Nilsson *et al* (1996) reported comparable results between SPME and Tenax adsorption (160). The SPME method has been applied by several research teams to qualitatively characterise fungal emissions (57, 58, 69, 94, 160, 207).

Attempts have been made to establish evaluation criteria for indoor environments (135). The principal component analysis is one approach for interpreting chromatograms of house dust samples collected from areas with various grade of microbial contamination (224). Also the use of an electronic nose to detect fungal contamination in indoor environments has recently been reviewed. The electronic nose is aimed to recognise the patterns of compounds related to the occurrence of fungi. The authors concluded that at present, despite promising implications, the low MVOC concentrations and presence of interfering substances restrict the use of the electronic nose in indoor settings (121).

## 6. Exposure data

Measurements of MVOCs in field settings have focused on indoor environments, especially buildings with water and microbial damage and/or unspecified indoor air problems. Published data on MVOC measurements in problem buildings are available mainly from Sweden, Germany, and the USA (40, 66, 106, 135, 148, 149, 192, 197, 210, 212-214, 239). The aim has not been to get exposure data but rather to reveal contaminated areas/buildings. The data are, however, far too limited for the evaluation of global or even local exposure to MVOCs. Also, the lack of standardised and validated analytical methods for MVOCs makes comparison between studies difficult (106). Even though the number of determined MVOCs and the analytical method has been the same in different studies, the sum of MVOCs in problem buildings may differ by three orders of magnitude (66, 149, 197, 213, 239).

Table 5 summarises the available data regarding concentrations of MVOCs in problem and normal buildings or areas and outdoor air. The indoor air concentrations have been measured in residences or in non-industrial work sites such as schools. In some of these studies (135, 148, 192), the study design and the presentation of the data have not allowed a satisfactory differentiation between problem and reference buildings or areas. The concentrations of *individual* compounds in problem buildings have varied from a few ng/m<sup>3</sup> to 1 mg/m<sup>3</sup>, and the same compounds have also been identified in reference buildings or areas and even outdoor air. Generally, the maximum reported levels of individual MVOCs

Compound	R	Range of reported MVOC concentration $(\mug/m^3)$	concentration $(\mu g/m^3)$		References
	Problem buildings/ complaint areas	Reference buildings/ non-complaint areas	Building/area with unspecified identification	Outdoor air	
2-Methyl-1-propanol	n.d1.74	0.38	ı	n.d0.08	(197)
3-Methyl-1-butanol	0.175-260	0.07-8.7	n.d110	n.d3.8	(66, 135, 144, 148, 149, 192, 197)
3-Methyl-2-butanol	0.19-1.19	0.16	1	n.d0.43	(197)
2-Pentanol	0.012-1.4	0.18-1.7	n.d0.45	n.d0.63	(66, 135, 144, 192, 197)
3-Octanol	1.3-8.86	n.d.	n.d0.04	n.d0.14	(40, 66, 192, 197)
1-Octen-3-ol	0.08-904	0.04-7	n.d4.8	n.d1.9	(40, 66, 135, 144, 149, 164, 192, 197)
2-Octen-1-ol	1.56-266	0.30-13.1	n.d16.1	n.d6.82	(40, 66, 148, 149, 192, 197)
3-Methylfuran	n.d0.6	0.02-0.1	n.d1.8	n.d0.11	(135, 144, 149, 164, 192, 197)
2-Hexanone	0.50-8.8	0.46-2.9	traces-0.19	n.d0.8	(66, 135, 197)
2-Heptanone	0.24 - 97	0.58 - 1.2	traces-44	n.d1.1	(66, 135, 148, 149, 192, 197)
3-Octanone	0.03-3.02	0.14-3.0	n.d0.41	n.d2	(66, 135, 144, 192, 197)
2-Methylisoborneol	0.41-2.8	0.56	n.d0.02	n.d1.18	(192, 197)
2-Isopropyl-3-methoxy-pyrazine	0.6-9.5	n.d.	n.d0.003	n.d0.34	(192, 197)
Geosmin	0.006-0.55	n.d.	n.d0.05	n.d0.014	(192, 197)
Dimethyl disulphide	0.03 - 0.09	I	n.d0.05	n.d0.01	(135, 144)

Table 5. Reported concentrations of individual MVOCs in buildings and outdoor air.

n.d. = not detected - = not analysed

are 0.1-10  $\mu$ g/m<sup>3</sup> in problem buildings. However, levels of approximately 100  $\mu$ g/m<sup>3</sup> of 2-heptanone (148) and 270  $\mu$ g/m<sup>3</sup> of 3-methyl-1-butanol (148) and 2-octen-1-ol (149) have been reported. The highest individual MVOC concentration (approximately 900  $\mu$ g/m<sup>3</sup>) is reported for 1-octen-3-ol (149).

When it comes to the use of the sum of certain MVOCs - or *total* MVOCs, as they are sometimes denoted – investigators have included different MVOCs. In fact, different research groups have included from 7 to 23 compounds in the sum of total MVOCs. The decision on which MVOCs to include has differed signifycantly even if the number of compounds was the same. Bearing in mind the great variation in the number of compounds, some ranges for total MVOC levels can still be indicated. In problem buildings/areas, total MVOCs have been reported to be 0.05-84  $\mu$ g/m<sup>3</sup>, in reference areas <0.01-30.1  $\mu$ g/m<sup>3</sup>, and in outdoor air from n.d.-4.6  $\mu$ g/m<sup>3</sup> (66, 84, 106, 135, 149, 164, 197, 210, 213, 214, 239).

Because of overlapping concentrations of both individual compounds and the sum of selected MVOCs in problem and reference buildings, it is difficult to recognise problem buildings on the basis of MVOC measurements, or to establish reference values for MVOCs, though some suggestions have recently been presented. Based on the presented data, concentrations  $\geq 20 \ \mu g/m^3$  of 2-octen-1-ol,  $\geq 10 \ \mu g/m^3$  of 1-octen-3-ol,  $\geq 1 \ \mu g/m^3$  of 3-methyl-2-butanol, 2-methyl-1-propanol, 2-isopropyl-3-methoxypyrazine, and  $\geq 0.1 \, \mu g/m^3$  of geosmin and 3-octanol could be assumed to indicate an abnormal level (Table 5). According to Lorenz et al (135), the detection of main indicators, i.e. 3-methylfuran, 1-octen-3-ol, and/or dimethyl disulphide at concentrations above  $0.05 \ \mu g/m^3$  would clearly indicate a microbial source. In addition, the presence of at least one of the main indicators and the sum of 8 MVOCs exceeding 0.6  $\mu$ g/m<sup>3</sup> or 1.0  $\mu$ g/m<sup>3</sup> would be an indication of a probable or a very probable microbial source, respectively. To avoid false positive results, new buildings (< 6 months), rooms with flower pots, waste, and pet cages, and interference from smoking, cooking and baking should be excluded (135). At present, the concentration limits suggested by Lorenz et al are difficult to apply universally, since different research groups are measuring different MVOCs. Differences in methods of sampling and analysis add additional variability.

In some studies (66, 212), increased levels of some MVOCs (3-methylfuran, 3-methyl-1-butanol, dimethyl disulphide, 2-hexanone, 2-heptanone, 1-octen-3-ol, and 3-octanone) were detected before remedial actions and moisture and/or microbial contamination control measures in buildings. A decrease in the levels were observed after the mitigation, though measurements in some cases showed contradictory results (66). Again, the selection of compounds (which has never been justified in the MVOC literature) and the analysis methods affect the results.

Only little attention has been paid to MVOCs in work environments with productive microbial sources or high levels of contamination (70). MVOCs, like 3-methyl-1-butanol, 2-methyl-1-butanol, ketones, furans, sulphur compounds, geosmin, and terpenes, have been identified in the air of compost facilities (71, 72, 133, 152, 237). Individual MVOC concentrations have varied from 0.1 to 1000

 $\mu$ g/m<sup>3</sup> (152). In a laboratory study, typical VOCs in composts included carbonyl derivatives, organosulphur compounds, pyrazines, pyridines, and oxygenated monoterpenes. Concentrations of organic sulphur compounds (thioethers, disulphides, and trisulphides) in garden waste were concluded to be sufficiently high  $(10-50 \text{ mg/m}^3)$  to cause irritation and other symptoms of toxicity among waste handling personnel (220). Herr et al reported gradually decreasing concentrations of 11 MVOCs (in the range from 0.005  $\mu$ g/m<sup>3</sup> to 6  $\mu$ g/m<sup>3</sup>) measured at different distances (200-550 m) from a large-scale composting site. The authors demonstrated an association between concentrations of residential bioaerosol pollution including MVOCs (< 200 m from the plant) and complaints of airway irritation (85). In a similar study by Muller et al, compost-derived MVOCs (especially terpenes) were registered by measurements at distances of up to 800 m from the composting facilities. Dispersal of volatile contaminants from the composting plant was associated with odour complaints and irritation symptoms (153). Lappalainen et al measured MVOCs in a horse stable. The concentrations were  $\leq 0.5 \ \mu g/m^3$  for 2-hexanone,  $\leq 4.6 \ \mu g/m^3$  for 2-heptanone, and  $\leq 1.5$  $\mu g/m^3$  for 3-octanone. These authors estimated that the concentrations of potential MVOCs were only approximately 0.07-0.31 % of the concentrations of total VOCs in the horse stable. The emission rates for single VOCs from bedding materials in the stable varied between 0.2 and 2  $\mu$ g/kg/hour, being about ten times higher than the corresponding rates in the laboratory experiments (123).

To conclude, reported individual and total MVOC levels are quite low and barely exceed  $1 \text{ mg/m}^3$  even in fairly contaminated areas.

# 7. Toxicokinetics

The body burden of MVOCs, as of any chemical substance, is influenced by the rate of absorption, distribution, biotransformation, and excretion. MVOCs are by definition volatile hence the dominating route of exposure is via inhalation. Based on experiences with organic solvents, some of which may also be considered as MVOCs, it can safely be assumed that the respiratory uptake of MVOCs is in general considerable and that the dermal uptake of vapour is not more than a few percent of the respiratory uptake. Since MVOCs are small and mostly uncharged molecules they easily diffuse across cellular membranes. Hence, they are readily transported between alveolar air and blood and between blood and other tissues. The distribution of MVOCs in the body depends on their tissue:blood partition coefficients. The log octanol:water partition coefficients of MVOCs are listed in Table 4. Substances with high octanol:water partition coefficients tend to have high tissue:blood and especially fat:blood partition coefficients (172). Examples of such substances are long-chained alcohols (e.g. octanol and decanol), hydrocarbons (e.g. heptene) and ketones (e.g. undecanone), aromatic hydrocarbons and terpenoids (Table 3). For example, the high solubility of terpenes in blood and other tissues suggest a high respiratory uptake and thus accumulation in adipose tissue (65).

Aliphatic and aromatic alcohols and carboxylic acids undergo conjugation with glucuronic acid in liver, kidney, intestine, skin, brain, and spleen. Glucuronides are excreted from the body via urine or bile. Another important conjugation reaction for hydroxyl groups (present in aliphatic alcohols) is sulphation, yielding conjugates that are excreted mainly in urine. Oxidation-reduction systems prevail in the body for the biotransformation of aldehydes, ketones, and alcohols. Aldehydes and ketones can be reduced to alcohols by aldehyde/ketone reductases, alcohols can be oxidised to aldehydes by alcohol dehydrogenase, and then further oxidised to acids by aldehyde dehydrogenase (190). Ketones may also undergo an omega-minus 1 oxidation process to form hydroxyketones and be further metabolised to the corresponding diones (16, 17, 218). One example is 2-hexanone, which undergoes biotransformation to the neurotoxic  $\gamma$ -diketone 2,5-hexanedione (32).

The aromatic ether 3-methylfuran is metabolically activated via microsomal oxidation, cleaving the furan ring to a highly reactive unsaturated dialdehyde (methyl butenedial) that binds covalently to tissue macromolecules (174).

For sulphur-containing compounds, the oxidation of sulphur or desulphuration occurs by the addition of oxygen via cytochrome P450. Alkene epoxidation and oxidative *N*-, *O*-, or *S*-dealkylation proceed via cytochrome P450-mediated reactions to form epoxide and hydroxyalkyl moieties, respectively. The formed aliphatic epoxides are then hydrolysed to dihydrodiol products, and the hydroxyalkyl part decomposes into an aldehyde or ketone and a metabolite containing a free amino, hydroxyl, or sulphhydryl group (190).

#### 8. Biological monitoring

Methods for biological exposure monitoring are available for many organic solvents that also appear as MVOCs, such as acetone, benzene, 1-butanol, 2-butanone, ethylbenzene, 2-hexanone, toluene, terpenes and xylenes or their metabolites (3, 60). However, these methods have been developed and are applicable for much higher exposure levels than those typically encountered in the MVOC setting. For the 15 MVOCs listed in Table 2, no such methods are routinely available. Moreover, although background levels in human blood or breath are found for only a few of the typical MVOCs in the scientific literature, e.g. acetone, ethanol, isoprene, and methanol (62, 193), most MVOCs are likely to be present in small amounts in human tissues, as a result of endogenous or microbial metabolism, or both. Thus, a number of the alcohols, aldehydes, ethers, and esters listed in Tables 3-4 have been detected in exhaled breath of humans (141). The endogenous production and the resulting background levels for these compounds are not known exactly, although some VOC measurements have been performed from the exhaled breath of humans (67, 171). Obviously, endogenous background exposure may invalidate biomonitoring of MVOC at low exposure levels. High metabolism rates and low exposure levels restrict the application of biomonitoring for the exposure assessment of MVOCs. In addition, two or more exposing agents may produce the same metabolites, as it is with 2-hexanone and

*n*-hexane, thus hampering the selection of a specific biomarker for exposure to 2-hexanone.

#### 9. Mechanism of toxicity

MVOCs can elicit a variety of toxic systemic effects at concentrations far higher than relevant in this context. Such toxicity is not discussed in this chapter. However, sensory irritation is a known effect of exposure to MVOCs.

Irritation of the eyes and upper airways, i.e. sensory irritation (also called pungent sensation), is due to stimulation of the trigeminal nerve (52). It has been suggested that the strength of the response depends on the number of occupied receptors. Such receptors have though not yet been identified, but several investigators have suggested their existence (157, 230). It has also been proposed that the magnitude of the responses in turn depends on the chemical structure of the compounds (157). Even small differences in the chemical structure, such as different enantiomers of the same compounds, may affect the potency (104, 158).

Wolkoff *et al* (230) have recently proposed that it is possible to distinguish between four types of different organic compounds in the indoor environment that could provoke sensory irritation in the airways. The groups of the proposed compounds are as follows: 1) chemically non-reactive, stable organic compounds, i.e. octane, toluene, butanol and alike; 2) chemically reactive organic compounds like alkenes that react with ozone alone or with nitrogen dioxide in the presence of light to produce new oxygenated products; 3) organic compounds that form chemical bonds to receptor sites in the mucous membranes; 4) organic compounds with (known) toxic properties; the latter compounds are characterised by effects developed over long duration of exposure. Receptors, if any, that mediate the effects of MVOCs remain to be elucidated and characterised. However, it is most likely that at least some of the MVOC-mediated effects are due to activation of receptors in the airways (5).

It has been proposed that sensory irritation receptors can be activated in different ways. Physical binding to the receptor is typical for non-reactive VOCs including most MVOCs. Chemicals with high water solubility and high reactivity, such as ammonia, formaldehyde and acrolein would lead to receptor activation through modification of the receptor structure or adjacent structures important for receptor activation (5). Other mechanisms that activate receptors associated with sensory irritation include chemical reactions of amines and nucleophilic addition of isocyanates (8).

It has also been suggested that irritation may arise through activation of polymorphonuclear neutrophils (PMNs), alveolar macrophages or other professional phagocytes in the lung tissue (5, 156, 230). These effects are thought to be due to proinflammatory and other bioactive mediators released from the phagocytic cells upon their activation (158). It should be noted that irritation due to an inflammatory reaction and due to stimulation of nerve endings are not related. Finally, pulmonary irritation may appear due to stimulation of vagal nerve endings at the alveolar level. Direct compound-stimulation of vagal nerve endings occurs rapidly in relation to the onset of exposure, and disappears when exposure is terminated. However, these nerve endings can also be stimulated due to oedema. As oedema develops and dissipates slowly, the onset of such an irritative process is slow.

# 10. Effects in animals and in vitro studies

#### 10.1 Irritation and sensitisation

The most apparent effect following acute MVOC exposure is irritation. In this document only irritation from vapours was considered. Skin irritation studies with application of concentrated solutions of the test substance were regarded irrelevant in the context of MVOCs.

Alarie (1966) introduced the so-called mouse bioassay, which was later established as a standard test method for estimating sensory irritancy of airborne chemicals by the American Society for Testing of Materials (ASTM) (14). This method has been widely used to assess the sensory irritation potency of aerosols, gases, and vapours. Thus, the method is also applicable for estimation of airway irritation due to inhalation of MVOCs. In this model, four mice are simultaneously exposed head-only to the exposure agent, and their respiratory function is continuously monitored. When a compound stimulates the trigeminal nerve endings in the nasal mucosa, time of breaking (i.e. when inhalation goes into exhalation) increases, and the respiratory rate of the animals decreases in a dose-dependent manner. The sensory irritation potency can be quantified by the changes in breathing patterns and respiratory functions at a given exposure level (airborne concentrations of a chemical used in the experiments). From these relationships, the concentration causing a 50 % decrease in the respiratory rate ( $RD_{50}$ ) can be estimated (4, 5).

Schaper (1993) compiled a large database of results obtained by this bioassay. In addition to the bioassay, the sensory irritation potencies of non-reactive volatile compounds, such as alkylbenzenes, saturated alcohols, and ketones, can be estimated on the basis of physicochemical descriptors (e.g. molecular weight, vapour pressure or Ostwald gas-liquid partition coefficient). This is possible because the effect of the compounds are probably induced via physical adsorption to the biological receptors (1, 6-8). Since most MVOCs are non-reactive (towards SH or OH groups in proteins), their RD<sub>50</sub>s can be calculated theoretically by means of the physicochemical variables. RD<sub>50</sub>s calculated or determined by the mouse bioassay for some MVOCs are presented in Table 6.

In the case of exposure to MVOCs, low exposure concentrations of several MVOCs frequently occur (in the range of  $ng/m^3 - \mu g/m^3$ ). Effects of exposure to chemical mixtures can be additive, antagonistic, or synergistic. If we assume that additivity prevails at low exposure levels to VOC mixtures, the total sensory irritation potency would be the sum of the effects that each compound would elicit

Compound	RD	<b>)</b> <sub>50</sub>
	mg/m <sup>3</sup>	ppm
2-Methyl-1-propanol	5 499	1 815
3-Methyl-1-butanol	9 325	2 583
3-Methyl-2-butanol	9 645	2 672
2-Pentanol	9 907	2 744
3-Octanol	1 359	255
1-Octen-3-ol	182	35
2-Octen-1-ol <sup>a</sup>	-	-
3-Methylfuran <sup>a</sup>	-	-
2-Hexanone	10 449	2 550
2-Heptanone	4 163	891
3-Octanone	17 586	3 359
2-Methylisoborneol	811	118
2-Isopropyl-3-methoxy-pyrazine <sup>a</sup>	-	-
Geosmin	216	29
Dimethyl disulphide <sup>b</sup>	37 330	9 700

**Table 6.** The  $RD_{50}$  values for some MVOCs (114, 168, 180).

<sup>a</sup> fundamental understanding or data about the substance's potency as a sensory irritant is missing <sup>b</sup> estimated by document authors by using the formula:

 $\log RD_{50} (ppm) = 2.693 + (0.887 \cdot \log P^{\circ}) (P^{\circ}, mmHg) (8)$ 

alone (156). The estimation is based on the sum of ratios of the fractional concentration (c) and the  $RD_{50}$  of each compound:

 $1/RD_{50mixture} = \sum (c_n/RD_{50n})$ 

It has been proposed that effects at higher exposure concentrations might show synergistic interactions (156). This was verified by Korpi *et al* who investigated the sensory irritation potency of 1-octen-3-ol, 3-octanol and 3-octanone, separately, and the potency of the MVOC mixture including 2-methyl-1-propanol, 3-methyl-1-butanol, 1-octen-3-ol, 2-heptanone and 3-octanone by the mouse bioassay. The RD<sub>50</sub>s for these MVOCs are presented in Table 6. The RD<sub>50</sub> for the mixture was 3.6 times lower than estimated from the sum of fractional concentrations and the RD<sub>50</sub>s of individual compounds (the estimated RD<sub>50</sub> was 504 mg/m<sup>3</sup> and the observed RD<sub>50</sub> 142 mg/m<sup>3</sup>). As expected, if a particular compound in the mixture is much more potent than the other ones, it may dominate the effect (114).

Korpi *et al* also investigated the effect of repeated exposures (30 minutes per day during four consecutive days) to 3-octanone ( $3531 \text{ mg/m}^3$ ), 1-octen-3-ol ( $36 \text{ mg/m}^3$ ), or to a mixture of 2-methyl-1-propanol ( $\sim 6 \text{ mg/m}^3$ ), 3-methyl-1-butanol ( $\sim 6 \text{ mg/m}^3$ ), 1-octen-3-ol ( $\sim 19 \text{ mg/m}^3$ ), 2-heptanone ( $\sim 19 \text{ mg/m}^3$ ) and 3-octanone ( $\sim 6 \text{ mg/m}^3$ ) by the mouse bioassay. The levels for *individual* MVOC experiments were chosen to produce a clear respiratory rate decrease (from 11-20 % for 1-octen-3-ol with a very steep dose-response curve to 32 % for 3-octanone). The proportions of the MVOCs in the *mixture* were chosen to reflect those measured in mouldy buildings or mouldy building materials, and test concentrations were chosen to maximally yield a 32 % decrease in the respiratory frequency. For single MVOCs, no changes in the responses were observed between the exposures,

and only a very slight adaptation in respiratory function occurred along with the exposures to the MVOC mixture. The authors concluded that MVOCs seem to act as "pure" sensory irritants and the effects of a short-term, repeated exposure seem non-cumulative and transient (115).

Histamine induces inflammation, and in an *in vitro* study by Larsen *et al* human bronchoalveolar lavage cells were incubated with MVOCs of *Trichoderma viride* (e.g. 2-methyl-1-propanol, 3-methyl-1-butanol, 2-methyl-1-butanol, 1-pentanol) with the result of evoked histamine release from the cells. The histamine release was stastistically significant when it increased from approximately 12 to 20 % at VOC concentrations ranging from 0.04 to 10 % (v/v) (124). Hypothetically, exposure to MVOCs, especially to reactive oxygen metabolites (after reactions with ozone and other oxidants) may then function as adjuvants by inducing nonspecific airway hyperresponsiveness and inflammation (156).

In the reactions between oxidants and unsaturated hydrocarbons intermediates formed may be much more irritating than the original reactants and end-products (179, 208, 228, 229).

In summary, irritation is the most probable effect of MVOC exposure and  $RD_{50}$ s for a number of MVOCs have been estimated by the mouse bioassay. Assuming additivity at low exposure concentrations the total sensory potency of an MVOC mixture can be calculated. However, at higher concentrations synergy may occur. Effects of short-term, repeated exposures at levels high enough to produce a clear respiratory rate decrease seem to be non-cumulative and transient.

#### **10.2 Effects of single exposure**

The most apparent effect following acute MVOC exposure is irritation (Chapter 10.1). However, acute high-level vapour exposure to compounds denoted MVOCs generally has the potential to cause narcosis, central nervous system disturbance and death (11).

The lethal dose for 50 % of the exposed animals at single administration ( $LD_{50}$ ) and the corresponding lethal concentration for 50 % of the animals at single inhalation exposure ( $LC_{50}$ ) are measures of the general acute toxic potency of a chemical (11) for comparison with other substances. In Table 7,  $LC_{50}$ -values, lowest observed lethal concentrations ( $LC_{L0}$ ), and dermal  $LD_{50}$ s are presented for some of the 15 selected MVOCs. However, for several of those MVOCs such values have not been determined. According to the Hodge and Sterner scale the substances listed in Table 7, with the possible exception of dimethyl disulphide, would be classified as slightly toxic, i.e. having an  $LC_{50}$  in the range 1 000-10 000 ppm in rats (42). Also the dermal  $LD_{50}$ s indicate a low acute toxicity termed as slightly (350-2 810 mg/kg) or practically non-toxic (2 820-22 590 mg/kg) according to the same toxicity scale. The concentrations of MVOCs need to be thousands of mg/m<sup>3</sup> (Table 7) in order to produce lethal effects in animals, whereas the concentrations of individual MVOCs indoors in general are in the range of hundred ng to <1 mg/m<sup>3</sup> (Table 5).

Compound/ Species		LC <sub>50</sub> <sup>a</sup> ration Dura	tion		LC <sub>Lo</sub> <sup>b</sup> ration Du	ration	Dermal LD <sub>50</sub> <sup>c</sup>	Reference
	mg/m <sup>3</sup>	ppm	h	mg/m <sup>3</sup>	ppm	h	mg/kg	
2-Methyl-1-pro	panol							
Rat	19 200- 24 200	6 300- 8 000	4	24 200 8 000	8 000 2 600	4 4		(88, 162, 176)
Guinea pig Mouse Rabbit	19 900 15 500 26 200	6 600 5 100 8 600	4 2 -					(88, 176) (88, 176) (88, 176)
	2 600	860	4					
Rabbit, male Rabbit, female							>2 000 2 460 (1 790-3 390	(24) (24)
Rabbit Rabbit, occlusive							3 400 4 240 (2 520-7 120	(24) (24)
3-Methyl-1-but	anol						(2 320-7 120	)
Rabbit <i>3-Octanol</i>							3 240	(2)
Rabbit <i>1-Octen-3-ol</i>							>5 000	(25)
Rabbit <i>3-Methylfuran</i>							3 300	(26)
Mouse Rat	3 000 6 700	900 2 000	1 1					(78) (79)
Hamster 2-Hexanone	>26 400	>7 900	1					(79)
Rat Guinea pig	32 700	8 000	4	82 000 40 000	20 000 9 800	1.2 0.5		161, 162, 176) (162, 176)
Rabbit					,	0.0	4 800	(31)
2-Heptanone								
Rat Guinea pig Rabbit				18 600 22 400	4 000 4 800	4 4	13 000	(176) (176) (218)
<i>3-Octanone</i> Rabbit,							>5 000	(188)
occlusive Dimethyl disulp	phide							
Rat Mouse Rat	15.8 <sup>d</sup> 12.3 <sup>d</sup> 3 100							(47, 176) (47, 176) (47)
Rat Rabbit	4 800	1 200	4				>2 000	(47) (13)

Table 7.  $LC_{50}s$ ,  $LC_{Lo}s$  and  $LD_{50}s$  for some MVOCs most often reported in field studies.

 $^{a}$  = lethal concentration for 50 % of the animals at single exposure

<sup>b</sup> = lowest observed lethal concentration

 $^{\circ}$  = lethal doses for 50 % of the animals at single administration

 $^{d}$  = the validity of the value has been questioned (47)

The cytotoxicity of 13 so called MVOCs including e.g. 1-octen-3-ol, 3-octanol, 3-methyl-1-butanol, 2-methyl-1-propanol, 3-octanone, 2-heptanone and 2-hexanone was studied using a human lung carcinoma epithelial cell line A549 in a colony formation assay and 2 colorimetric assays. 1-Octen-3-ol and 3-octanol were approximately 10-100 times more cytotoxic than the other MVOCs. However, all tested MVOCs were more than 1 000-fold less toxic than the known cytotoxic substance gliotoxin measured as the concentration resulting in 50 % inhibition of colony growth or absorbance (120).

Other toxicological data of relevance on the 15 selected substances are presented below:

2-Methyl-1-propanol: Signs of toxicity reported in acute inhalation studies (> 19 000 mg/m<sup>3</sup>) in several species include narcosis, irritant effects on the mucous membranes, and effects on the liver and kidneys (24).

*3-Methylfuran:* Acute inhalation studies of 3-methylfuran have revealed damage to the epithelium lining the small airways at high doses. Mice appear to be more sensitive than rats and hamsters with extensive necrosis of the bronchiolar epithelium within one day following a 1-hour exposure to initial concentrations of 343-906 ppm (1 149-3 038 mg/m<sup>3</sup>). Virtually complete regeneration of the epithelium was observed within 21 days. 3-Methylfuran-induced injury occurred also in the liver, lymphoid system, and nasal mucosa although species differences were present (78, 79, 150).

#### Conclusion

Single exposure data are lacking or are very scarce for all of the 15 selected substances. Available data on the alcohols, ketones and 3-methylfuran suggest a slight, and those on dimethyl disulphide a slight to moderate acute toxicity. Reported effect levels are more than three orders of magnitude higher than reported MVOC indoor air levels.

#### 10.3 Effects of short-term and long-term exposure

Repeated exposure data, available for 7 of the 15 selected MVOCs, are presented below. Focus is on inhalation and the lowest administered doses. If inhalation data are lacking or scarce, however, oral data have been included.

*3-Methyl-1-butanol:* No compound-related, toxicologically relevant effects were found in rats exposed in drinking water to doses up to 1 068 (males) or 1 431 (females) mg/kg body weight for 90 days (82). Oral exposure of rats to doses up to 1 000 mg/kg body weight/day, 7 days/week for 17 weeks, was not accompanied by adverse effects. However, a decreased body weight gain, which was ascribed a reduced food intake was reported at the highest dose (82).

2-Methyl-1-propanol: A comprehensive set of neurotoxicity tests including an assessment of complex behaviour dependent on learning and memory, full histopathology and blood chemistry evaluations were conducted following the exposure of rats to 0, 250, 1 000, and 2 500 ppm (0, 760, 3 030, and 7 600 mg/m<sup>3</sup>) for 90 days. A slight decrease in response to external stimuli was observed during

the actual exposures at all concentrations. There were no morphological or behavioural effects indicative of a specific, persistent or progressive effect on the nervous system at any exposure level (132).

In two 90-day studies in male and female rats and according to OECD guidelines, no-effect-levels of approximately 316 and 1 450 mg/kg body weight/day were determined following administration by gavage and in drinking water, respectively (24, 181).

*3-Octanol:* In a recent subchronic oral toxicity study, no effects were observed in rats dosed with 25 mg/kg body weight. Treatment-related lesions in the kidney (100 mg/kg body weight) and liver (400 mg/kg body weight) were observed at higher dose-levels (134).

*3-Methylfuran:* In male and female hamsters and mice exposed for a total of 2 hours to an initial concentration of 8 400 ppm (28 200 mg/m<sup>3</sup>) and a final ditto of 1 900 ppm (6 400 mg/m<sup>3</sup>), and for 1 hour to an initial concentration of 700 ppm and a final concentration of 400 ppm (2 400 and 1 300 mg/m<sup>3</sup>), respectively, once a week for 10 consecutive weeks, the result of respiratory function tests and the histopathologic evaluation of the lungs did not reveal any major long-lasting changes 10 months later. In mice, the tumour incidence in exposed animals was not increased when compared to controls 2 years after exposure (227).

2-Hexanone: The target tissue in 2-hexanone toxicity is the nervous system. Neurotoxic effects, expressed as peripheral neuropathy, and weight loss/retarded weight gain, have been observed in several species after repeated exposures and different modes of administration (31, 32, 138).

Exposure to 40 ppm (164 mg/m<sup>3</sup>) of 2-hexanone vapour 8 hours/day, 5 days/ week for 22-88 days did not result in any clinical or pathological signs of neuropathy in rats, whereas 3/20 rats showed demyelination of the sciatic nerve after 13 weeks of exposure to 50 ppm (205 mg/m<sup>3</sup>) of 2-hexanone vapour under similar conditions (LOEL). In rats exposed for 6 months a reduced nerve conduction velocity was observed within 17 weeks. Even after 6 months of exposure no clinical signs of neuropathy were observed, but wide-spread demyelination of the sciatic nerve was seen in 32/40 rats. It cannot be ruled out that prolongation of the 40 ppm exposure would have revealed signs of neuropathy at a later time.

In another study, exposure to 100 ppm (410 mg/m<sup>3</sup>) of 2-hexanone vapour for 6 hours/day, 5 days/week resulted in a reduced motor conduction velocity of the sciatic-tibial nerve in male rats within 29 weeks (n=30) as well as in male monkeys (within 9 months) (n=24). Complete recovery occurred in monkeys within 2 months post-exposure.

Rats exposed to 200 ppm (820 mg/m<sup>3</sup>) of 2-hexanone vapours for 6 weeks (8 hours/day, 5 days/week) showed several signs of neuropathy (axonal hypertrophy and segmental breakdown of myelin). Continuous inhalatory exposure of 12 rats to 225 ppm (920 mg/m<sup>3</sup>) resulted in clinical paralysis after 66 days.

Chicken continuously exposed to 100 ppm (410 mg/m<sup>3</sup>) 2-hexanone vapour developed hind limb dragging after 4-5 weeks.

The neurotoxicity of 2-hexanone is enhanced by co-exposure to other chemicals, especially other ketones like 2-butanone. 2-Hexanone can also potentiate the toxicity of other compounds, such as chloroform and carbon tetrachloride (31, 32, 138).

2-*Heptanone:* Repeated (n=19) inhalation exposures of 115-1 500 ppm (540-7 000 mg/m<sup>3</sup>) for 6-8 hours did not cause behavioural changes in rats, but some reduction in response rate occurred at exposure to 1 575-1 900 ppm (7 400-8 900 mg/m<sup>3</sup>). Tolerance developed over the course of the study (12).

No adverse effects on cardiopulmonary function, clinical chemistry, or signs of neurotoxicity were noted in male rats and monkeys after inhalation exposures up to approximately 1 000 ppm (4 700 mg/m<sup>3</sup>) for 9-10 months, 6 hours/day, 5 days/ week (97, 98, 140). 2-Heptanone has been shown to increase the chloroform induced nephro- and hepatotoxicity in rats (86).

*Dimethyl disulphide:* No toxic effects were noted in rats after inhalation of 100 ppm ( $390 \text{ mg/m}^3$ ), 6 hours/day for 20 days (137).

Two other inhalation studies of rats exposed to dimethyl disulphide 6 hours/ day, 5 days/week for 13 weeks, and according to OECD guidelines, have been performed. In the first study, reduced body weight and food intake, and changes in some serum biochemical parameters were observed at 25 ppm (96 mg/m<sup>3</sup>) in the males. At 125 ppm (480 mg/m<sup>3</sup>) the same effects and in addition an increase in some organ weights were seen in both genders. No treatment-related effects were observed at the lowest exposure level of 5 ppm (19 mg/m<sup>3</sup>) (108). In the other study, hypoactivity, reduced body weight and food consumption, and changes in organ weights and in white blood cell counts were reported. Reversible microscopic changes in the nasal mucosa were noted at all exposure levels (10, 50, 150 and 250 ppm) (38, 190, 580 and 960 mg/m<sup>3</sup>). The NOEL was judged to be slightly lower than 10 ppm (13).

### Conclusion

The animal short- and long-term toxicity database is poor for most of the 15 selected substances. For some of the alcohols, and for 2-heptanone, the available data indicate a slight toxicity. Two 13-week inhalation studies on dimethyl disulphide in rats suggest a NOEL below 10 ppm (38 mg/m<sup>3</sup>). In rats, peripheral neuropathy has been observed after exposure to 50 ppm (205 mg/m<sup>3</sup>) 2-hexanone (LOEL). Thus, adverse effects from some of the selected substances have been reported only at doses far higher than what can be obtained when their main source is microbial.

#### 10.4 Mutagenicity and genotoxicity

Genotoxicity tests on the 15 substances selected for further investigation are summarised in Table 8. With few exceptions results are negative. Data are lacking for some of the substances.

As examples of recent studies on MVOCs, Kreja and Seidel (119) investigated genotoxic, clastogenic, and mutagenic potential of 16 MVOCs *in vitro* (Table 8).

Each compound induced DNA damage only under cytotoxic conditions. Clastogenic (causing chromosomal breaks) and mutagenic effects were not detected.

In contrast, SOS<sup>a</sup>-inducing activity was reported for 15 out of 20 MVOCs including 2-methyl-1-propanol, 3-methyl-1-butanol, 3-methyl-2-butanol, 2-pentanol, 3-octanol, 1-octen-3-ol, 2-hexanone, 2-heptanone, and 3-octanone in the luminescent *umu* test. 3-Methyl-2-butanone and 3-methyl-2-butanol were the only MVOCs that were tested positive also in the less sensitive conventional light absorption *umu* test. These two MVOCs were subsequently tested also in the Ames test and were reported positive (154).

## **10.5** Carcinogenicity

The International Agency for Research on Cancer (IARC) has not evaluated the carcinogenicity of any of the 15 MVOCs listed in Table 2. The United States National Toxicology Program (NTP) has not listed those chemicals in its report on carcinogens (165). In the broader list of identified MVOCs (Table 3), some substances (such as formaldehyde) are classified as human carcinogens or as possible human carcinogens (such as acetaldehyde, ethylbenzene, isoprene, and styrene). Considering the low concentrations encountered in the MVOC context, cancer is not likely to be a concern. The very few studies that were retrieved from the literature are described below.

*3-Methyl-1-butanol*: Oral administration of approximately 81 mg/kg body weight, twice a week for 135 weeks, or subcutaneous injection of approximately 32 mg/kg body weight, once a week, for 95 weeks, induced an increase in the incidence of malignant tumours in rats (4 and 10, respectively). No malignant tumours were found in the controls (82). The study design and documentation of reported findings have several flaws, which hampers the interpretation of the data, e.g. a low number of animals, unknown sex ratios, lack of information on tumour incidences in historical controls, and the exceeding of the maximum tolerated dose evidenced by chronic toxic effects on the liver and haematopoietic system.

2-Methyl-1-propanol: In rats, oral administration of approximately 160 mg/kg body weight, twice a week for 72 weeks, or subcutaneous injection of approximately 41 mg/kg body weight, twice a week, for 90 weeks led to an increase in the number of malignant tumours. No malignant tumours were seen in the controls (24). The study does not comply with current standards (see text on 3-methyl-1-butanol).

<sup>&</sup>lt;sup>a</sup> The SOS response in bacteria describes changes in gene expression in response to DNA damage.

Commund/Protections	Endnoint	Concentration	Matabalia	Docu14	Defension
Compound test system	EIIIIIDOIIII	Concentration	Metabolic activation <sup>a</sup>	Kesult	Kelerence
2-Methyl-1-propanol					
Ames test /S. Typhimurium TA98, 100, 1535, 1537, 1538	Gene mutation	5-5 000 µg/plate	No/Yes	Negative/Negative	(43)
Ames test /S. Typhinurium TA97, 98, 100, 1535, 1537, 1538	Gene mutation	100-10 000 μg/plate	No/Yes	Negative/Negative	(43)
Light absorption umu test / S. Typhimurium TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test / S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
E. coli/ WP2 uvrA	Gene mutation	5-5 000 $\mu$ g/plate	No/Yes	Negative/Negative	(43)
Chinese hamster V-79 / HPRT assay	Gene mutation	107 mM	No/Yes	Negative/Negative	(43)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	53, 270 mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts / Alkaline comet assay	DNA damage	53, 270 mM	No	Negative/Positive <sup>b</sup>	(119)
Human peripheral blood cells / Alkaline comet assay	DNA damage	53, 270 mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts: Micronucleus test HPRT assay	Clastogenic effects: Micronuclei Gene mutation	11, 53 mM 0-107 mM	No No/Yes	Negative Negative/Negative	(119)
3-Methyl-1-butanol					
Ames test/ S. Typhimurium TA98, 100, 1535, 1537	Gene mutation	Not given	No/Yes	Negative/Negative	(82)
Light absorption <i>unu</i> test / <i>S. Typhimurium</i> TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test /S. <i>Typhimurium</i> TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)

Table 8. Genotoxicity tests with some MVOCs often reported in field studies.

Compound/Test system Endpoint	Endpoint	Concentration	Metabolic activation <sup>a</sup>	Result	Reference
Chinese hamster V-79 / HPRT assay	Gene mutation	51.5 mM	No/Yes	Negative/Negative	(43)
Human blood cells / Comet assay	DNA damage (strand breaks, alkali-labile sites)			Negative	(82)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	23, 46, 91mM	No	Negative	(119)
V-79 Chinese hamster fibroblasts / Alkaline comet assay	DNA damage	46, 91 mM	No	Negative	(119)
Human peripheral blood cells / Alkaline comet assay	DNA damage	23, 91 mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts: Micronucleus test HPRT assav	Clastogenic effects: Micronuclei Gene mutation	5, 9, 23 mM 0-51.5 mM	No/Yes No/Yes	Negative/Negative Negative/Negative	(119)
Rat bone marrow <i>in vivo</i>	Chromosmal abberation Polyploid cells	1/5 LD <sub>50</sub>		Positive Negative	(82)
	Chromosome gaps			Negative	
3-Methyl-2-butanol					
Ames test/S. Typhimurium TA98, 100	Gene mutation	0-2.5 $\mu$ l/plate	No/Yes	Positive/Positive	(154)
Light absorption <i>umu</i> test / S. Typhimurium TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Positive	(154)
Luminescent <i>umu</i> test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	45, 90 mM	No	Negative	(119)
V-79 Chinese hamster fibroblasts: Micronucleus test	Clastogenic effects: Micronuclei	45, 90 mM	No	Negative	(119)

Compound/Test system Endpoint	Endpoint	Concentration	Metabolic	Result	Reference
			acuvation		
2-Pentanol					
Light absorption <i>umu</i> test /S. Typhimurium TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
3-Octanol					
Light absorption <i>umu</i> test /S. Typhimurium TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
Human lung carcinoma epithelial A549 cells/ Alkaline comet assay	DNA damage	6.2, 31mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts/ Alkaline comet assay	DNA damage	6.2, 31 mM	No	Negative/Negative	(119)
V-79 Chinese hamster fibroblasts	Micronuclei	3.1, 6.2  mM	No	Negative	(119)
I-Octen-3-ol					
Light absorption <i>umu</i> test /S. <i>Typhimurium</i> TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive? °	(154)
Chinese hamster V-79 / HPRT assay	Gene mutation	5 mM	No/Yes	Negative/Negative	(43)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	0.6, 6.4  mM	No	Negative	(119)
V-79 Chinese hamster fibroblasts / Alkaline comet assay	DNA damage	0.6, 6.4  mM	No	Negative/Positive <sup>b</sup>	(119)

.

Compound/Test system	Endpoint	Concentration	Metabolic activation <sup>a</sup>	Result	Reference
Human peripheral blood cells / Alkaline comet assay	DNA damage	0.6, 6.4  mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts Micronucleus test HPRT test	Clastogenic effects: Micronuclei Gene mutation	0.6, 3.2 6.4 mM 0-5 mM	NoYes No/Yes	Negative/Negative Negative/Negative	(119)
2-Octen-I-ol				0	
No data					
3-Methylfuran					
No data					
2-Hexanone					
Light absorption <i>umu</i> test /S. <i>Typhimurium</i> TA1535 / pSK1002 and	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
Chinese hamster V-79 / HPRT assay	Gene mutation	40 mM	No/Yes	Negative/Negative	(43)
Saccharomyces cervisiae	Chromosome loss	48 mM	No	Weakly positive <sup>d</sup>	(143)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	40, 80 mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts / Alkaline comet assay	DNA damage	40, 80 mM	No	Negative	(119)
Human peripheral blood cells / Alkaline comet assay	DNA damage	40, 80  mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts: Micronucleus test HPRT assay	Clastogenic effects: Micronuclei Gene mutation	40, 80mM 0-40 mM	No/Yes No/Yes	Negative/Negative Negative/Negative	(119)

Compound/Test system Endpoint	Endpoint	Concentration	Metabolic activation <sup>a</sup>	Result	Reference
2-Heptanone					
Light absorption <i>umu</i> test /S. Typhimurium TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent <i>umu</i> test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
Chinese hamster V-79 / HPRT assay	Gene mutation	18.2 mM	No/Yes	Negative/Negative	(43)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	7, 35, 70 mM	No	Negative/Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts / Alkaline comet assay	DNA damage	7, 35, 70 mM	No	Negative/Negative/Positive <sup>b</sup>	(119)
Human peripheral blood cells/ Alkaline comet assay	DNA damage	7, 35 mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts	Clastogenic effects:				(119)
Micronucleus test	Micronuclei	17, 35 mM	No	Negative	
HPRT test	Gene mutation	18.2 mM	No/Yes	Negative/Negative	
3-Octanone					
Light absorption umu test /S. Typhimurium TA1535 / pSK1002	DNA damage (SOS-induction)	Not given	No	Negative	(154)
Luminescent umu test /S. Typhimurium TA1535 / pTL210	DNA damage (SOS-induction)	Not given	No	Positive	(154)
Chinese hamster V-79 / HPRT assay	Gene mutation	$14.4 \mathrm{mM}$	No/Yes	Negative/Negative	(43)
Human lung carcinoma epithelial A549 cells / Alkaline comet assay	DNA damage	6, 31, 62 mM	No	Negative/Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts / Alkaline comet	DNA damage	6, 31, 62  mM	No	Negative/Negative/Negative	(119)
assay					

Compound/Test system	Endpoint	Concentration	Metabolic activation <sup>a</sup>	Result	Reference
Human peripheral blood cells / Alkaline comet assay	DNA damage	6, 62 mM	No	Negative/Positive <sup>b</sup>	(119)
V-79 Chinese hamster fibroblasts:	Clastogenic effects:				(119)
Micronucleus test	Micronuclei	16, 31 mM	No	Negative	
HPRT test	Gene mutation	0-14.4 mM	No/Yes	Negative/Negative	
Geosmin					
Ames test/ S. Typhimurium TA98, 100,	Gene mutation	3.93-2 000 и g/plate	No/Yes	Negative/Negative	(43)
Ames test/ S. Typhimurium TA98, 100, 1535, 1537, 1538	Gene mutation	Not given		Negative	(107)
Arnes test/ S. Typhimurium TA98, 100, 102, 1535, 1537, 1538	Gene mutation	Not given	No/Yes	Negative/Negative	(155)
Umu test/ S. Typhimurium	DNA damage (SOS-induction)	$0-400 \ \mu \mathrm{g/ml}$	No/Yes	Negative/Negative	(155)
Chinese hamster lung fibroplast cell line CHL	Chromosome abberation	0-0.15 mg/ml	No/Yes	Negative/Negative	(142)
2-Methylisoborneol					
Ames test/ S. Typhimurium TA98, 100	Gene mutation	9.85-5 000 μg/plate	No/Yes	Negative/Negative	
Ames test/ S. Typhimurium TA98, 100, 102, 1535, 1537, 1538	Gene mutation	Not given	No/Yes	Negative/Negative	(155)
Umu test/ S. Typhimurium	DNA damage (SOS-induction)	$0-400 \mu \mathrm{g/ml}$	No/Yes	Negative/Negative	(155)
Chinese hamster lung fibroplast cell line CHL	Chromosome abberation	0-0.2 mg/ml	No/Yes	Negative/Negative	(142)

Table 8. Genotoxicity tests with some MVOCs often reported in field studies.	ften reported in field studies.				
Compound/Test system	Endpoint	Concentration	Metabolic activation <sup>a</sup>	Result	Reference
Dimethyl disulphide					
Ames test/ S. Typhinurium TA98, 100, 102	Gene mutation	0.011-1 100 μg/plate	No/Yes	Negative/Negative	(43)
Ames test/ S. Typhimurium TA98, 100, 1535, 1537, 1538	Gene mutation	50-5 000 μg/plate	No/Yes	Negative/Negative	(13)
Chinese hamster ovary / HPRT assay	Gene mutation	$0.46-1\ 000\ \mu g/ml$	No/Yes	Negative/Negative	(13)
Rat hepatocytes in primary culture	DNA damage and repair assay	$1-300 \ \mu g/ml$	No	Negative	(13)
Human lymphocytes	Chromosome aberration	$3.7-300 \ \mu g/ml$	No/Yes	Ambiguous/ Ambiguous	(13)
Mouse bone marrow in vivo	Micronuclei	0, 250, 500  ppm		Negative	(13)
Rat hepatocytes <i>in vivo</i>	Unscheduled DNA synthesis	0, 500 ppm		Negative	(13)
2-Isopropyl-3-methoxy-pyrazine					
No data					

<sup>a</sup> Addition of a metabolising system, usually the microsomal fraction of an Aroclor 1254-induced rat liver, a so-called S9-mix.

<sup>b</sup> At cytotoxic concentrations.

 $^{\circ}$ ? = Å result where the number of revertant colonies was 1.5-2 times that of spontaneous colonies. <sup>d</sup> Combined treatment with methyl ethyl ketone potentiated the effect. 2,5-Hexandione was strongly positive alone and in combination with acetone or methyl ethyl ketone.

#### **10.6 Reproductive and developmental studies**

Studies on reproductive and developmental effects of MVOC exposure are rare. Of the selected 15 MVOCs, data exist for 4 compounds.

*3-Methyl-1-butanol* was tested for developmental toxicity in pregnant Wistar rats and Himalayan rabbits. Twenty-five female rats and 15 female rabbits per group were exposed to 3-methyl-1-butanol vapour at concentrations of 510, 2 500, or 9 800 mg/m<sup>3</sup>, 6 hours/day. The rats were exposed on days 6-15 postcoitum and the rabbits were exposed on days 7-19 after insemination. The dams were sacrificed on day 20 respectively day 29. The foetuses were removed from the uterus and examined for malformations and variations externally, in soft, and in skeletal tissue. Exposure to 9 800 mg/m<sup>3</sup> caused eye irritation in dams of either species during the exposure. Pregnancy and litter data were similar in all groups, and no signs of embryo- or foetotoxicity were observed in foetuses of either species. The overall incidence of variations was significantly increased in rabbit offspring in the highest exposure group, however the number of malformations exhibited no differences between groups (109).

2-Methyl-1-propanol was tested for developmental toxicity in a study of similar design as described above for 3-methyl-1-butanol, with exposure concentrations of 510, 2 500, or 9 800 mg/m<sup>3</sup>. Pregnancy and litter data were similar in all groups, and no signs of embryo- or foetotoxicity were observed in foetuses of either species. The overall incidence of variations was significantly increased in rabbit offspring in the highest exposure group, whereas the number of foetuses with retarded ossification was lowest in this group. The number of malformations exhibited no differences between groups (109).

2-Hexanone: Male rats (strain not specified) were exposed to 700 ppm (2 870 mg/m<sup>3</sup>) 2-hexanone for 72 hours weekly (two 20-hour and two 16-hour exposure periods) for 11 weeks. Exposure was associated with a marked reduction in body weight. Generally, organ weight changes reflected the reduction in body weight, however both absolute and relative weight of testes were significantly reduced. Microscopically, atrophy of the testicular germinal epithelium was observed (105).

Peters *et al* exposed groups of 25 pregnant Fisher 344 rats by inhalation to 0, 500, 1 000, or 2 000 ppm (2 050, 4 100 or 8 200 mg/m<sup>3</sup>) 2-hexanone for 6 hours/day throughout gestation. Five male and female pups per group (one from a litter) were examined for developmental landmarks and tested for simple reflexes, activity in open field and running wheel, food maze behaviour, a swimming stress test, and shock avoidance. Tests were performed at puberty, adulthood, and old age, however all exposure groups were not tested in all tests at all ages. Additional animals were tested for pentobarbital-induced sleeping time, clinical chemistry, and haematology. Offspring from the 500 ppm group had to be discarded due to non-treatment related circumstances. Maternal weight gain was non-significantly reduced at the two higher concentrations, and a detectable change in neurological function was observed in 2 000 ppm dams. These females produced smaller litters of lower weight pups, the latter persisting in male offspring throughout life.

Behavioural alterations were detected in most tests. In the food maze, offspring from exposed animals performed better at puberty but poorer as adults. Treated offspring exhibited reduced activity in the open field early in life (2 000 ppm) and increased activity in the running wheel up until adulthood. Performance in avoidance conditioning was poorer in puberty females, and the treated animals generally exhibited increased random movement during the intertrial interval (1 000 ppm). A few sporadic changes were also noted in clinical chemistry values, however consistency lacked. Haematology revealed no differences between exposed and control groups (170).

*Dimethyl disulphide:* Rats were exposed to 0, 5, 15 and 50 ppm (0, 19, 58 and 190 mg/m<sup>3</sup>) dimethyl disulphide for 6 hours/day from day 6 to day 15 of gestation. Exposure to 50 ppm caused marked maternal toxicity and foetal growth retardation. At 15 ppm, there were less marked maternal toxicity and no foetal effects. No maternal or foetal effects were observed at 5 ppm (13).

Smotherman and Robinson examined the behavioural responses of near term rat foetuses to a range of potential chemosensory fluids, including milk and dimethyl disulphide, a constituent of pup saliva promoting postnatal nipple attachment. Only milk and dimethyl disulphide altered foetal motor activity and foetal responsiveness to perioral cutaneous stimulation, suggesting dimethyl disulphide acts as a chemical messenger during the neonatal period. Furthermore, the opiod antagonist naloxone reversed the pup behavioural response to dimethyl disulphide, indicating that this chemical is capable of promoting opiod activity, as is milk (194).

### Conclusion

Studies are lacking for most of the 15 selected substances. Some effects on postnatal development and behaviour have been reported after 2-hexanone exposure of pregnant rats. However, considering the low levels encountered in MVOC settings reproductive and developmental effects caused by MVOCs are unlikely.

## 11. Observations in man

## 11.1 Odour sensation, irritation and sensitisation

Complaints of unpleasant odours are often presented in damp buildings with microbial contamination (173, 192, 197). Because many MVOCs have musty, earthy, mushroom-like, sweet, and/or fruity smell, MVOCs have been assumed to be responsible for odour sensations in problem buildings (106, 178). However, the occurrence of MVOCs or odours is not a direct measure of the extent of microbial growth in a building (96, 197) because many factors affect MVOC levels indoors as well as the human sensation of odours (the odour threshold of an MVOC, occupants' susceptibility to odour). The odour thresholds may vary at least by a factor of 10<sup>8</sup> (from 10<sup>-7</sup> to 10<sup>1</sup> ppm) between individual MVOCs and by a factor of

 $10^{1}$ - $10^{4}$  within the same compound between different studies. In one study, odour complaints in problem buildings were reported at the sum concentration of 13 MVOCs of 15  $\mu$ g/m<sup>3</sup>, while in the same study odour was not noticed even at concentrations up to 40  $\mu$ g/m<sup>3</sup> (197). The authors stated that odour complaints are related to the occurrence of individual compounds or their combinations rather than the sum of selected MVOCs.

Despite the lack of studies on concentration-response relationships, MVOCs have been associated with general discomfort (i.e. headache, dizziness and fatigue) in buildings when occurring in concentrations above the odour thresholds. In addition, in epidemiological and case studies, the presence of MVOCs or musty, earthy odours has been related to the prevalence of eye, nose, and throat irritation, wheezing and other asthma-like symptoms (63, 92, 111, 177, 204, 212). Perceived mould odour was found to be a risk indicator for occurrence of nasal congestion, excretion, cough, phlegm, wheeze, and the occurrence of symptoms was related to the frequency of the days with mould odour expressed (92). In this study, the data on symptoms and causes were obtained by questionnaires, and no measurements (on e.g. MVOC levels) were performed. In general, when the concentration of a non-reactive MVOC exceeds a certain limit, it begins to evoke the odour sensation, and if the concentration is high enough, the symptoms of irritation appear. Available odour and irritation thresholds for some MVOCs are presented in Table 9.

Some MVOCs, like 3-methylfuran, 2-heptanone, 1-octen-3-ol, and 2-methylisoborneol, were observed to be related to asthma among subjects working in schools with elevated levels of airborne fungi (192). The authors point out,

Compound	Odour characterisation	Odour threshold (10, 47, 80, 88,	d 106, 178, 219)	(51) or	n threshold irritating tration (178)
		ppm	mg/m <sup>3</sup>	ppm	mg/m <sup>3</sup>
2-Methyl-1-propanol	Mild, musty, fungus-like	0.001-74	0.003-225	99	300
3-Methyl-1-butanol	Sour, sharp, malty	0.01-35	0.045-126	100	360
1-Octen-3-ol	Raw mushroom	-	-	-	-
2-Hexanone	Acetone-like	0.076	0.31		
2-Heptanone	Fungus-like, musty	0.02-0.35	0.094-1.6	281	1310
3-Octanone	Mild, fruity, fresh, herbal, lavender, sweet, fungus-like	6	31.2	50	260
2-Methylisoborneol	Musty, earthy	0.000001	0.000007	-	-
2-Isopropyl-3- methoxy-pyrazine	Musty, mouldy	0.0000002	0.000001	-	-
Geosmin	Musty, earthy	0.0009	0.0076	-	-
Dimethyl disulphide	Uncomfortable	0.00003-0.09	0.0001-0.3465	-	-

**Table 9.** Odour and sensory irritation thresholds/ irritating concentrations for some MVOCs most frequently searched for in problem buildings.

<sup>a</sup> nasal pungency threshold from human anosmics; available for 2-heptanone

however, that the occurrence of MVOCs is an indication of active growth of microorganisms and not a signal that MVOCs cause asthma.

There are few studies on MVOCs and irritation/inflammatory responses. In human experimental studies by Wålinder *et al*, some signs of inflammatory responses and respiratory reactions were reported after MVOC exposure (see also Chapter 11.2). Human subjects (n=30) were exposed to 10 mg/m<sup>3</sup> of 1-octen-3-ol for 2 hours. Increases in eye, nose and throat irritation, headache, dizziness, nausea, intoxication, blinking frequency, and in the amounts of lysozyme, myeloperoxidase and eosinophilic cationic protein in the nasal lavage fluid were reported (232). In another study, 29 healthy volunteers were randomly exposed to sham or  $1 \text{ mg/m}^3$  3-methylfuran for 2 hours. No subjective symptom ratings (related to smell, irritative and general symptoms) were increased during exposure. However, blinking frequency, tear film break-up time, and the lavage biomarkers myeloperoxidase and lysozyme were increased and forced vital capacity was decreased during exposure to 3-methylfuran compared to ambient air exposure (233). The exposure to 3-methylfuran caused an immediate obstructive (during the last 30 minutes of exposure) and late (three days after exposure) pulmonary reaction in a subject with previous occupational fungal exposure and presence of mould allergy (234). These experimental exposures were performed with 10 and 500 times higher concentrations of 1-octen-3-ol and 3-methylfuran, respectively, than measured in field samples (Table 5).

Koren et al reported a two-fold increase in PMNs in nasal lavage of 14 subjects immediately after a 4-hour exposure to a mixture of 22 VOCs at a total of 25  $mg/m^3$  (112), whereas Pappas *et al* (166) observed no significant increase in PMNs in 15 subjects after a for the most part compositionally similar exposure of 21 VOCs at 25 mg/m<sup>3</sup> and 50 mg/m<sup>3</sup>. However, increases in lower and upper respiratory symptoms were reported both immediately and two hours after exposure to VOCs at 50 mg/m<sup>3</sup> and increases in upper respiratory symptoms immediately after exposure to VOCs at 25 mg/m<sup>3</sup>. No changes were observed in lung function (forced expiratory flow in one second, forced vital capacity, forced expiratory flow between 25 and 75 % of forced vital capacity), cellularity or cell differentials, biomarkers of airway inflammation including interleukin-8, leukotriene  $B_4$ , or albumin in nasal lavage or induced sputum samples (166). The VOC mixtures of Koren et al and Pappas et al, respectively, were designed to mimic the levels and types of VOCs found in homes in the United States excluding suspected carcinogens and very irritating compounds. However, none of the 15 substances evaluated in the present document were included but e.g. p-xylene (8.25 mg/m<sup>3</sup>), 1-butanol, ethylbenzene, hexanal,  $\alpha$ -pinene (825  $\mu$ g/m<sup>3</sup> each), 2-butanone and 3methyl-2-butanone (75  $\mu$ g/m<sup>3</sup> each), which can be regarded as MVOCs.

In a study by Laumbach *et al*, 130 healthy women were exposed three times for 135 minutes each to clean air, or to a mixture of 23 VOCs at 25 mg/m<sup>3</sup> with and without the addition of 40 ppb ozone. The test mixture was claimed to contain some MVOCs but these were not identified nor were their individual concentrations specified. Laumbach *et al* reported no significant differences in nasal

irritation symptoms or nasal lavage PMNs between VOC + ozone, VOC alone, or clean air conditions, and concluded that MVOCs appear an unlikely cause of acute upper respiratory irritation or inflammation (129).

In a study reported by Claeson (48), 17 women and 10 men were exposed to low level emissions from 5 mould species grown on particle board and pine wood for 60 minutes. In a second exposure set-up, 13 women and 11 men with or without nose-clip were exposed to moderate level emissions from mouldy building materials for 10 minutes. The emissions included 1-butanol, 2-methyl-1-propanol, 3-methyl-1-butanol, 2-hexanone and 2-heptanone at individual MVOC concentrations in the range 0.56-3.4  $\mu$ g/m<sup>3</sup> and 13.2-214  $\mu$ g/m<sup>3</sup> representing low and medium (moderate) level exposures, respectively. Exposure to moderate MVOC levels in the condition without nose-clip increased the reports of perceived poor air quality (stuffy air, smell), and skin irritation. No such outcome was observed when participants were exposed to low MVOC levels or to moderate MVOC levels with nose-clip. Irrespective of exposure level or duration, no effects on the cornea reflected by self-reported tear film break-up time or on attention and processing speed were found (48).

Certain pathophysiological effects, such as the production of inflammatory mediators, cell influx into the airways, antibody responses and cell desquamation, have been more intensively studied after exposure to other compounds related to microbial exposure than the MVOCs. These microbial particulate or non-volatile fractions include e.g. fungal and actinomycete spores, cell wall components, proteins, glycoproteins, polysaccharides, mycotoxins, and bacterial-derived polypeptides and endotoxin. The immune responses may be noticed as the activation of PMNs and/or alveolar macrophages resulting in secretion of inflammatory mediators like cytokines (9, 156, 173).

#### Conclusion

Microbial VOCs have been related to complaints of general symptoms, as well as eye, nose, and throat irritation, and even asthma-like symptoms. However, there is a general lack of studies on dose-effect or dose-response relationships regarding irritation from single MVOC exposures and from MVOC mixtures. Human irritation thresholds, determined as nasal pungency thresholds by human anosmics or reported irritating concentrations, are available for 4 out of the 15 selected compounds and are considerably higher than measured indoor air levels for these compounds. Inflammatory responses have not been definitely confirmed although a few experimental studies indicate inflammatory responses and pulmonary reactions after exposure to 1-octen-3-ol and 3-methylfuran, respectively, at exposure levels 10 and 500 times higher than measured in field. The very few experimental studies on exposure to MVOC mixtures and inflammatory effects are inconclusive.

## 11.2 Effects of single and short-term exposure

Experimental single exposure studies have been conducted for 5 of the 15 substances selected for further investigation. In the text below focus is on

inhalation and the lowest administered doses. However, if inhalation data are lacking or scarce oral data have been included.

2-Methyl-1-propanol: Human toxicological data are virtually missing. In a drinking study in which 10 volunteers were given ethanol in orange juice with or without the addition of 2-methyl-1-propanol a clear increase in the frequency of errors and subjective hangover symptoms in the post-alcoholic phase was recorded with the addition of 2-methyl-1-propanol. No data on ingested amounts were given. In another report, it was stated that 2-methyl-propanol vapours cause narcosis and irritation of the upper airways. No further details were given (24).

*3-Methyl-1-butanol*: Throat irritation was reported in a respiratory uptake study in which 4 healthy, male volunteers were exposed through a mouthpiece to 25 ppm (90 mg/m<sup>3</sup>) for 10 minutes. In another study, slight throat irritation was reported in human volunteers after exposure to 100 ppm (360 mg/m<sup>3</sup>) 3-methyl-1-butanol for 3-5 minutes. Following exposure to 150 ppm (540 mg/m<sup>3</sup>) also eye and nose irritation were noted (82).

*1-Octen-3-ol*: Mucosal irritation and weak general symptoms were reported in human subjects (n=30) exposed to 10 mg/m<sup>3</sup> of 1-octen-3-ol for 2 hours. Subjective ratings of smell, eye, nose and throat irritation, dizziness, headache, intoxication and nausea were increased as were some of the results from objective measurements (blinking frequency and levels of the nasal lavage fluid biomarkers eosinophilic cationic protein, lysozyme, and myeloperoxidase) (232).

*3-Methylfuran:* Twenty-nine healthy volunteers were randomly exposed to sham or 1 mg/m<sup>3</sup> 3-methylfuran for 2 hours. No subjective symptom ratings (discomfort in the eyes, nose, and throat, dyspnoea, headache, fatigue, dizziness, nausea, and feeling of intoxication) were increased during exposure. However, blinking frequency, tear film break-up time, and the lavage biomarkers myeloperoxidase and lysozyme were increased and forced vital capacity was decreased during exposure to 3-methylfuran compared to sham exposure. In conclusion, the acute effects from eyes, nose, and airways indicate mucosal reactive properties of 3-methylfuran (233). One subject was removed from the study because of a two-phased pulmonary reaction. This suspected adverse reaction was described separately in a case-report. The subject suffered an acute obstructive reaction and a delayed pulmonary reaction with flu-like symptoms. Previous occupational exposure to fungi and presence of mould allergy may have contributed to the reaction (234).

2-*Hexanone*: Human subjects exposed to 1 000 ppm (4 100 mg/m<sup>3</sup>) of 2-hexanone for a few minutes reported transient moderate eye and nasal irritation (31).

In a toxicokinetic study where 3 healthy volunteers were exposed to 50 ppm  $(205 \text{ mg/m}^3)$  of 2-hexanone vapours for 7.5 hours or to 100 ppm  $(410 \text{ mg/m}^3)$  for 4 hours symptoms were not mentioned (31).

In conclusion, for all of the 15 selected substances data are either totally lacking or very scanty. In most of the reported studies doses are high when compared to actual MVOC levels in houses. However, acute effects from eyes, nose, and airways following exposure to 1 mg/m<sup>3</sup> of 3-methylfuran, and 10 mg/m<sup>3</sup> of 1-

octen-3-ol, respectively, have been reported. Still, such exposure levels are 10 and 500 times higher than levels reported indoors.

#### 11.3 Effects of long-term exposure

Relevant long-term studies were found for two of the 15 selected substances, all from work environments where the source of exposure was not microbial metabolism.

2-Hexanone: Occupational exposure to 2-hexanone, mostly as a paint thinner (usually at least 4 months), has resulted in changes in both motor and sensory nerves with symptoms such as muscular weakness and/or trembling of the extremities, and difficulty in walking and handling objects. Also weight reduction has been reported in many cases. Medical examination has shown a reduced nerve conduction velocity and electromyographic changes. Nerve biopsies have shown neurotoxic effects such as axonodal swelling and demyelination. Recovery after termination of exposure has been slow and not always complete. In most cases nerve function gradually improved but some cases even showed a slight worsening. In many cases exposure was both dermal and inhalatory.

In an epidemiological study, peripheral neuropathy was reported in workers in a coated fabrics plant. Four months before the onset of symptoms 2-hexanone had been introduced in the solvents in the printing department. Measurements indicated 2-hexanone levels in the range 1-156 ppm (4-640 mg/m<sup>3</sup>) and on average 9.2 ppm in front of the printers. However, the air samples were collected after the problem had arisen and only 9/17 printing machines were in operation during the 2-day sampling period. The true exposure levels could therefore not be estimated and data were insufficient to correlate 2-hexanone air levels with neurotoxic effects. In addition, percutaneous uptake could not be excluded, and also 2-butanone (methyl-ethyl ketone) was present in the area (highest measured concentration approximately 500 ppm) (31, 32, 138).

*Dimethyl disulphide:* Eighty-one pulp mill workers exposed to dimethyl disulphide (0-1.5 ppm, 0-5.8 mg/m<sup>3</sup>), dimethyl sulphide (0-14 ppm), hydrogen sulphide (0-6 ppm), and methyl mercaptan (0-15 ppm) complained of inability to concentrate, headaches, restlessness, and lack of vigour. However, only the increased frequency of headache reached statistical significance compared with controls (102).

The relationship between exposure to organic sulphides and disturbances in iron metabolism was investigated in 18 workers at a pulp and paper plant. Measured mean exposure levels were low, generally below the detection limits, i.e.<0.2 ppm for methylmercaptan, <0.05 ppm for dimethyl sulphide and <0.05 ppm (<0.19 mg/m<sup>3</sup>) for dimethyl disulphide. However, peak concentrations of one or two orders of magnitude higher were registered. Five subjects experiencing such peaks within 2 months before blood sampling had significantly elevated concentrations of serum iron and transferrin, and lower ferritin concentrations than referents. As an incidental finding, 6 workers not included in the study group involved in the

clean up following an explosion had significantly increased serum iron levels at 2 days compared to 10 days post-exposure (110).

In conclusion, long-term exposure studies are virtually missing, except for 2-hexanone, which is an established neurotoxicant. It cannot be excluded that peripheral neuropathy may develop in workers exposed to only a few mg/m<sup>3</sup> of 2-hexanone. For comparison, reported levels in non-industrial settings are <9  $\mu$ g/m<sup>3</sup>, i.e. about three orders of magnitude lower. A disturbed iron metabolism among workers exposed to organic sulphides has been reported. Levels of dimethyl disulphide were several orders of magnitude above those reported in indoor air.

## **11.4 Genotoxic effects**

No studies on genotoxic effects in humans were found for the 15 substances selected for further investigation.

#### 11.5 Carcinogenic effects

No studies assessing the carcinogenic potential in humans were found following exposure to any of the 15 substances selected for further investigation.

#### 11.6 Reproductive and developmental effects

No studies on reproductive and developmental effects in humans were found for the 15 substances selected for further investigation.

# 12. Dose-effect and dose-response relationships

Data on exposure levels and toxicological effects presented earlier in the document are summarised below. Data are given for both MVOC mixtures and 15 individual substances representing the MVOCs most often analysed and reported in previous MVOC studies.

#### 12.1 Animal data

Sensory irritation potency has been assessed for several MVOCs by determining the concentration causing a 50 % decrease in respiratory rate in mice ( $RD_{50}$ ) (see also 12.3). At low exposure levels to MVOC mixtures it has been assumed that sensory irritation effects are additive. If so, the total sensory potency of a specific MVOC mixture can be calculated. However, at higher exposure concentrations MVOCs may have synergistic effects on sensory irritation (156). It has been shown that the determined  $RD_{50}$  for an MVOC mixture was lower than estimated from the sum of fractional concentrations and the  $RD_{50}$ s of individual compounds (114). Irritation effects of short-term, repeated exposure (30 minutes/day for 4 days) to 3-octanone, 1-octen-3-ol or to a mixture of 2-methyl-1-propanol, 3-methyl-1-butanol, 1-octen-3-ol, 2-heptanone and 3-octanone by the mouse bioassay at levels ranging from <0.001 to  $0.2 \cdot \text{RD}_{50}$  seemed non-cumulative and transient (115). Levels were chosen to produce a clear respiratory rate decrease, and proportions of the MVOCs in the mixture to reflect those measured in mouldy buildings or mouldy building materials.

Based on acute toxicity studies on the 15 selected substances the alcohols, ketones and 3-methylfuran would be classified as slightly toxic. The database is, however, poor for most of the compounds. Virtually no data exist for 2-octen-1-ol, 2-pentanol, 3-octanone, geosmin, 2-methylisoborneol and 2-isopropyl-3-methoxy-pyrazine.

2-Hexanone is a well-established neurotoxicant. Neurotoxic effects, expressed as peripheral neuropathy, and weight loss/retarded weight gain have been observed in several species after repeated exposures and different modes of administration. The lowest inhalation exposure level reported to produce adverse effects in animals is 50 ppm (205 mg/m<sup>3</sup>). Although no effects were observed at an exposure of 40 ppm (164 mg/m<sup>3</sup>) for up to 88 days, it cannot be ruled out that signs of neuropathy may have developed later. The neurotoxicity of 2-hexanone is enhanced by co-exposure to other chemicals e.g. 2-butanone (methyl ethyl ketone), and 2-hexanone can also potentiate the toxicity of other compounds (31, 32, 138). Reported indoor air levels of 2-hexanone as an MVOC are <9  $\mu$ g/m<sup>3</sup>.

Regarding dimethyl disulphide, two inhalation studies of rats exposed 6 hours/ day, 5 days/week for 13 weeks have been performed according to current standards. Both studies indicate a NOEL below 10 ppm ( $38 \text{ mg/m}^3$ ) (13, 108). The highest reported concentrations of dimethyl disulphide in buildings are approximately 0.1  $\mu$ g/m<sup>3</sup>.

Moreover, the few available repeated exposure studies on 3-methyl-1-butanol, 2-methyl-1-propanol, 3-octanol and 2-heptanone suggest NOELs at least three orders of magnitude higher than the reported concentrations in problem or complaint buildings.

#### 12.2 Human data

Reported concentrations of single MVOCs in buildings with indoor air problems (microbial growth, odour complaints) range from a few ng/m<sup>3</sup> to  $<1 \text{ mg/m}^3$ . However, comparisons between studies are hampered by a lack of standardised and validated analytical methods for MVOCs. There is a considerable overlap of the results for both single and sums of several MVOCs in contaminated areas, clean areas and outdoor air.

There is a general lack of studies on dose-effect or dose-response relationships regarding irritation and other effects from exposure to individual MVOCs and MVOC mixtures. In epidemiological and case studies, MVOCs or musty, earthy odours have been related to complaints of general symptoms (i.e. headache, dizziness and fatigue) as well as eye, nose, and throat irritation, and even asthma-

like symptoms (63, 92, 111, 177, 204, 212). It has been suggested that odour complaints may be related to the occurrence of individual compounds or their combinations rather than the sum of selected MVOCs (92). However, if MVOCs are present in the air, other agents, e.g. fungal components will presumably be present as well.

Fifteen substances most often determined as MVOCs in field samplings were selected for further examination (Table 2). Available odour and irritation thresholds for those MVOCs are presented in Table 9. Odour thresholds may differ considerably both between substances and between studies of the same compound. Irritation thresholds/irritating concentrations are in the range 50-280 ppm, but are available only for 4 of the 15 selected compounds.

Inflammatory responses of single or repeated MVOC exposures have not been unequivocally confirmed in human experimental studies. The very few studies on MVOC mixtures are inconclusive. An increase in PMNs in nasal lavage were reported in a study on 14 subjects exposed to 22 VOCs at 25 mg/m<sup>3</sup> (112) whereas no such change was fond in two other studies on MVOC mixtures (129, 166). However, signs of inflammatory responses, and respiratory reactions were reported after single exposures to 1-octen-3-ol (10 mg/m<sup>3</sup>) (232), and 3-methylfuran (1 mg/m<sup>3</sup>) (233, 234), respectively. The experimental exposure levels in these studies were approximately 10 and 500 times higher than levels measured in field.

Dose-effect relationships after single and short-term exposure to VOCs, single or in combinations, are presented in Table 10. In general, the toxicological data on these compounds are poor. Virtually no data exist for 2-octen-1-ol, 2-pentanol, 3-octanone, geosmin, 2-methylisoborneol and 2-isopropyl-3-methoxy-pyrazine.

In an epidemiological study, peripheral neuropathy was reported in workers exposed to 2-hexanone. Measurements performed after the onset of problems indicated levels in the range 1-156 ppm (4-640 mg/m<sup>3</sup>) and also 2-butanone was present. Respiratory and percutaneous uptake could not be separated and precise exposure levels were not known, (31, 32, 138). It can therefore not be ruled out that severe neuropathy has developed at exposure concentrations down to one or a few ppm. Such 2-hexanone levels would still be more than 400 times higher than those encountered in indoor air.

3-Methyl-1-butanol       10 min       4 he.         25 ppm (90 mg/m <sup>3</sup> )       10 min       4 he.         100 ppm (360 mg/m <sup>3</sup> )       3-5 min       Ca 1         150 ppm (540 mg/m <sup>3</sup> )       3-5 min       Ca 1         150 ppm (540 mg/m <sup>3</sup> )       3-5 min       Ca 1         19 ppm (10 mg/m <sup>3</sup> )       2 h       30 h         3-Methylfuran       2 h       30 h         3-Methylfuran       2 h       1 vo         0.30 ppm (1 mg/m <sup>3</sup> )       2 h       1 vo	4 healthy, male volunteers Ca 10 male and female subjects Ca 10 male and female subjects	Throat irritation. Slight throat irritation.	
10 min 3) 3-5 min 3) 3-5 min 2 h 2 h	ealthy, male volunteers 10 male and female subjects 10 male and female subjects		
<ul> <li><sup>3</sup>) 3-5 min</li> <li><sup>3</sup>) 3-5 min</li> <li><sup>2</sup> h</li> <li><sup>2</sup> h</li> <li><sup>2</sup> h</li> </ul>	10 male and female subjects 10 male and female subjects		(82)
<sup>3</sup> ) 3-5 min 2 h 2 h	10 male and female subjects		(82)
2 h 2 h		Throat, eye and nose irritation.	(82)
2 h 2 h			
2 ћ	30 healthy subjects serving as their own controls	Increases in eye, nose, throat irritation, headache, dizziness, nausea, (intoxication, blinking rate, and in the amounts of lysozyme, myeloperoxidase and eosinophilic cationic protein in the nasal lavage fluid.	(232)
2 h			
ATT	<ol> <li>volunteer with previous occupational fungal exposure and presence of mould allergy</li> </ol>	An immediate (during the last 30 min of exposure) obstructive reaction and a (delayed pulmonary reaction with flu-like symptoms.	(234)
0.30 ppm (1 mg/m <sup>3</sup> ) 2 h 29 h as th	29 healthy volunteers serving as their own controls	No increase in subjective symptom ratings (discomfort in eyes, nose, and throat, dyspnoea, headache, fatigue, dizziness, nausea, and feeling of intoxication). However, blinking frequency, tear film break-up time and the nasal lavage biomarkers myeloperoxidase and lysozyme were increased and forced vital capacity was decreased.	(233)
2-Hexanone			
50 ppm (205 mg/m <sup>3</sup> ) 7.5 h 3 he	3 healthy volunteers	Symptoms not mentioned.	(31)
$100 \text{ ppm} (410 \text{ mg/m}^3)$ $4 \text{ h}$ 3 he	3 healthy volunteers	Symptoms not mentioned.	(31)

**Table 10.** Dose-effect relationships in man after single or short-term inhalation exposure to VOCs. single or in combination.

Compound/Exposure level	Duration	Duration No. of subjects	Effects Re	Reference
VOC mixtures, total concentrations				
$22 \text{ VOCs at } 25 \text{ mg/m}^3$	4 h	14 subjects	A two-fold increase in PMNs in nasal lavage immediately after exposure.	(112)
21  VOCs $25 \text{ mg/m}^3 \text{ and } 50 \text{ mg/m}^3$	4 h	15 subjects	No significant difference in nasal lavage cellularity or differential cell counts. Increases in lower and upper respiratory symptoms both immediately and two hours after exposure to 50 mg/m <sup>3</sup> . Increases in upper respiratory symptoms immediately after exposure to 25 mg/m <sup>3</sup> . No changes in lung function (FEV <sub>1</sub> , FVC, or FEF <sub>25.75</sub> ), cellularity or cell differentials, biomarkers of airway inflammation including interleukin-8, leukotriene B <sub>4</sub> , or albumin in nasal lavage or induced sputum samples.	(166)
23 VOCs (not specified) at 0, 25 and 25 mg/m <sup>3</sup> +40 ppb ozone, respectively	3·135 min	130 healthy women	No significant differences in nasal irritation symptoms or nasal lavage PMNs between VOC + ozone, VOC alone, or clean air conditions.	(129)
Emissions from 5 mould species grown on particle board and pine wood: Emissions included 1-butanol, 2-methyl-1-propanol, 3-methyl-1-butanol, 2-hexanone and 2-heptanone at individual MVOC concentrations in the range 0.56-3.4 $\mu$ g/m <sup>3</sup>	60 min	17 women and 10 men	No effects on perceived air quality or skin symptoms. No effects on the cornea reflected by self-reported <sup>a</sup> tear film break-up time or on attention and processing speed.	(48)
Emissions as above, at individual MVOC concentrations in the range 13.2-214 $\mu$ g/m <sup>3</sup>	10 min	13 women and 11 men with or without nose- clip	In exposure without nose-clip, increased ratings of perceived poor air quality (stuffy air, smell) and increased reporting of skin irritation. No effects on the self-reported <sup>a</sup> corneal tear film break-up time or on attention and processing speed.	(48)

Table 10. Dose-effect relationshins in man after single or short-term inhalation exposure to VOCs single or in combination

FUC = forced expiratory volume in one second FVC = forced vital capacity FEF<sub>2575</sub> = forced expiratory flow between 25 and 75 % of FVC PMN = polymorphonuclear neutrophil

#### 12.3 Extrapolation of animal data on sensory irritation responses to humans

At the levels relevant in this context, the effects from MVOC exposure are those related to irritation in the eyes and upper airways. At higher exposure levels MVOCs may cause also other effects. Thus, the evaluation of human health risks caused by MVOC exposure focuses on the sensory irritation. Sensory irritation potency is assessed by determining the  $RD_{50}$  in mice (Chapter 10.1). However, the question of the application of  $RD_{50}$ s to human responses is a critical issue. Cometto-Muñiz and Cain showed that the sensory irritation potencies of 21 VOCs (consisting of e.g. alcohols, acetates, ketones, alkylbenzenes) estimated in the mouse bioassay were well correlated (r = 0.85) with the human potencies, measured as the nasal pungency thresholds (51). The extrapolation of the mouse bioassay to human exposures predicted that, in general, slight but tolerable irritation would occur at  $0.1 \cdot RD_{50}$  and minimal or no effect at  $0.01 \cdot RD_{50}$  (101). Further, regression analyses suggest that RD<sub>50</sub>s correlate with the threshold limit values (TLVs) elaborated by American Conference of Governmental Industrial Hygienists (ACGIH) (r = 0.88) with a slope factor (regression coefficient) of 0.03 (the mid-point between 0.1 and 0.01 on a logarithmic scale) (180). As a pragmatic approach, such levels  $(0.03 \cdot RD_{50})$  could constitute an acceptable level of human exposure to prevent sensory irritation in work environments. The RD<sub>50</sub>s and corresponding 0.03 RD<sub>50</sub>s for the selected MVOCs are presented in Table 11. For the tabulated MVOCs the  $0.03 \cdot \text{RD}_{50}$ s are in the range 5-530 mg/m<sup>3</sup> (approximately 0.9-100 ppm).

The levels of the compounds usually measured in water damaged and/or mould problem buildings (Table 5), are generally several orders of magnitude lower than the  $0.03 \cdot \text{RD}_{50}$ s given in Table 11. For such low-concentration mixtures of MVOCs, it seems prudent to apply the additivity rule (7). The underlying assumption is that all the compounds act on the same biological site and individual compounds act as

Compound	RD <sub>50</sub> (mg/m <sup>3</sup> )	0.03 RD <sub>50</sub> (mg/m <sup>3</sup> )	
2-Methyl-1-propanol	5 499	165	
3-Methyl-1-butanol	9 325	280	
3-Methyl-2-butanol	9 645	289	
2-Pentanol	9 907	297	
3-Octanol	1 359	41	
1-Octen-3-ol	182	5.5	
2-Hexanone	10 449	313ª	
2-Heptanone	4 163	125	
3-Octanone	17 586	528	
2-Methylisoborneol	811	24	
Geosmin	216	6.5	
Acrolein	4	0.12	
Formaldehyde	5	0.15	

**Table 11.** The RD<sub>50</sub>s (114, 168, 180) and  $0.03 \cdot \text{RD}_{50}$ s for some MVOCs. The values for acrolein and formaldehyde were added for comparison.

<sup>a</sup> Neurotoxicity appears at lower exposure levels (Chapters 10.3 and 11.3).

dilutions (depending on their relative potencies) of the same toxic compound (122). For the occupational setting the additive effect could be calculated as follows:

Additive effect =  $\sum (c_n / 0.03 \cdot RD_{50n})$ , where

c= measured concentration of a chemical

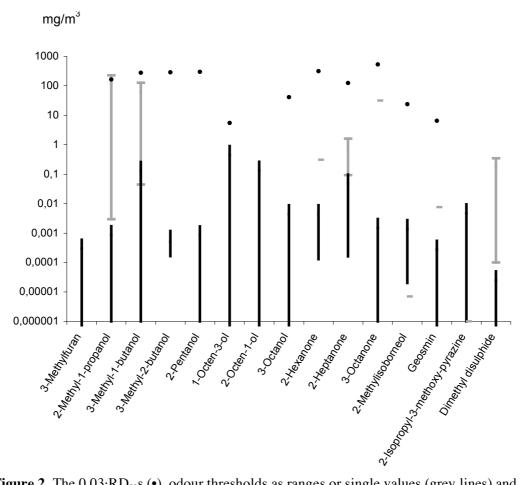
If the additive effect exceeds the value 1, sensory irritation may be expected, whereas levels below 1 should be of no concern (7).

The interest in the indoor environment is to protect the general, rather than the working, population. This includes sensitive individuals (e.g. asthmatics and children) and continuous exposure for 24 hours per day. On this basis, Nielsen *et al* proposed that the  $0.03 \cdot \text{RD}_{50}$  should be divided by 40 when calculating a recommended indoor air level (RIL) for individual non-reactive VOCs outside the occupational settings. The factor 40 includes a four times longer duration of indoor air exposure compared to occupational exposure and a safety factor of 10 for potential risk groups (156). The assumption of additivity and, thus the same procedure for calculating the additive effect, applies also for RILs.

However, it should be pointed out that the approach to calculate "acceptable" levels is applicable only for sensory irritation effects. For reactive substances or substances with effects other than sensory irritation as the primary concern, other extrapolations to protect humans should be applied.

Pasanen *et al* calculated RILs for 27 MVOCs in a theoretical setting and for 3-14 MVOCs with reported concentrations in some problem buildings. Individual RILs for single MVOCs approach hundreds (e.g. 1-octen-3-ol, 2-methylisoborneol, geosmin) or thousands of  $\mu$ g/m<sup>3</sup> (e.g. 2-methyl-1-butanol, 3-methyl-1-butanol, 3-methyl-2-butanol, 2-pentanol, 3-octanol, 2-hexanone, 2-heptanone) (168). Such high MVOC concentrations rarely occur in regular indoor environments. Laumbach *et al* and Sigsgaard and Bornehag have recently reached the same conclusion (129, 189). Similarly, Böck concluded, based on literature data, that the indoor concentrations of single MVOCs are 4-6 orders below their RD<sub>50</sub>s (35).

Both in theoretical calculations and based on MVOC concentrations measured in some problem buildings, when assuming additivity the total effect has remained below unity even when using RILs instead of  $0.03 \cdot \text{RD}_{50}$ s as acceptable limits (see the above mentioned formula), indicating that irritation symptoms due to MVOCs should not be expected. These authors also estimated that microbial growth seems to have only marginal effects on the total VOC load in the room (168). Odour thresholds, indoor air concentrations, and the  $0.03 \cdot \text{RD}_{50}$ s are combined and presented in Figure 2.



**Figure 2.** The  $0.03 \cdot \text{RD}_{50}$ s (•), odour thresholds as ranges or single values (grey lines) and indoor air concentrations (black lines) of selected MVOCs, when available. The indoor air concentrations have been measured in residences or in non-industrial work sites such as schools.

# 13. Previous evaluations by national and international bodies

No evaluation related to health risks of MVOCs in general was found in the literature. Considering the 15 selected compounds, health risks have been evaluated for five of them, as presented in Table 12. However, it should be noted that the purpose of these evaluations has been to estimate the health risks in industrial work environments and processes where workers are exposed to much higher concentrations of one or a few of these chemicals. This contrasts with exposure to chemicals of microbial origin (e.g. in buildings with moisture and microbial damage) where people are exposed to a wide range of MVOCs albeit at much lower concentrations.

Compound/Organisation (Year)	Summary of conclusion/assessment	Reference
2-Methyl-1-propanol		
Swedish Criteria Group for Occupational Standards (SCG) (1984)	Reported effects of long-term exposure are primarily irritation of eyes and mucous membranes and some dizziness. In experimental animals central nervous system effects have been shown after relatively high doses.	(136)
International Programme on Chemical Safety (IPCS), World Health Organization (WHO) (1987)	The available data are inadequate to set an OEL. In line with good manufacturing practice, exposure to isobutanol should be minimised. Isobutanol is severely irritating to the eyes and moderately irritating to the skin. From the animal studies available, it is not possible to determine a no-observed-adverse-effect-level for long-term exposure. No adequate data are available to assess mutagenicity or teratogenicity of isobutanol or effects on reproduction.	(06)
American Conference of Governmental Industrial Hygienists (ACGIH) (2001)	A TLV-TWA of 50 ppm is recommended for occupational exposure to isobutanol to minimise the potential for skin and ocular irritation. Sufficient data were not available to recommend skin, sensitiser or carcinogenicity notations or a STEL.	(2)
3-Methyl-1-butanol		
American Conference of Governmental Industrial Hygienists (ACGIH) (2001)	A TLV-TWA of 100 ppm and a TLV-STEL of 125 ppm are recommended for occupational exposure to isoamyl alcohol, in part by analogy with the irritation data for $n$ -butanol. This value is intended to minimise the potential for upper respiratory tract, and ocular irritation, with possible corneal damage. Sufficient data were not available to recommend skin, sensitiser or carcinogenicity notations.	(2)
2-Hexanone		
German Research Foundation (DFG) (1975)	Concentrations down to 6 ppm lead to occurrence of polyneuritis in chronically exposed humans. Long- term studies in animals have also shown neurotoxic effects at 100-200 ppm. The MAK value is therefore established at 5 ppm.	(59)

Compound/Organisation (Year)	Summary of conclusion/assessment	Reference
US National Institute for Occupational Safety and Health (NIOSH) (1978)	Studies on a variety of animals have conclusively demonstrated that repeated exposure to methyl $n$ -butyl ketone produced peripheral neuropathy and data indicated that the no effect concentration in animals was probably less than 100 ppm. Human data indicate that apparently 2.3 ppm cannot be ruled out as causing neuropathy. Because of the severity of the toxic effects and the incomplete reversibility of the lesions in workers a cautious approach is needed. The ketone has the ability to penetrate skin as well as to cause local skin effects. It is recommended that methyl $n$ -butyl ketone concentrations in workplace air not exceed 1 ppm (10-hour TWA).	(161)
Dutch Expert Committee on Occupational Standards (DECOS) and the Swedish Criteria Group for Occupational Standards (SCG) (1990)	The primary target organ for 2-hexanone is the nervous system. It cannot be ruled out that severe neuro- toxic effects that are not always completely reversible may develop in man at exposure levels as low as 2 ppm. Percutaneous absorption may contribute significantly to the occupational 2-hexanone exposure. Attention should be paid to the potentiation of 2-hexanone neurotoxicity by other chemicals. DECOS recommends (55) a health-based OEL for 2-hexanone of 0.5 ppm as an 8-hour TWA concentration.	(31, 55)
Swedish Criteria Group for Occupational Standards (SCG) (1992)	The critical effect of occupational exposure to 2-hexanone is its effect on the nervous system. It should be noted that 2-hexanone is readily absorbed by the skin.	(138)
Agency for Toxic Substances and Disease Registry (ATSDR) (1992)	The most important health effect from exposure to 2-hexanone is its harmful effect on the nervous system. These effects were seen in workers who were exposed to 2-hexanone for almost a year. The major effects were weakness, numbness, and tingling in the skin of the hands and feet. Similar effects were seen in animals that ate or breathed high levels of 2-hexanone, these effects included weakness, clumsiness, and paralysis.	(17)
American Conference of Governmental Industrial Hygienists (ACGIH) (2001)	A TLV-TWA of 5 ppm and a STEL of 10 ppm are recommended. These values are intended to minimise the potential for distal peripheral neuropathy primarily nerve fibre conduction, with weakness in the hands and feet and loss of coordination. A STEL is recommended to control exposure concentrations, which have the potential to induce testicular toxicity. A skin notation is assigned based on data reporting skin uptake in humans contributing substantially to the total body burden. Sufficient data were not available to recommend sensitiser or carcinogenicity notations.	(2)

Compound/Organisation (Year)	Summary of conclusion/assessment	Reference
2-Heptanone		
Dutch Expert Committee on Occupational Standards (DECOS) and the Nordic Expert Group for Documentation of Occupational Exposure Limits (NEG) (1990)	The target organs for exposure to 2-heptanone are the upper respiratory tract for its irritation properties, the central nervous system, the liver and kidneys. Based on animal inhalation data 1 000 ppm was considered an overall no adverse effect level. DECOS therefore recommended (56) a health-based OEL of 50 ppm as an 8-hour TWA concentration.	(56, 218)
Swedish Criteria Group for Occupational Standards (SCG) (1992)	Judging from animal data, the critical effect of 2-heptanone is irritation of the upper respiratory tract.	(139)
American Conference of Governmental Industrial Hygienists (ACGIH) (2001)	Although based on limited toxicity data a TLV-TWA of 50 ppm is recommended to minimise the potential for eye and skin irritation. The lack of objective signs of toxicity, including neurotoxicity, in rats and monkeys inhaling 131 ppm of 2-heptanone on a daily basis for 9 months provide the basis for the recommended TLV.	(2)
Dimethyl disulphide		
Swedish Criteria Group for Occupational Standards (SCG) (1987)	There are no data on which to base dose-response or dose-effect relationships for occupational exposure. The critical effect of dimethyl disulphide is the discomfort caused by the smell.	(137)
MAK = Maximale Arbeitsplatz-Konzentration (maximum workplace concentration) OEL = occupational exposure limit STEL = short-term exposure limit TLV® = threshold limit value TWA = time weighted average	n (maximum workplace concentration)	

# 14. Evaluation of human health risks

## 14.1 Assessment of health risks

It is difficult to evaluate human health risks because of the lack of sufficient knowledge of the specific MVOCs, exposure to MVOCs (in particular mixtures) especially in work environments, and of the mechanism of possible health effects of MVOCs. Furthermore, if there are MVOCs in the air there will most certainly be other agents, e.g. fungal components, present as well. The toxicological database is poor, at least for the 15 typically analysed MVOCs. Considering typical MVOCs - as reported qualitatively and quantitatively in the field so far - it appears evident that eye and upper respiratory tract (sensory) irritation is the most probable response to MVOC exposure, and no long-term or more toxic effects are expected. Inflammatory responses (e.g. increase in the count of inflammatory cells or other mediators) after single or repeated MVOC exposures have not been unequivocally confirmed in controlled human exposure studies. Thus, sensory irritation by the stimulation of the trigeminal nerves seems to be the most likely mechanism of toxicity of typical MVOCs. Occupational exposure limits (OELs) based on irritation as the critical effect have been shown to correlate roughly to 3 % of their respective RD<sub>50</sub>s. This would correspond to 5-530 mg/m<sup>3</sup> (0.9-100 ppm) for the 15 typical MVOCs, with geosmin and 1-octen-3-ol in the lower, and 3-octanone in the higher range. A further reduction by a factor of 40 has been proposed to protect the general population, including sensitive groups, which would correspond to  $0.1-13 \text{ mg/m}^3$  for the individual MVOCs evaluated in this document.

#### 14.2 Groups at extra risk

In the light of present knowledge, no long-term health effects are expected as a result of exposure to concentrations of MVOCs measured in buildings with moisture and microbial damage. There are no scientific data available suggesting that any particular group is at extra risk.

#### 14.3 Scientific basis for an occupational exposure limit

There are insufficient data to serve as basis to establish OELs for MVOC mixtures. There is no clear definition, but several hundred substances may be considered as MVOCs. With a few exceptions, little is known about the concentrations of these substances in indoor air and even less is known about their health hazards. Measurements in indoor air have generally focused on a relatively small number of substances.

It should be pointed out, however, that some of the substances considered as MVOCs are also used in industry and/or occur at relatively high levels in the work environment. For these substances more is known about exposure levels in work places and health effects and some have established OELs.

Sensory irritation seems to be the critical effect for many of the individual substances. Considering the low levels generally occurring in indoor air, it seems prudent to apply the additivity rule to calculate the risk for sensory irritation. Such calculations based on the  $RD_{50}$ s determined in mice suggest that the MVOC concentrations measured so far in indoor environments are well below the levels expected to cause sensory irritation. However, such exercises must be executed and interpreted with care and cannot be applied to reactive substances, and substances with other endpoints than irritation as the major concern (2-hexanone).

# 15. Research needs

In the past, MVOCs have been a focus of research in two ways: as indicators of microbial growth in a substrate (foodstuffs, building constructions) or as possible causative agents for adverse health effects in buildings with moisture and microbial damage. However, among the compounds identified so far, none has been verified as a "pure", MVOC, i.e. of solely microbial origin. Possible candidate groups for such specific MVOCs could include sesquiterpenes, furans, and very volatile compounds. The search requires further development of analytical methods. The other approach to increase the reliable interpretation of MVOC results might be to focus on statistical data handling of chromatograms. Some attempts on application of the principal component analyses have been made, but this area needs more research. On the other hand, statistical analyses require large databases on MVOC concentrations (exposure data) in different environments and occasions collected with the identical methodology. Before databases can be gathered, consensus on recommended sampling and analytical methods should be reached among researchers. These measures are necessary, if MVOC analysis is intended to be utilised further either in the research or in field settings, although according to the present knowledge, the use of MVOC concentrations for both the above mentioned purposes appears to be questionable.

Considering health effects of MVOCs, more *in vitro* and *in vivo* data on the inflammatory and other immunological responses to MVOCs are needed. Furthermore, information on co-effects of several microbiological agents is still missing, though it is evident that no single agent studied so far is responsible for the health effects observed in subjects exposed to microorganisms in living and work environments. One possibility is to direct the measurements towards more reactive compounds (such as amines, acids), as they are more likely to affect human health.

# 16. Summary

Pasanen A-L, Järnberg J, Korpi A. *The Nordic Expert Group for Criteria* Documentation of Health Risks from Chemicals. 138. Microbial volatile organic compounds (MVOCs). Arbete och Hälsa 2006;13:1-78.

Microbial volatile organic compounds (MVOCs) consist of a variety of compounds (mainly alcohols, ketones, terpenes, esters, aldehydes, sulphur and nitrogen compounds) formed as side-products in the primary and secondary metabolism of fungi and bacteria. More than 200 compounds have been identified as MVOCs in laboratory experiments, but none can be regarded as exclusively of microbial origin or specific for certain microbial species. Thus, these compounds also have many other, often much stronger, sources than microbial metabolism in the environment. Furthermore, the techniques chosen for sampling and analysis will effect which MVOCs are detected. In many cases, the typcially 10-15 compounds analysed have been selected beforehand based on previous investigations as well as existing analytical facilities.

In this review, the basic physical and chemical properties of 96 typical MVOCs have been summarised. Of these, the 15 MVOCs listed below have most often been analysed and reported in buildings with moisture and microbial damage and sometimes in occupational settings (in agriculture or compost facilities). These were also the MVOCs for which the toxicological and exposure data were gathered in this report:

2-Methyl-1-propanol	1-Octen-3-ol	3-Octanone
3-Methyl-1-butanol	2-Octen-1-ol	2-Methylisoborneol
3-Methyl-2-butanol	3-Methylfuran	2-Isopropyl-3-methoxy-pyrazine
2-Pentanol	2-Hexanone	Geosmin
3-Octanol	2-Heptanone	Dimethyl disulphide

In studies with quantitative MVOC results, single MVOC levels have ranged from a few ng/m<sup>3</sup> up to 1 mg/m<sup>3</sup> both in indoor and work environments, however, quantitative data from work settings are limited to compost facilities. The comparable data on MVOC levels in the environment published so far are inadequate for drawing reliable conclusions on the MVOC exposure.

The main exposure route for MVOCs is absorption through the lungs. Typical MVOCs are rapidly metabolised and excreted in the urine and bile. Generally, MVOCs do not accumulate in tissues to any great extent.

The toxicological database is poor for the 15 listed MVOCs. In epidemiological studies on buildings with moisture and microbial damage, MVOCs, in addition to many other microbial agents, have been associated with unpleasant odours, eye and upper airway irritation, unspecific symptoms and even asthma-like symptoms. However, inflammatory responses of single or repeated MVOC exposures have not been unequivocally confirmed in controlled human exposure studies. The most obvious health effect of MVOC exposure is eye and upper airway irritation,

due to stimulation of the trigeminal nerves (sensory irritation), which seems to be the critical effect also for many individual MVOCs although there are exceptions (2-hexanone). In human experimental exposure studies, symptoms of irritation have appeared at MVOC concentrations several orders of magnitude higher than those measured under field conditions in indoor environments. This is supported by data from animal studies. According to the database determined by the ASTM mouse bioassay, dose-dependent sensory irritation has been detected for many typical MVOCs. Assuming additivity of the sensory irritation reaction, the irritation responses anticipated from exposure to MVOC mixtures lead to the conclusion that MVOCs - as combinations and concentrations reported so far - are well below the levels needed to cause sensory irritation.

Overall, considering the very low levels encountered in the MVOC context, no toxic effects besides irritation, and very seldom also this effect, are expected. On the other hand, the present document covers the toxicological data of only 15 out of more than 200 MVOCs recognised so far. Thus, the conclusions do not necessarily apply to all MVOCs, and not even for all of the 15 compounds evaluated, as there may be more potent compounds and/or other endpoints not yet evaluated.

So far, attempts to recognise microbially contaminated buildings or areas, or to verify the success of remedial measures by MVOC measurements have failed because of considerable overlap of the results (both for individual MVOCs and sums of several MVOCs) between suspected and control areas. Thus, in order to identify contaminated buildings by MVOC measurements, MVOCs of purely microbial origin and/or sophisticated data handling procedures are needed.

*Keywords:* health effect, microbial volatile organic compound, MVOC, occupational exposure limit, respiratory effects, review, risk assessment, sensory irritation, toxicity

# 17. Summary in Swedish

Pasanen A-L, Järnberg J, Korpi A. *The Nordic Expert Group for Criteria* Documentation of Health Risks from Chemicals. 138. Microbial volatile organic compounds (MVOCs). Arbete och Hälsa 2006;13:1-78.

Mikrobiella flyktiga organiska ämnen (MVOC) består av en mängd olika ämnen (huvudsakligen alkoholer, ketoner, terpener, estrar, aldehyder, svavel- och kväveföreningar) som bildas som biprodukter vid svampars och bakteriers primära och sekundära metabolism. Över 200 ämnen har identifierats som MVOC vid laboratorieförsök, men inga av dessa kan anses ha enbart mikrobiellt ursprung eller vara specifika för en speciell mikroorganism eftersom MVOC har andra, ofta mycket mer betydelsefulla källor i miljön. Även provtagnings- och analysmetodiken har betydelse för vilka ämnen som detekteras. I många fall har antalet ämnen som analyseras begränsats till 10-15 st vilka valts på förhand utifrån tidigare laboratorieeller fältförsök och på tillgänglig analysutrustning.

I det här kriteriedokumentet sammanfattades fysikaliska och kemiska egenskaper för 96 typiska MVOC. Av dessa har följande 15 ämnen oftast analyserats och rapporterats förekomma i byggnader med fukt- och mikrobiella skador samt ibland i arbetsmiljöer (inom jordbruk och kompostering). För dessa ämnen ges exponeringsdata och en toxikologisk översikt.

2-Metyl-1-propanol	2-Okten-1-ol	3-Oktanon
3-Metyl-1-butanol	3-Oktanol	2-Metylisoborneol
3-Metyl-2-butanol	3-Metylfuran	2-Isopropyl-3-metoxi-pyrazin
2-Pentanol	2-Hexanon	Geosmin
1-Okten-3-ol	2-Heptanon	Dimetyldisulfide

I studier med kvantitativa uppgifter om MVOC har nivåerna av enskilda ämnen varierat mellan några få ng/m<sup>3</sup> upp till 1 mg/m<sup>3</sup> både inomhus och i arbetsmiljöer, även om kvantitativa data från arbetsplatser är sällsynta. Hittills publicerade och jämförbara data på MVOC-nivåer i miljön är för bristfälliga för att man ska kunna dra några långtgående slutsatser om MVOC-exponering.

Upptag via lungorna är den huvudsakliga exponeringsvägen för MVOC. Typiska MVOC metaboliseras snabbt och utsöndras i urinen och gallan. Generellt ackumuleras inte MVOC i vävnader och organ i någon större utsträckning.

Den toxikologiska databasen är mager för de 15 listade MVOC. I epidemiologiska studier av byggnader med fukt- och mikrobiella skador har obehaglig lukt, ögon- och övre luftvägsirritation samt ospecifika och till och med astmaliknande symtom satts i samband med MVOC och även många andra mikrobiologiska agens. Det har emellertid inte otvetydigt påvisats något inflammatoriskt svar efter enstaka eller upprepade MVOC-exponeringar i kontrollerade humanstudier. Den mest uppenbara effekten av exponering för MVOC är ögon- och övre luftvägsirritation via stimulering av trigeminalnerven (sensorisk irritation), vilket tycks vara den kritiska effekten även för många enskilda MVOC även om det finns undantag (2-hexanon). I exponeringsstudier på människa har irritationssymtom uppkommit vid MVOC-nivåer som är flera tiopotenser högre än de man mätt i inomhusmiljöer. Detta stöds av data från djurstudier. Enligt den databas som bygger på resultat från en djurmodell (ASTM mouse bioassay) har dosberoende sensorisk irritation detekterats för många typiska MVOC. Förutsatt att additivitet gäller för sensorisk irritation av MVOC blir dock slutsatsen att de kombinationer och koncentrationer av MVOC som hittills rapporterats inte ger upphov till denna effekt.

Med tanke på de mycket låga nivåer som förekommer i MVOC-sammanhang förväntas inga andra toxiska effekter förutom irritation och således mycket sällan också det. Detta dokument täcker emellertid toxikologiska data för endast 15 av mer än 200 ämnen som anses vara MVOC. Slutsatserna gäller således inte nödvändigtvis alla MVOC, och kanske inte ens de 15 som utvärderats, eftersom det kan finnas mer potenta ämnen och/eller andra effekter som ännu inte utvärderats.

Ansträngningar att identifiera mikrobiologiskt kontaminerade byggnader eller områden, eller att verifiera effekten av åtgärder genom MVOC-mätningar, har hittills misslyckats på grund av betydande överlappning mellan misstänkta områden och kontrollområden, både vad gäller nivåer av enskilda MVOC och summan av flera MVOC. För att skadade byggnader ska kunna identifieras med hjälp av MVOC-mätningar behövs MVOC med enbart mikrobiellt ursprung och/eller avancerade databehandlingsmetoder.

*Nyckelord:* hygieniskt gränsvärde, hälsoeffekter, mikrobiella flyktiga organiska ämnen, MVOC, respiratoriska effekter, riskbedömning, sensorisk irritation, toxicitet, översikt

# 18. References

- 1. Abraham MH, Nielsen GD, Alarie Y. The Ferguson principle and an analysis of biological activity of gases and vapors. *J Pharm Sci* 1994;83:680-688.
- 2. ACGIH. *Documentation of the threshold limit values and biological exposure indices.* 7th ed. Cincinnati, OH: American Conference of Governmental Industrial Hygienists Inc., 2001.
- ACGIH. 2006 TLVs and BEIs. Based on the Documentation of the threshold limit values for chemical substances and physical agents & biological exposure indices. Cincinnati, OH: American Conference of Governmental Industrial Hygienists Inc., 2006.
- 4. Alarie Y. Irritating properties of airborne materials to the upper respiratory tract. *Arch Environ Health* 1966;13:433-449.
- 5. Alarie Y. Sensory irritation by airborne chemicals. CRC Crit Rev Toxicol 1973;2:299-363.
- 6. Alarie Y, Nielsen GD, Andonian-Haftvan J, Abraham MH. Physicochemical properties of nonreactive volatile organic chemicals to estimate RD50: alternatives to animal studies. *Toxicol Appl Pharmacol* 1995;134:92-99.
- Alarie Y, Schaper M, Nielsen GD, Abraham MH. Estimating the sensory irritating potency of airborne nonreactive volatile organic chemicals and their mixtures. SAR QSAR Environ Res 1996;5:151-165.
- 8. Alarie Y, Schaper M, Nielsen GD, Abraham MH. Structure-activity relationships of volatile organic chemicals as sensory irritants. *Arch Toxicol* 1998;72:125-140.
- Ammann HM. Mold toxicity: risk assessment for humans exposed indoors. In: Johanning, ed. Proceedings of the 5th international conference: Bioaerosols, fungi, bacteria, mycotoxins and human health. Fungal Research Group Foundation, Inc. Albany, NY: Boyd Publishing, 2005:52-59.
- 10. Amoore JE, Hautala E. Odor as an aid to chemical safety: odor thresholds compared with threshold limit values and volatilities for 214 industrial chemicals in air and water dilution. *J Appl Toxicol* 1983;3:272-290.
- Andrews LS, Snyder R. Toxic effects of solvents and vapors. In: Amdur MO, Doull J, Klaasen CD, eds. *Casarett and Doull's Toxicology. The basic science of poisons.* 4th ed. New York: Pergamon Press, 1991:681-722.
- 12. Anger WK, Jordan MK, Lynch DW. Effects of inhalation exposures and intraperitoneal injections of methyl n-amyl ketone on multiple fixed-ratio, fixed-interval response rates in rats. *Toxicol Appl Pharmacol* 1979;49:407-416.
- 13. Arkema Inc. *Dimethyl disulfide (CAS# 624-92-0). Test plan.* US Environmental Protection Agency High production volume (HPV) challenge program, 2005.
- 14. ASTM. Standard test method for estimating sensory irritancy of airborne chemicals. Designation: E981-84. Philadelphia, PA: American Society for Testing and Materials, 1984.
- 15. Atkinson R, Tuazon EC, Aschmann SM. Atmospheric chemistry of 2-pentanone and 2-heptanone. *Environ Sci Technol* 2000;34:623-631.
- ATSDR. Toxicological profile for 2-butanone. http://www.atsdr.cdc.gov/toxprofiles/tp29.html (October 25, 2006). Agency for Toxic Substances and Disease Registry. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service, 1992.
- ATSDR. Toxicological profile for 2-hexanone. http://www.atsdr.cdc.gov/toxprofiles/tp44.html (May 29, 2006). Agency for Toxic Substances and Disease Registry. Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service, 1992.

- 18. Barr JG. Effects of volatile bacterial metabolites on the growth, sporulation and mycotoxin production of fungi. *J Sci Food Agric* 1976;27:324-330.
- 19. Batterman SA. Sampling and analysis of biological volatile organic compounds. In: Burge HA, ed. *Bioaerosols*. Boca Raton, FL: CRC Press Inc., 1995:249-268.
- Bayer CW, Crow S. Odorous volatile emissions from fungal contamination. In: Teichman KY, ed. Proceedings from IAQ'93: Operating and maintaining buildings for health, comfort and productivity. Atlanta, GA: American Society of Heating, Refrigerating and Air-Conditioning Engineers, Inc., 1994:165-170.
- 21. Bentley R, Bennett JW. Biosynthesis of secondary metabolites. In: Berry DR, ed. *Physiology* of industrial fungi. Oxford: Blackwell Scientific Publications, 1988:161-183.
- 22. Bentley R, Meganathan R. Geosmin and methylisoborneol biosynthesis in streptomycetes. Evidence for an isoprenoid pathway and its absence in non-differentiating isolates. *FEBS Lett* 1981;125:220-222.
- 23. Berry DR. Products of primary metabolic pathways. In: Berry DR, ed. *Physiology of industrial fungi*. Oxford: Blackwell Scientific Publications, 1988:130-160.
- 24. Berufsgenossenschaft der chemischen Industrie. 2-Methylpropanol-1. In: *Toxicological* evaluations 15. Potential health hazards of existing chemicals. Berlin, Heidelberg: Springer-Verlag, 1999:1-45.
- 25. Bevan C. Monohydric alcohols C<sub>7</sub> to C<sub>18</sub>, aromatic, and other alcohols. In: Bingham E, Cohrssen B, Powell CH, eds. *Patty's toxicology. Vol. 1*. 5th ed. New York: John Wiley, 2001:461-541.
- 26. Birmingham DJ, ed. Amylvinylcarbinol. Fragrance raw materials monographs. *Food Cosmet Toxicol* 1976;14:681.
- 27. Bjurman J. Release of MVOCs from microorganisms. In: Salthammer T, ed. Organic indoor air pollutants: occurrence, measurement, evaluation. New York: Wiley-VCH, 1999:259-273.
- Bjurman J, Kristensson J. Production of volatile metabolites by the soft rot fungus. *Chaetomium globosum* on building materials and defined media. *Microbios* 1992;72:47-54.
- 29. Bjurman J, Kristensson J. Volatile production by *Aspergillus versicolor* as a possible cause of odor in houses affected by fungi. *Mycopathol* 1992;118:173-178.
- 30. Bjurman J, Nordstrand E, Kristensson J. Growth-phase-related production of potential volatile organic tracer compounds by moulds on wood. *Indoor Air* 1997;7:2-7.
- Bos PMJ. DEC and SCG basis for an occupational health standard: 2-hexanone. Dutch Expert Committee on Occupational Standards and the Swedish Criteria Group. Arbete och Hälsa 1990:12. Solna, Sweden: National Institute of Occupational Health, 1990.
- 32. Bos PM, de Mik G, Bragt PC. Critical review of the toxicity of methyl n-butyl ketone: risk from occupational exposure. *Am J Ind Med* 1991;20:175-194.
- Braathen OA, Schmidbauer N, Lunder C, Blom P, Mattsson J. ORM (optimal removing of moisture from water damaged building constructions) - MVOCS. In: *Proceedings of Indoor Air 2002, Monterey, CA.*, 2002;1:408-413.
- 34. Burge H. Bioaerosols: prevalence and health effects in the indoor environment. *J Allergy Clin Immunol* 1990;86:687-701.
- 35. Böck R. Sensorische Wirkung von flüchtigen Metaboliten (MVOC) in verschimmelten Innenräumen [Sensory effects of secondary metabolites (MVOC) from moulds]. *Umweltmed Forsch Prax* 2001;6:137-142 (in German, abstract in English).
- Börjesson T, Stöllman U, Adamek P, Kaspersson A. Analysis of volatile compounds for detection of molds in stored cereals. *Cereal Chem* 1989;66:300-304.
- 37. Börjesson T, Stöllman U, Schnürer J. Volatile metabolites and other indicators of Penicillium aurantiogriseum growth on different substrates. *Appl Environ Microbiol* 1990;56:3705-3710.
- Börjesson T, Stöllman U, Schnürer J. Volatile metabolites produced by six fungal species compared with other indicators of fungal growth on cereal grains. *Appl Environ Microbiol* 1992;58:2599-2605.

- 39. Börjesson T, Stöllman U, Schnürer J. Off-odorous compounds produced by molds on oat meal agar: Identification and relation to other growth characteristics. *J Agric Food Chem* 1993;41:2104-2111.
- Carlson N, Quraishi A. Anatomy of a fungal problem. In: Johanning E, ed. Proceedings of the 3rd international conference: Bioaerosols, fungi and mycotoxins: health effects, assessment, prevention and control. Fungal Research Group Foundation, Inc. Albany, NY: Boyd Printing 1999:245-253.
- 41. CAS. CAS Registry File (STN). Chemical Abstracts Service Registry System. Columbus, OH: American Chemical Society, June 2005.
- 42. CCOHS. http://www.ccohs.ca/oshanswers/chemicals/Id50.html (April 4, 2006). Hamilton, ON, Canada: Canadian Centre for Occupational Health and Safety.
- CCRIS. Chemical Carcinogenesis Research Information System. http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?CCRIS (March 2006). Rockville Pike, Bethesda, MD, US: National Library of Medicine.
- 44. Chalier P, Crouzet J. Production of lactones by *Penicillium roqueforti*. *Biotechnol Lett* 1992;14:275-280.
- 45. Chalier P, Crouzet J. Production of volatile components by *Penicillium roqueforti* cultivated in the presence of soya bean oil. *Flavour Fragr J* 1993;8:43-49.
- 46. *ChemFinder database*. http://chemfinder.cambridgesoft.com (April 2005). Cambridge, Massachusetts, USA: CambridgeSoft Corporation.
- 47. *CHEMINFO database*. http://www.ccohs.ca (June 2005) and CD-ROM. Hamilton, Ontario, Canada: Canadian Centre for Occupational Health and Safety.
- Claeson, A-S. Volatile organic compounds from microorganisms identification and health effects. Umeå University Medical Dissertations. Umeå. New Series No. 1052. The National Institue for Working Life, Umeå, 2006: 1-53.
- 49. Claeson AS, Levin JO, Blomquist G, Sunesson AL. Volatile metabolites from microorganisms grown on building materials. In: *Proceedings of Indoor Air 2002, Monterey, CA.*, 2002;1:437-442.
- 50. Claeson AS, Levin JO, Blomquist G, Sunesson AL. Volatile metabolites from microorganisms grown on humid building materials and synthetic media. *J Environ Monit* 2002;4:667-672.
- 51. Cometto-Muñiz JE, Cain WS. Sensory reactions of nasal pungency and odor to volatile organic compounds: the alkylbenzenes. *Am Ind Hyg Assoc J* 1994;55:811-817.
- 52. Cometto-Muñiz JE, Cain WS. Physicochemical determinants and functional properties of the senses of irritation and smell. In: Gammage RB, Berven BA, eds. *Indoor air and human health*. 2nd ed. Boca Raton, FL: CRC Press Inc., 1996:53-65.
- 53. Dainty RH, Edwards RA, Hibbard CM. Volatile compounds associated with the aerobic growth of some *Pseudomonas* species on beef. *J Appl Bacteriol* 1984;57:75-81.
- 54. Dainty RH, Edwards RA, Hibbard CM, Marnewick JJ. Volatile compounds associated with microbial growth on normal and high pH beef stored at chill temperatures. *J Appl Bacteriol* 1989;66:281-289.
- 55. DECOS. Health-based recommended occupational exposure limits for 2-hexanone. Dutch Expert Committee on Occupational Standards in collaboration with the Swedish Criteria Group for Occupational Standards. Voorburg: Department of Social Affairs and Employment-Directorate-General of Labour, Netherlands, 1990.
- 56. DECOS. Health-based recommended occupational exposure limits for 7/8-carbon chain aliphatic monoketones. Dutch Expert Committee on Occupational Standards in collaboration with the Nordic Expert Group for Documentation of Occupational Exosure Limits. Voorburg: Department of Social Affairs and Employment-Directorate-General of Labour, Netherlands, 1990.

- Demyttenaere JC, Moriña RM, De Kimpe N, Sandra P. Use of headspace solid-phase microextraction and headspace sorptive extraction for the detection of the volatile metabolites produced by toxigenic Fusarium species. *J Chromatogr A* 2004;1027:147-154.
- 58. Demyttenaere JC, Moriña RM, Sandra P. Monitoring and fast detection of mycotoxinproducing fungi based on headspace solid-phase microextraction and headspace sorptive extraction of the volatile metabolites. *J Chromatogr A* 2003;985:127-135.
- DFG. 2-Hexanon. In: Toxikologisch-arbeitsmedizinische Begründungen von MAK-Werten (Maximale Arbeitsplatzkonzentrationen). Band IV. Deutsche Forschungsgemeinschaft (German Research Foundation). Weinheim: Wiley-VCH, 1975.
- 60. DFG. *List of MAK and BAT values 2005*. Commission for the Investigation of Health Hazards of Chemical Compounds in the Work Area. Report No. 41. Deutsche Forschungsgemeinschaft (German Research Foundation). Weinheim: Wiley-VCH, 2005.
- 61. Dionigi CP, Ingram DA. Effects of temperature and oxygen concentration on geosmin production by *Streptomyces tendae* and *Penicillium expansum*. J Agric Food Chem 1994;42:143-145.
- 62. Diskin AM, Spanel P, Smith D. Time variation of ammonia, acetone, isoprene and ethanol in breath: a quantitative SIFT-MS study over 30 days. *Physiol Meas* 2003;24:107-119.
- 63. Elke K, Begerow J, Oppermann H, Kramer U, Jermann E, Dunemann L. Determination of selected microbial volatile organic compounds by diffusive sampling and dual-column capillary GC-FID--a new feasible approach for the detection of an exposure to indoor mould fungi? *J Environ Monit* 1999;1:445-452.
- 64. Ezeonu IM, Price DL, Simmons RB, Crow SA, Ahearn DG. Fungal production of volatiles during growth on fiberglass. *Appl Environ Microbiol* 1994;60:4172-4173.
- 65. Falk A, Gullstrand E, Löf A, Wigaeus-Hjelm E. Liquid/air partition coefficients of four terpenes. *Br J Ind Med* 1990;47:62-64.
- 66. Fedoruk MJ, Uhlman S, Baker D, Yang H. Analysis of microbial contamination of a ventilation system detected by measurement of microbial volatile organic compounds. In: Johanning E, eds. Proceedings of the 3rd international conference: Bioaerosols, fungi and mycotoxins: health effects, assessment, prevention and control. Fungal Research Group Foundation, Inc. Albany, NY: Boyd Printing 1999:386-395.
- 67. Fenske JD, Paulson SE. Human breath emissions of VOCs. *J Air Waste Manag Assoc* 1999;49:594-598.
- Fessenden RJ, Fessenden JS. Organic Chemistry. 4th ed. Pacific Grove, CA: Brooks/Cole Publishing Company, 1990:1137.
- 69. Fiedler K, Schutz E, Geh S. Detection of microbial volatile organic compounds (MVOCs) produced by moulds on various materials. *Int J Hyg Environ Health* 2001;204:111-121.
- 70. Fischer G, Dott W. Relevance of airborne fungi and their secondary metabolites for environmental, occupational and indoor hygiene. *Arch Microbiol* 2003;179:75-82.
- Fischer G, Schwalbe R, Moller M, Ostrowski R, Dott W. Species-specific production of microbial volatile organic compounds (MVOC) by airborne fungi from a compost facility. *Chemosphere* 1999;39:795-810.
- 72. Fischer G, Schwalbe R, Ostrowski R, Dott W. Airborne fungi and their secondary metabolites in working places in a compost facility. *Mycoses* 1998;41:383-388.
- 73. Gadd GM. Carbon nutrition and metabolism. In: Berry DR, ed. *Physiology of industrial fungi*. Oxford: Blackwell Scientific Publications, 1988:21-57.
- 74. Gao P, Korley F, Martin J, Chen BT. Determination of unique microbial volatile organic compounds produced by five *Aspergillus* species commonly found in problem buildings. *AIHA J (Fairfax, Va)* 2002;63:135-140.
- 75. Gao P, Martin J. Volatile metabolites produced by three strains of *Stachybotrys chartarum* cultivated on rice and gypsum board. *Appl Occup Environ Hyg* 2002;17:430-436.

- 76. Gustafsson H. Kemisk emission från byggnadsmaterial. Beskrivning av skadefall, mätteknik och åtgärder. [Building materials identified as sources for indoor air pollution]. Borås, Sweden: Swedish National Testing and Research Institute, SP Report 1990;25:65 (in Swedish).
- 77. Harris ND, Karahadian C, Lindsay RC. Musty aroma compounds produced by selected molds and actinomycetes on agar and whole wheat bread. *J Food Prot* 1986;49:964-970.
- Haschek WM, Boyd MR, Hakkinen PJ, Owenby CS, Witschi H. Acute inhalation toxicity of 3-methylfuran in the mouse: pathology, cell kinetics, and respiratory rate effects. *Toxicol Appl Pharmacol* 1984;72:124-133.
- 79. Haschek WM, Morse CC, Boyd MR, Hakkinen PJ, Witschi HP. Pathology of acute inhalation exposure to 3-methylfuran in the rat and hamster. *Exp Mol Pathol* 1983;39:342-354.
- Hau KM, Connell DW. Quantitative structure–activity relationships (QSARs) for odor thresholds of volatile organic compounds (VOCs). *Indoor Air* 1998;8:23-33.
- 81. Hawke JC. The formation and metabolism of methyl ketones and related compounds. *J Dairy Res* 1966;33:225-243.
- 82. Health Council of the Netherlands: Committee on Updating of Occupational Exposure Limits. *3-Methylbutan-1-ol; Health-based reassessment of administrative occupational exposure limits.* The Hague: Health Council of the Netherlands, 2005; 2000/15OSH/085 (R).
- Helmig D, Klinger LF, Guenther A, Vierling L, Geron C, Zimmerman P. Biogenic volatile organic compound emissions (BVOCs). I. Identifications from three continental sites in the U.S. *Chemosphere* 1999;38:2163-2187.
- Herr C, Schenke S, Harpel S, Fischer G, Rethage T, Ulu F, Bergmann A. Stilianakis N, Lindemann H, Eikmann, T. Exposure assessment of microbial volatile organic compounds (MVOC) and organic contaminants in normal bedrooms of children with airway disease. *Epidemiology* 2004;15:68.
- 85. Herr C, Harpel S, zur Nieden A, Stilianakis N, Eikmann TH. Assessing health effects of bioaerosols measuring viable spores and microbial volatile organic compounds (MVOC) in residential air. In: *Proceedings of Indoor Air 2002*, Monterey, CA, 2002:3:29-34.
- 86. Hewitt WR, Brown EM. Nephrotoxic interactions between ketonic solvents and halogenated aliphatic chemicals. *Fundam Appl Toxicol* 1984;4:902-908.
- 87. Horner WE, Morey PR, Black MS. MVOC and VOC emission pattern from multiple strains of indoor fungi. In: *Proceedings of Indoor Air 1999*, Edinburgh, Scotland, 1999:4:915-920.
- HSDB. Hazardous Substances Data Bank. http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB (June 2005). Rockville Pike, Bethesda, MD, US: National Library of Medicine.
- 89. IPCS INCHEM. *International chemical safety cards (ICSCs)*. http://www.inchem.org (June 2005). International Programme on Chemical Safety (IPCS) Inchem and the Commission of the European Communities (CEC).
- 90. IPCS/WHO. *Environmental health criteria 65: Butanols: four isomers*. Geneva: International Programme on Chemical Safety, World Health Organization, 1987:9-42.
- 91. IPCS/WHO. *Environmental health criteria 89: Formaldehyde*. Geneva: International Programme on Chemical Safety, World Health Organization, 1989:1-219.
- Jaakkola JJ, Jaakkola N, Ruotsalainen R. Home dampness and molds as determinants of respiratory symptoms and asthma in pre-school children. *J Expo Anal Environ Epidemiol* 1993;3 (Suppl 1):129-142.
- 93. Jelen H, Wasowicz E. Volatile fungal metabolites and their relation to the spoilage of agricultural commodities. *Food Rev Int* 1998;14:391-426.
- 94. Jelen HH, Majcher M, Zawirska-Wojtasiak R, Wiewiorowska M, Wasowicz E. Determination of geosmin, 2-methylisoborneol, and a musty-earthy odor in wheat grain by SPME-GC-MS, profiling volatiles, and sensory analysis. *J Agric Food Chem* 2003;51:7079-7085.

- 95. Jelen HH, Mirocha CJ, Wasowicz E, Kaminski E. Production of volatile sesquiterpenes by *Fusarium sambucinum* strains with different abilities to synthesize trichothecenes. *Appl Environ Microbiol* 1995;61:3815-3820.
- 96. Johansson P. Moldy odor and geosmin from soil-contaminated construction timber. In: *Proceedings from Healthy Buildings 2000*, Helsinki, Finland, 2000;4:369-374.
- Johnson BL, Anger WK, Setzer JV, Lynch DW, Lewis TR. Neurobehavioral effects of methyl N-butyl ketone and methyl N-amyl ketone in rats and monkeys: a summary of NIOSH investigations. *J Environ Pathol Toxicol* 1979;2:113-133.
- 98. Johnson BL, Setzer JV, Lewis TR, Hornung RW. An electrodiagnostic study of the neurotoxicity of methyl n-amyl ketone. *Am Ind Hyg Assoc J* 1978;39:866-872.
- 99. Kaminski E, Libbey LM, Stawicki S, Wasowicz E. Identification of the predominant volatile compounds produced by *Aspergillus flavus*. *Appl Microbiol* 1972;24:721-726.
- 100. Kaminski E, Stawicki S, Wasowicz E. Volatile flavor compounds produced by molds of *Aspergillus, Penicillium*, and *Fungi imperfecti. Appl Microbiol* 1974;27:1001-1004.
- 101. Kane LE, Barrow CS, Alarie Y. A short-term test to predict acceptable levels of exposure to airborne sensory irritants. *Am Ind Hyg Assoc J* 1979;40:207-229.
- 102. Kangas J, Jappinen P, Savolainen H. Exposure to hydrogen sulfide, mercaptans and sulfur dioxide in pulp industry. *Am Ind Hyg Assoc J* 1984;45:787-790.
- 103. Karlshøj K, Larsen TO. Differentiation of species from the *Penicillium roqueforti* group by volatile metabolite profiling. *J Agric Food Chem* 2005;53:708-715.
- 104. Kasanen JP, Pasanen AL, Pasanen P, Liesivuori J, Kosma VM, Alarie Y. Stereospecificity of the sensory irritation receptor for nonreactive chemicals illustrated by pinene enantiomers. *Arch Toxicol* 1998;72:514-523.
- 105. Katz GV, O'Donoghue JL, DiVincenzo GD, Terhaar CJ. Comparative neurotoxicity and metabolism of ethyl n-butyl ketone and methyl n-butyl ketone in rats. *Toxicol Appl Pharmacol* 1980;52:153-158.
- 106. Keller R, Senkpiel K, Ohgke H. Use of MVOC measurements and odour perception as indicator of mould in indoor areas. In: Johanning E, eds. Proceedings of the 3rd international conference: Bioaerosols, fungi and mycotoxins: health effects, assessment, prevention and control. Fungal Research Group Foundation, Inc. Albany, NY: Boyd Printing 1999:532-537.
- 107. Kier LD, Brusick DJ, Auletta AE, Von Halle ES, Brown MM, Simmon VF, Dunkel V, McCann J, Mortelmans K, Prival M, Rao T, Ray V. The Salmonella typhimurium/mammalian microsomal assay. A report of the U.S. Environmental Protection Agency Gene-Tox Program. *Mutat Res* 1986;168:69-240.
- 108. Kim HY, Lee SB, Chung YH, Lim CH, Yu IJ, Park SC, Shin JY, Kim SH, Shin DH, Kim JC. Evaluation of subchronic inhalation toxicity of dimethyl disulfide in rats. *Inhal Toxicol* 2006;18:395-403.
- Klimisch HJ, Hellwig J. Studies on the prenatal toxicity of 3-methyl-1-butanol and 2-methyl-1-propanol in rats and rabbits following inhalation exposure. *Fundam Appl Toxicol* 1995;27:77-89.
- 110. Klingberg J, Beviz A, Ohlson CG, Tenhunen R. Disturbed iron metabolism among workers exposed to organic sulfides in a pulp plant. *Scand J Work Environ Health* 1988;14:17-20.
- 111. Knasko SC. Human responses to ambient olfactory stimuli. In: Gammage RB, Berven BA, eds. *Indoor air and human health.* 2nd ed. Boca Raton, FL: CRC Press Inc., 1996:107-126.
- 112. Koren HS, Graham DE, Devlin RB. Exposure of humans to a volatile organic mixture. III. Inflammatory response. *Arch Environ Health* 1992;47:39-44.
- 113. Korpi A. *Fungal volatile metabolites and biological responses to fungal exposure*. Kuopio University, Faculty of Natural and Environmental Sciences. Kuopio, Finland: Kuopio University Publications, 2001:1-129 (Doctoral Thesis).

- 114. Korpi A, Kasanen JP, Alarie Y, Kosma VM, Pasanen AL. Sensory irritating potency of some microbial volatile organic compounds (MVOCs) and a mixture of five MVOCs. Arch Environ Health 1999;54:347-352.
- 115. Korpi A, Kasanen JP, Pasanen AL. Sensory irritation of microbially produced volatile organic compounds in mice during repeated exposures. In: Johanning E, eds. *Proceedings of the 3rd international conference: Bioaerosols, fungi and mycotoxins: health effects, assessment, prevention and control.* Fungal Research Group Foundation, Inc. Albany, NY: Boyd Printing 1999:106-111.
- 116. Korpi A, Pasanen AL, Pasanen P. Volatile compounds originating from mixed microbial cultures on building materials under various humidity conditions. *Appl Environ Microbiol* 1998;64:2914-2919.
- 117. Korpi A, Pasanen AL, Pasanen P, Kalliokoski P. Microbial growth and metabolism in house dust. *Int Biodeterior Biodegrad* 1997;40:19-27.
- 118. Korpi A, Pasanen AL, Viitanen H. Volatile metabolites of *Serpula lacrymans, Coniophora puteana, Poria placenta, Stachybotrys chartarum* and *Chaetomium globosum. Build Environ* 1999;34:205-211.
- 119. Kreja L, Seidel HJ. Evaluation of the genotoxic potential of some microbial volatile organic compounds (MVOC) with the comet assay, the micronucleus assay and the HPRT gene mutation assay. *Mutat Res* 2002;513:143-150.
- 120. Kreja L, Seidel HJ. On the cytotoxicity of some microbial volatile organic compounds as studied in the human lung cell line A549. *Chemosphere* 2002;49:105-110.
- 121. Kuske M, Romain AC, Nicolas J. Microbial volatile organic compounds as indicators of fungi. Can an electronic nose detect fungi in indoor environments? *Build Environ* 2005;40:824-831.
- 122. Könemann WH, Pieters MN. Confusion of concepts in mixture toxicology. *Food Chem Toxicol* 1996;34:1025-1031.
- 123. Lappalainen S, Pasanen AL, Pasanen P, Kalliokoski P. Production of fungal volatile organic compounds in bedding materials. *Agric Food Sci Finl* 1997;6:219-227.
- 124. Larsen FO, Clementsen P, Hansen M, Maltbæk N, Ostenfeldt-Larsen T, Nielsen KF, Gravesen S, Skov PS, Norn S. Volatile organic compounds from the indoor mould *Trichoderma viride* cause histamine release from human bronchoalveolar cells. *Inflamm Res* 1998;47 (Suppl 1):S5-6.
- 125. Larsen TO, Frisvad JC. Production of volatiles and presence of mycotoxins in conidia of common *Penicillia* and *Aspergilli*. In: Samson RA, Flannigan B, Flannigan ME, Verhoeff AP, Adan OCG, Hoekstra ES, eds. *Health implications of fungi in indoor environments*. Air Quality Monographs 2. Amsterdam: Elsevier Science, 1994:251-279.
- 126. Larsen TO, Frisvad JC. Characterization of volatile metabolites from 47 *Penicillium taxa*. *Mycol Res* 1995;99:1153-1166.
- 127. Larsen TO, Frisvad JC. Chemosystematics of fungi in genus *Penicillium* based on profiles of volatile metabolites. *Mycol Res* 1995;99:1167-1174.
- 128. Larsen TO, Frisvad JC. Comparison of different methods for collection of volatile chemical markers from fungi. *J Microbiol Methods* 1995;24:135-144.
- 129. Laumbach R, Fiedler N, Weschler C, Gardner C, Laskin D, Fan ZH, Zhang J, Lioy P, Ohman-Strickland P, Kelly-McNeil K, Kipen H. Do MVOCs cause irritation?: Nasal effects of VOCs and VOC oxidation products in controlled human exposures. In: Johanning, ed. *Proceedings of the 5th international conference: Bioaerosols, fungi, bacteria, mycotoxins and human health*. Fungal Research Group Foundation, Inc. Albany, NY: Boyd publishing, 2005:154-161.
- 130. Laussmann D, Eis D, Schleibinger H. [Comparison of mycological and chemical analytical laboratory methods for detecting mold damage in indoor environments]. *Bundesgesund-heitsblatt Gesundheitsforschung Gesundheitsschutz* 2004;47:1078-1094 (in German).

- 131. Lee ML, Smith DL, Freeman LR. High-resolution gas chromatographic profiles of volatile organic compounds produced by microorganisms at refrigerated temperatures. *Appl Environ Microbiol* 1979;37:85-90.
- 132. Li AA, Thake DC, Kaempfe TA, Branch DK, O'Donnell P, Speck FL, Tyler TR, Faber WD, Jasti SL, Ouellette R, Banton MI. Neurotoxicity evaluation of rats after subchronic inhalation exposure to isobutanol. *Neurotoxicology* 1999;20:889-900.
- 133. Li HF, Imai T, Ukita M, Sekine M, Higuchi T. Compost stability assessment using a secondary metabolite: geosmin. *Environ Technol* 2004;25:1305-1312.
- 134. Lindecrona RH, Molck AM, Gry J, Poulsen M, Andersen R, Thorup I. Subchronic oral toxicity study on the three flavouring substances: octan-3-ol, 2-methylcrotonic acid and oct-3-yl 2-methylcrotonate in Wistar rats. *Food Chem Toxicol* 2003;41:647-654.
- Lorenz W, Diederich T, Conrad M. Practical experiences with MVOC as an indicator for microbial growth. In: *Proceedings of Indoor Air 2002*, Monterey, CA., 2002:4:341-346.
- 136. Lundberg P, ed. Swedish Criteria Group for Occupational Standards. Consensus report for 2-butanol, isobutanol and tert-butanol. In: *Scientific basis for Swedish occupational standards* V. Arbete och Hälsa 1984;44:98-105. Solna, Sweden: National Board of Occupational Safety and Health.
- 137. Lundberg P, ed. Swedish Criteria Group for Occupational Standards. Consensus report for dimethyl sulfide and dimethyl disulfide. In: *Scientific basis for Swedish occupational standards VIII*. Arbete och Hälsa 1987;39:12-18, Solna, Sweden: National Institute of Occupational Health.
- 138. Lundberg P, ed. Swedish Criteria Group for Occupational Standards. Concensus report for 2-hexanone. In: Scientific basis for Swedish occupational standards. XII. Arbete och Hälsa 1992;6:1-6, Solna, Sweden: National Institute of Occupational Health.
- 139. Lundberg P, ed. Swedish Criteria Group for Occupational Standards. Consensus report for some aliphatic monoketones. In: *Scientific basis for Swedish occupational standards XII*. Arbete och Hälsa 1992;6:7-12, Solna, Sweden: National Institute of Occupational Health.
- 140. Lynch DW, Lewis TR, Moorman WJ, Plotnick HB, Schuler RL, Smallwood AW, Kommineni C. Inhalation toxicity of methyl n-amyl ketone (2-heptanone) in rats and monkeys. *Toxicol Appl Pharmacol* 1981;58:341-352.
- 141. Mathews JM, Raymer JH, Etheridge AS, Velez GR, Bucher JR. Do endogenous volatile organic chemicals measured in breath reflect and maintain CYP2E1 levels *in vivo? Toxicol Appl Pharmacol* 1997;146:255-260.
- Matsuoka A, Hayashi M, Sofuni T. *In vitro* clastogenicity of 19 organic chemicals found in contaminated water and 7 structurally related chemicals. *Environ Mutagen Res* 1998;20:159-165.
- 143. Mayer VW, Goin CJ. Induction of chromosome loss in yeast by combined treatment with neurotoxic hexacarbons and monoketones. *Mutat Res* 1994;341:83-91.
- 144. Mehrer A, Lorenz W. Potential influences on MVOC measurements. In: *Proceedings of Indoor Air 2005*, Bejing, China, 2005:2:2444-2449.
- Menetrez M, Foarde K. Microbial volatile organic compound emission rates and exposure model. *Indoor Built Environ* 2002;11:208-213.
- 146. Meruva NK, Penn JM, Farthing DE. Rapid identification of microbial VOCs from tobacco molds using closed-loop stripping and gas chromatography/time-of-flight mass spectrometry. *J Ind Microbiol Biotechnol* 2004;31:482-488.
- 147. Miller A, III, Scanlan RA, Lee JS, Libbey LM. Volatile compounds produced in sterile fish muscle (*Sebastes melanops*) by *Pseudomonas putrefaciens*, *Pseudomonas fluorescens*, and an *Achromobacter* species. *Appl Microbiol* 1973;26:18-21.
- 148. Morey P, Horner E, Gareis M, Johanning E, Worthan A, Lstiburek J, Krell R. Microbial evaluation in a partially remediated residence. In: *Proceedings of Healthy Buildings 2000*, SYI Indoor Air Information Oy, Helsinki, Finland, 2000;3:385-390.

- 149. Morey P, Worthan A, Weber A, Horner E, Black M, Muller W. Microbial VOCs in moisture damaged buildings. In: *Proceedings from IAQ*'97, Bethesda, Maryland 1997;1:245-250.
- 150. Morse CC, Boyd MR, Witschi H. The effect of 3-methylfuran inhalation exposure on the rat nasal cavity. *Toxicology* 1984;30:195-204.
- 151. MSDS. *Material Safety Data Sheet database*. http://physchem.ox.ac.uk/MSDS (June 2005). Oxford, UK: The Physical and Theoretical Chemistry Laboratory.
- 152. Muller T, Thissen R, Braun S, Dott W, Fischer G. (M)VOC and composting facilities. Part 1: (M)VOC emissions from municipal biowaste and plant refuse. *Environ Sci Pollut Res Int* 2004;11:91-97.
- 153. Muller T, Thissen R, Braun S, Dott W, Fischer G. (M)VOC and composting facilities. Part 2: (M)VOC dispersal in the environment. *Environ Sci Pollut Res Int* 2004;11:152-157.
- 154. Nakajima D, Ishii R, Kageyama S, Onji Y, Mineki S, Morooka N, Takatori K, Goto S. Genotoxicity of microbial volatile organic compounds. *Journal of Health Science* 2006;52:148-153.
- 155. Nakamura SI, Oda Y, Shimada T, Oki I, Sugimoto K. SOS-inducing activity of chemical carcinogens and mutagens in *Salmonella typhimurium* TA1535/pSK1002: examination with 151 chemicals. *Mutat Res* 1987;192:239-246.
- 156. Nielsen GD, Alarie Y, Poulsen OM, Nexø BA. Possible mechanisms for the respiratory tract effects of noncarcinogenic indoor-climate pollutants and bases for their risk assessment. *Scand J Work Environ Health* 1995;21:165-178.
- 157. Nielsen GD, Hansen LF, Alarie Y. Irritation of the upper airways. Mechanisms and structureactivity relationships. In: Knöppel H, Wolkoff P, eds. *Chemical, microbiological, health and comfort aspects of indoor air quality - State of the art in SBS*. Dordrecht: Kluwer Academic Publishers, 1992:99-114.
- 158. Nielsen GD, Larsen ST, Hougaard KS, Hammer M, Wolkoff P, Clausen PA, Wilkins CK, Alarie Y. Mechanisms of acute inhalation effects of (+) and (-)-alpha-pinene in BALB/c mice. *Basic Clin Pharmacol Toxicol* 2005;96:420-488.
- 159. Nikunen E, Leinonen R, Kemiläinen B, Kultamaa A. *Environmental properties of chemicals*. Vol 1. Finnish Environment Institute. Helsinki, Edita Ltd., 2000.
- 160. Nilsson T, Larsen TO, Montarella L, Madsen JØ. Application of headspace solid-phase microextraction for the analysis of volatile metabolites emitted by *Penicillium* species. J Microbiol Methods 1996;25:245-255.
- 161. NIOSH. Criteria for a recommended standard. Occupational exposure to ketones. US National Institute for Occupational Safety and Health. Center for Disease Control. Public Health Service. U.S. Department of Health, Education and Welfare. DHEW (NIOSH) Publication No. 78-173, 1978.
- 162. NIOSH. Documentation for Immediately Dangerous to Life or Health Concentrations (IDLH). http://www.cdc.gov/niosh/idlh/intridl4.htm (January 2007). US National Institute for Occupational Safety and Health.
- 163. NIOSH. Conversion calculator: "mg/m3 to ppm" or "ppm to mg/m3". http://www.cdc.gov/niosh/docs/2004-101/calc.htm (April 30, 2006). US National Institute for Occupational Safety and Health. Centers for Disease Control and Prevention. Department of Health and Human Services.
- 164. Norbäck D, Mi YH, Larsson L, Wady L, Tao J, Mi YL. Moulds, bacteria and MVOC in classroom and outdoor air, and microbial components in settled dust from schools in Shanghai, China. In: *Proceedings of Healthy Buildings 2003*, Singapore, 2003:600-606.
- 165. NTP. Report on carcinogens, Eleventh edition; U.S. Department of Health and Human Services, Public Health Service, National Toxicology Program.
- 166. Pappas GP, Herbert RJ, Henderson W, Koenig J, Stover B, Barnhart S. The respiratory effects of volatile organic compounds. *Int J Occup Environ Health* 2000;6:1-8.

- 167. Pasanen AL, Heinonen-Tanski H, Kalliokoski P, Jantunen MJ. Fungal microcolonies on indoor surfaces an explanation for the base-level fungal spore counts in indoor air. *Atmos Environ* 1992;26B:121-124.
- 168. Pasanen AL, Korpi A, Kasanen JP, Pasanen P. Critical aspects on the significance of microbial volatile metabolites as indoor air pollutants. *Environ Int* 1998;24:703-712.
- 169. Pasanen AL, Lappalainen S, Pasanen P. Volatile organic metabolites associated with some toxic fungi end their mycotoxins. *Analyst* 1996;121:1949-1953.
- 170. Peters MA, Hudson PM, Dixon RL. The effect totigestational exposure to methyl n-butyl ketone has on postnatal development and behavior. *Ecotoxicol Environ Saf* 1981;5:291-306.
- 171. Phillips M, Herrera J, Krishnan S, Zain M, Greenberg J, Cataneo RN. Variation in volatile organic compounds in the breath of normal humans. *J Chromatogr B Biomed Sci Appl* 1999;729:75-88.
- 172. Poulin P, Krishnan K. An algorithm for predicting tissue: blood partition coefficients of organic chemicals from n-octanol: water partition coefficient data. *J Toxicol Environ Health* 1995;46:117-129.
- 173. Putus T. Health effects of moisture damage associated microbes. In: Johanning, ed. Proceedings of the 5th international conference: Bioaerosols, fungi, bacteria, mycotoxins and human health. Fungal Research Group Foundation, Inc. Albany, NY: Boyd publishing, 2005:94-107.
- 174. Ravindranath V, Burka LT, Boyd MR. Reactive metabolites from the bioactivation of toxic methylfurans. *Science* 1984;224:884-886.
- 175. Rivers JC, Pleil JD, Wiener RW. Detection and characterization of volatile organic compounds produced by indoor air bacteria. *J Expo Anal Environ Epidemiol* 1992; (Suppl 1):177-188.
- 176. RTECS. *The Registry of Toxic Effects of Chemical Substances database* (October, 2005). Compiled by the US National Institute for Occupational Safety and Health. Licensed through MDL Information Services, Inc. San Leandro, CA, US: MDL Information Services, Inc.
- 177. Ruotsalainen R, Jaakkola N, Jaakkola JJ. Dampness and molds in day-care centers as an occupational health problem. *Int Arch Occup Environ Health* 1995;66:369-374.
- 178. Ruth JH. Odor thresholds and irritation levels of several chemical substances: a review. *Am Ind Hyg Assoc J* 1986;47:A142-151.
- 179. Sauer F, Schäfer C, Neeb P, Horie O, Moortgat GK. Formation of hydrogen peroxide in the ozonolysis of isoprene and simple alkenes under humid conditions. *Atmos Environ* 1999;33:229-241.
- 180. Schaper M. Development of a database for sensory irritants and its use in establishing occupational exposure limits. *Am Ind Hyg Assoc J* 1993;54:488-544.
- Schilling K, Kayser M, Deckardt K, Kuttler K, Klimisch HJ. Subchronic toxicity studies of 3-methyl-1-butanol and 2-methyl-1-propanol in rats. *Hum Exp Toxicol* 1997;16:722-726.
- 182. Schleibinger H, Brattig C, Mangler M, Laußmann D, Eis D, Braun P, Marchl D, Nickelmann A, Rueden H. Are microbial volatile organic compounds (MVOC) useful predictors for a hidden mould damage? In: *Proceedings of Healthy Buildings 2003*, Singapore, 2003:706-710.
- 183. Schleibinger H, Brattig C, Mangler M, Samwer H, Laußmann D, Eis D, Braun P, Marchl D, Nickelmann A, Rueden H. Microbial volatile organic compounds (MVOC) as indicators for fungal damage. In: *Proceedings of Indoor Air 2002*, Monterey, CA, 2002;4:707-712.
- 184. Schleibinger H, Laußmann D, Brattig C, Mangler M, Eis D, Ruden H. Emission patterns and emission rates of MVOC and the possibility for predicting hidden mold damage? *Indoor Air* 2005;15 (Suppl 9):98-104.
- 185. Schleibinger H, Laußmann D, Samwer H, Eis D, Rüden H. Sind MVOC geeignete Indikatoren für einen verdeckten Schimmelpilzbefall? [Are MVOC useful predictors for a hidden mould damage? *Umweltmed Forsch Prax* 2004;9:151-162 (in German, English abstract).

- Schleibinger H, Rüden H. Air filters from HVAC systems as possible source of volatile organic compounds (VOC) - laboratory and field assays. *Atmos Environ* 1999;33:4571-4577.
- 187. Seifert RM, King AD, Jr. Identification of some volatile constituents of *Aspergillus clavatus*. *J Agric Food Chem* 1982;30:786-790.
- 188. Shelanski, MV. Ethyl amyl ketone. Fragrance raw materials monographs. *Food Cosmet Toxicol* 1973;12:715.
- 189. Sigsgaard T, Bornehag CG. What do we know about dampness in buildings and health? Conclusions from two multidisciplinary reviews of the entire literature on dampness in buildings and associated health effect. In: Johanning, ed. *Proceedings of the 5th international conference: Bioaerosols, fungi, bacteria, mycotoxins and human health.* Fungal Research Group Foundation, Inc. Albany, NY: Boyd Publishing, 2005:24-31.
- 190. Sipes IG, Gandolfi AJ. Biotransformation of toxicants. In: Amdur MO, Doull J, Klaasen CD, eds. Casarett and Doull's Toxicology. The basic science of poisons. 4th ed. New York: Pergamon Press, 1991:88-126.
- 191. Slaughter JC. Nitrogen metabolism. In: Berry DR, ed. *Physiology of industrial fungi*. Oxford: Blackwell Scientific Publications, 1988:58-76.
- 192. Smedje G, Norbäck D, Wessén B, Edling C. Asthma among school employees in relation to the school environment. In: *Proceedings of Indoor Air 1996*, Nagoya, Japan, 1996;1:611-616.
- 193. Smith D, Spanel P, Davies S. Trace gases in breath of healthy volunteers when fasting and after a protein-calorie meal: a preliminary study. *J Appl Physiol* 1999;87:1584-1588.
- 194. Smotherman WP, Robinson SR. Dimethyl disulfide mimics the effects of milk on fetal behavior and responsiveness to cutaneous stimuli. *Physiol Behav* 1992;52:761-765.
- 195. Sprecher E, Hanssen HP. Influence of strain specificity and culture conditions on terpene production by fungi. *J Med Plant Res* 1982;44:41-43.
- 196. SRC. *Interactive LogKow (KowWin) Demo*. http://www.syrres.com/esc/est\_kowdemo.htm (July 2005). Syracuse, NY, US: Syracuse research corporation.
- 197. Ström G, West J, Wessén B, Palmgren U. Quantitative analysis of microbial volatiles in damp Swedish houses. In: Samson RA, Flannigan B, Flannigan ME, Verhoeff AP, Adan OCG, Hoekstra ES, eds. *Health implications of fungi in indoor environments*. Air Quality Monographs 2. Amsterdam: Elsevier Science, 1994;2:291-305.
- 198. Sunesson AL. Volatile metabolites from microorganisms in indoor environments sampling, analysis and identification. Umeå University and National Institute for Working Life. Umeå, Sweden: National Institute for Working Life, 1995:1-88 (Doctoral Thesis).
- 199. Sunesson AL, Nilsson CA, Andersson B. Evaluation of adsorbents for sampling and quantitative analysis of microbial volatiles using thermal desorption-gas chromatography. *J Chromatogr A* 1995;699:203-214.
- 200. Sunesson AL, Nilsson CA, Andersson B, Blomquist G. Volatile metabolites produced by two fungal species cultivated on building materials. *Ann Occup Hyg* 1996;40:397-410.
- 201. Sunesson AL, Nilsson CA, Carlson R, Blomquist G, Andersson B. Production of volatile metabolites from *Streptomyces albidoflavus* cultivated on gypsum board and tryptone glucose extract agar - influence of temperature, oxygen and carbon dioxide levels. *Ann Occup Hyg* 1997;41:393-413.
- 202. Sunesson AL, Vaes WHJ, Nilsson CA, Blomquist G, Andersson B, Carlson F. Identification of volatile metabolites from five fungal species cultivated on two media. *Appl Environ Microbiol* 1995;61:2911-2918.
- 203. Takken W, Knols BG. Odor-mediated behavior of Afrotropical malaria mosquitoes. *Annu Rev Entomol* 1999;44:131-157.
- 204. Tobin RS, Baranowski E, Gilman AP, Kuiper-Goodman T, Miller JD, Giddings M. Significance of fungi in indoor air: report of a working group. *Can J Public Health* 1987;78 (Suppl):S1-32.
- 205. Turner WB. Fungal metabolites. London: Academic Press, 1971:1-446.

- 206. Turner WB, Aldridge DC. Fungal metabolites II. London: Academic Press, 1983:1-631.
- 207. Wady L, Bunte A, Pehrson C, Larsson L. Use of gas chromatography-mass spectrometry/ solid phase microextraction for the identification of MVOCs from moldy building materials. *J Microbiol Methods* 2003;52:325-332.
- Weschler CJ. Ozone in indoor environments: concentration and chemistry. *Indoor Air* 2000;10:269-288.
- Wessén B, Hall T. Directed non-destructive VOC-sampling a method for source location of indoor pollutants. In: *Proceedings of Indoor Air 1999*, Edinburgh, Scotland, 1999;4:420-425.
- 210. Wessén B, Schoeps KO. Microbial volatile organic compounds what substances can be found in sick buildings? *Analyst* 1996;121:1203-1205.
- 211. Wessén B, Schoeps KO. MVOC ratios an aid for remediation of sick buildings. In: *Proceedings of Indoor Air 1996*, Nagoya, Japan, 1996;3:557-561.
- Wessén B, Schoeps KO. Location of emitting sources in buildings with IAQ problems. In: *Proceedings of Healthy Buildings 2000*, Helsinki, Finland, 2000;1:591-595.
- 213. Wessén B, Ström G, Palmgren U. Microbial problem buildings analysis and verification. In: *Proceedings of Indoor Air 1999*, Edinburgh, Scotland, 1999;4:875-879.
- 214. Wessén B, Ström G, Schoeps KO. MVOC profiles a tool for indoor-air quality assessment. In: Morawska L, Bofinger ND, Maroni M, eds. *Proceedings of the international workshop Indoor Air - An Integrated Approach*, Gold Coast Australia, 1994. Oxford: Elsevier Science & Technology Books, 1995:67-70.
- 215. Wheatley RE, Hackett C, Bruce A, Kundzewicz A. Effect of substrate composition on the production of volatile organic compounds from *Trichoderma spp*. Inhibitory to wood decay fungi. *Int Biodeterior Biodegrad* 1997;39:199-205.
- 216. Whillans FD, Lamont GS. Fungal volatile metabolites released into indoor air environments: variation with fungal species and growth media. In: Morawska L, Bofinger ND, Maroni M, eds. *Proceedings of the international workshop Indoor Air An Integrated Approach*, Gold Coast Australia, 1994. Oxford: Elsevier Science & Technology Books, 1995:47-50.
- Whillans FD, Lamont GS. Volatile metabolites from fungi: adsorbent choice and breakthrough, and quantitation. In: *Proceedings of Indoor Air 1996*, Nagoya, Japan, 1996;3:215-220.
- 218. Wibowo AAE. DEC and NEG basis for an occupational health standard. 7/8-Carbon chain aliphatic monoketones (2-heptanone, 3-heptanone, ethylamylketone and methylisoamyl-ketone). Arbete och Hälsa 1990;2:1-44. Solna, Sweden: National Institute of Occupational Health.
- 219. Wilkins CK. Microbial VOC (MVOC) in buildings, their properties and potential use. In: *Proceedings of Indoor Air 2002*, Monterey, CA., 2002;1:431-436.
- 220. Wilkins CK, Larsen K. Identification of volatile (micro)biological compounds from household waste and building materials by thermal desorption capillary gas chromatography-mass spectroscopy. *J High Resol Chromatogr* 1995;18:373-377.
- 221. Wilkins CK, Larsen K. Variation of volatile organic compounds patterns of mold species from damp buildings *Chemosphere* 1995;31:3225-3236.
- 222. Wilkins CK, Larsen K, Simkus M. Volatile metabolites from mold growth on building materials and synthetic media. *Chemosphere* 2000;41:437-446.
- 223. Wilkins CK, Larsen K, Simkus M. Volatile metabolites from indoor molds grown on media containing wood constituents. *Environ Sci Pollut Res Int* 2003;10:206-208.
- 224. Wilkins CK, Nielsen E, Wolkoff P. Patterns in volatile organic compounds in dust from moldy buildings. *Indoor Air* 1997;7:128-134.
- 225. Wilkins CK, Nielsen KF, Hoekstra E, Larson K. MVOC analysis for detection of microbial growth in buildings I. Variation of MVOC and mycotoxin patterns in *Aspergillus versicolor*. In: *Proceedings of Indoor Air 1999*, Edinburgh, Scotland, 1999;4:897-901.

- 226. Wilkins CK, Scholl S. Volatile metabolites of some barley storage molds. *Int J Food Microbiol* 1989;8:11-17.
- 227. Witschi HP, Tryka AF, Mauderly JL, Haschek WM, Satterfield LC, Bowles ND, Boyd MR. Long-term effects of repeated exposure to 3-methylfuran in hamsters and mice. *J Toxicol Environ Health* 1985;16:581-592.
- 228. Wolkoff P, Clausen PA, Wilkins CK, Hougaard KS, Nielsen GD. Formation of strong airway irritants in a model mixture of (+)-alpha-pinene/ozone. *Atmos Environ* 1999;33:693-698.
- 229. Wolkoff P, Clausen PA, Wilkins CK, Nielsen GD. Formation of strong airway irritants in terpene/ozone mixtures. *Indoor Air* 2000;10:82-91.
- Wolkoff P, Wilkins CK, Clausen PA, Nielsen GD. Organic compounds in office environments sensory irritation, odor, measurements and the role of reactive chemistry. *Indoor Air* 2006;16:7-19.
- 231. Wurzenberger M, Grosch W. Wurzenberger M, Grosch W. Origin of the oxygen in the products of the enzymatic clevage reaction of linoleic acid to 1-octen-3-ol and 10-oxo-trans-8-decanoic acid in mushrooms (*Psalliota bispora*). *Biochim Biophys Acta* 1984;794:18-24.
- 232. Wålinder R, Ernstgård L, Gullstrand E, Johansson G, Norbäck D, Venge P, Wieslander G. Acute effects of experimental exposure to four volatile compounds associated with waterdamaged buildings and microbial growth. In: *Proceedings of Indoor Air 1999*, Edinburgh, Scotland, 1999;2:606-611.
- 233. Wålinder R, Ernstgård L, Johanson G, Norbäck D, Venge P, Wieslander G. Acute effects of a fungal volatile compound. *Environ Health Perspect* 2005;113:1775-1778.
- 234. Wålinder R, Norbäck D, Johanson G. Pulmonary reactions after exposure to 3-methylfuran vapour, a fungal metabolite. *Int J Tuberc Lung Dis* 1998;2:1037-1039.
- 235. Zeringue HJ, Jr, Bhatnagar D, Cleveland TE. C15H24 volatile compounds unique to aflatoxigenic strains of *Aspergillus flavus*. *Appl Environ Microbiol* 1993;59:2264-2270.

Late additions:

- 236. Claeson A-S, Sunesson A-L. Identification using versatile sampling and analytical methods of volatile compounds from *Streptomyces albidoflavus* grown on four humid building materials and one synthetic medium. *Indoor Air 2005*;15(Suppl 9):41-47.
- 237. Fischer G, Müller T, Schwalbe R, Ostrowski R, Dott W. Exposure to airborne fungi, MVOC and mycotoxins in biowaste-handling facilities. *Int J Hyg Environ Health* 2000;203:97-104.
- 238. Pasanen P, Korpi A, Kalliokoski P, Pasanen A-L. Growth and volatile metabolite production of *Aspergillus versicolor* in house dust. *Environ Int* 1997;23:425-432.
- 239. Wessén B, Nilsson M, Sisell Å. Odor problems in buildings caused by MVOC and biocides. In: Proceedings of Healthy Buildings 2000, Helsinki, Finland, 2000;4:411-415.

## 19. Data bases used in search of literature

In the search for literature the following databases were used:

CAB Abstracts CAS registry file (STN) HSELINE IARC cancer databases ISI Web of science NIOSHTIC PubMed RISKLINE RTECS Toxnet databases (including e.g. HSDB, Toxline, CCRIS, and Genetox)

In addition, literature cited in the thesis of Anne Korpi (113) was used. Major searches were performed during March-April 2005 for MVOCs and December 2005-June 2006 for single substances. A final search in PubMed and ISI on MVOCs (no single substances) was performed 28 November, 2006.

Submitted for publication January 26, 2007.